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UTILITY PATENT APPLICATION TRANSMITTAL

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Attorney Docket No. 7682-052-999 Total Pages 155

First Named Inventor or Application Identifier

Hong Jin

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09/72416

APPLICATION ELEMENTS See MPEP chapter 600 concerning utility patent application contents.

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1. ☒ Fee Transmittal Form [Total Pages 2]
Submit an original, and a duplicate for fee processing

2. ☒ Specification [Total Pages 123]
(preferred arrangement set forth below)

- Descriptive title of the Invention
- Cross Reference to Related Applications
- Statement Regarding Fed sponsored R&D
- Reference to Microfiche Appendix
- Background of the Invention
- Brief Summary of the Invention
- Brief Description of the Drawings (if filed)
- Detailed Description of the Invention (including drawings, if filed)
- Claim(s)
- Abstract of the Disclosure

3. ☒ Drawing(s) (35 USC 113) [Total Sheets 28]

4. ☒ Oath or Declaration [Total Sheets 2]

- a. ☐ Newly executed (original or copy)
- b. ☐ Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 17 completed)
[Note Box 5 below]

i. ☐ DELETION OF INVENTORS(S)

Signed statement attached deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33 (b).

5. ☐ Incorporation By Reference (useable if Box 4b is checked)
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

- 6. ☐ Microfiche Computer Program (Appendix)
- 7. ☐ Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)
 - a. ☐ Computer Readable Copy
 - b. ☐ Paper Copy (identical to computer copy)
 - c. ☐ Statement verifying identity of above copies

ACCOMPANYING APPLICATION PARTS

- 8. ☐ Assignment Papers
- 9. ☐ 37 CFR 3.73(b) Statement ☐ Power of Attorney (when there is an assignee)
- 10. ☐ English Translation Document (if applicable)
- 11. ☐ Information Disclosure ☐ Copies of IDS Statement (IDS)/PTO-1449 Citations
- 12. ☐ Preliminary Amendment
- 13. ☒ Return Receipt Postcard (MPEP 503) (Should be specifically itemized)
- 14. ☐ Small Entity ☐ Statement filed in prior application, Statement(s) Status still proper and desired
- 15. ☐ Certified Copy of Priority Document(s) (if foreign priority is claimed)
- 16. ☐ Other: Copy of Request for Preparation of a Computer Readable Sequence Listing and Statement including a paper copy of the Sequence Listing from 09/375,514, filed August 17, 1999

17. If a CONTINUING APPLICATION, check appropriate box and supply the requisite information:

Continuation ☐ Divisional ☒ Continuation-in-part (CIP) of prior application No: 09/368,076 filed August 3, 1999.

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09/724416
11/28/00

Sir:

This is a request for filing a ☐ continuation-in-part application under 37 CFR § 1.53(b), of pending prior application no. 09/368,076, filed on August 3, 1999.

of Jin Hong, Roderick Tang, Shengqiang Li, Marty Bryant.
(inventor(s) currently of record in prior application)

for RECOMBINANT RSV EXPRESSION SYSTEMS AND VACCINES
(title of invention)

1. ☒ The filing fee is calculated below:

PATENT APPLICATION FEE VALUE

TYPE	NO. FILED	LESS	EXTRA	EXTRA RATE	FEE
Total Claims	35	-20	15	\$18.00 each	\$ 270.00
Independent	14	-3	11	\$80.00 each	\$ 880.00
Basic Fee					\$ 710.00
Multiple Dependency Fee If Applicable (\$270.00)					\$ 0.00
Total					\$ 1,860.00
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2. ☒ Please charge the required fee to Pennie & Edmonds LLP Deposit Account No. 16-1150. A copy of this sheet is enclosed.
3. ☒ Amend the specification by inserting before the first line the following sentence: This is a continuation-in-part of application no. 09/368,076, filed August 3, 1999.

Respectfully submitted, *by: Jacqueline Benn*
Reg No: 43,492

Date November 28, 2000

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RECOMBINANT RSV VIRUS EXPRESSION SYSTEMS AND VACCINES

CROSS REFERENCE TO RELATED APPLICATIONS

5 This application is a continuation-in-part of Application No. 09/368,076, filed August 3, 1999, which is a continuation-in-part of Application Serial No. 09/161,122, filed September 25, 1998, which claims priority benefit under 35 U.S.C. §119(e) of provisional Application Nos. 60/060,153, filed September 26, 1997, 60/084,133, filed May 1, 1998, and 60/089,207, filed June 12, 1998, each of which is incorporated herein by reference in its entirety.

1. INTRODUCTION

10 The present invention relates to recombinant negative strand virus RNA templates which may be used to express heterologous gene products in appropriate host cell systems and/or to construct recombinant viruses that express, package, and/or present the heterologous gene
15 product. The expression products and chimeric viruses may advantageously be used in vaccine formulations. In particular, the present invention relates to methods of generating recombinant respiratory syncytial viruses and the use of these recombinant viruses as expression vectors and vaccines. The invention is described by way of examples in which recombinant respiratory syncytial viral genomes are used to generate infectious viral particles.

2. BACKGROUND OF THE INVENTION

20 A number of DNA viruses have been genetically engineered to direct the expression of heterologous proteins in host cell systems (e.g., vaccinia virus, baculovirus, etc.). Recently, similar advances have been made with positive-strand RNA viruses (e.g., poliovirus). The
25 expression products of these constructs, i.e., the heterologous gene product or the chimeric virus which expresses the heterologous gene product, are thought to be potentially useful in vaccine formulations (either subunit or whole virus vaccines). One drawback to the use of viruses such as vaccinia for constructing recombinant or chimeric viruses for use in vaccines is the lack of variation in its major epitopes. This lack of variability in the viral strains places strict
30 limitations on the repeated use of chimeric vaccinia, in that multiple vaccinations will generate

host-resistance to the strain so that the inoculated virus cannot infect the host. Inoculation of a resistant individual with chimeric vaccinia will, therefore, not induce immune stimulation.

By contrast, negative-strand RNA viruses such as influenza virus and respiratory syncytial virus, demonstrate a wide variability of their major epitopes. Indeed, thousands of variants of influenza have been identified; each strain evolving by antigenic drift. The negative-strand viruses such as influenza and respiratory syncytial virus would be attractive candidates for constructing chimeric viruses for use in vaccines because its genetic variability allows for the construction of a vast repertoire of vaccine formulations which will stimulate immunity without risk of developing a tolerance.

2.1. RESPIRATORY SYNCYTIAL VIRUS

Virus families containing enveloped single-stranded RNA of the negative-sense genome are classified into groups having non-segmented genomes (Paramyxoviridae, Rhabdoviridae) or those having segmented genomes (Orthomyxoviridae, Bunyaviridae and Arenaviridae).

Paramyxoviridae have been classified into three genera: paramyxovirus (sendai virus, parainfluenza viruses types 1-4, mumps, newcastle disease virus); morbillivirus (measles virus, canine distemper virus and rinderpest virus); and pneumovirus (respiratory syncytial virus and bovine respiratory syncytial virus).

Human respiratory syncytial virus (RSV) is the leading cause of severe lower respiratory tract disease in infants and young children and is responsible for considerable morbidity and mortality. Two antigenically diverse RSV subgroups A and B are present in human populations. RSV is also recognized as an important agent of disease in immuno-compromised adults and in the elderly. Due to the incomplete resistance to RSV reinfection induced by natural infection, RSV may infect multiple times during childhood and life. The goal of RSV immunoprophylaxis is to induce sufficient resistance to prevent the serious disease which may be associated with RSV infection. The current strategies for developing RSV vaccines principally revolve around the administration of purified viral antigen or the development of live attenuated RSV for intranasal administration. However, to date there have been no approved vaccines or highly effective antiviral therapy for RSV.

Infection with RSV can range from an unnoticeable infection to severe pneumonia and death. RSV possesses a single-stranded nonsegmented negative-sense RNA genome of 15,221

nucleotides (Collins, 1991, *In* The paramyxoviruses pp. 103-162, D.W. Kingsbury (ed.) Plenum Press, New York). The genome of RSV encodes 10 mRNAs (Collins et al., 1984, J. Virol. 49: 572-578). The genome contains a 44 nucleotide leader sequence at the 3' termini followed by the NS1-NS2-N-P-M-SH-G-F-M2-L and a 155 nucleotide trailer sequence at the 5' termini
5 (Collins. 1991, *supra*). Each gene transcription unit contains a short stretch of conserved gene start (GS) sequence and a gene end (GE) sequences.

The viral genomic RNA is not infectious as naked RNA. The RNA genome of RSV is tightly encapsidated with the major nucleocapsid (N) protein and is associated with the phosphoprotein (P) and the large (L) polymerase subunit. These proteins form the nucleoprotein
10 core, which is recognized as the minimum unit of infectivity (Brown et al., 1967, J. Virol. 1: 368-373). The RSV N, P, and L proteins form the viral RNA dependent RNA transcriptase for transcription and replication of the RSV genome (Yu et al., 1995, J. Virol. 69:2412-2419; Grosfeld et al., 1995, J. Virol. 69:5677-86). Recent studies indicate that the M2 gene products (M2-1 and M2-2) are involved and are required for transcription (Collins et al., 1996, Proc.
15 Natl. Acad. Sci. 93:81-5).

The M protein is expressed as a peripheral membrane protein, whereas the F and G proteins are expressed as integral membrane proteins and are involved in virus attachment and viral entry into cells. The G and F proteins are the major antigens that elicit neutralizing antibodies in vivo (as reviewed in McIntosh and Chanock, 1990 "Respiratory Syncytial Virus"
20 2nd ed. Virology (D. M. Knipe et al., Ed.) Raven Press, Ltd., N.Y.). Antigenic dimorphism between the subgroups of RSV A and B is mainly linked to the G glycoprotein, whereas the F glycoprotein is more closely related between the subgroups.

Despite decades of research, no safe and effective RSV vaccine has been developed for the prevention of severe morbidity and mortality associated with RSV infection. A formalin-
25 inactivated virus vaccine has failed to provide protection against RSV infection and its exacerbated symptoms during subsequent infection by the wild-type virus in infants (Kapikian et al., 1969, Am. J. Epidemiol. 89:405-21; Chin et al., 1969, Am. J. Epidemiol. 89:449-63) Efforts since have focused on developing live attenuated temperature-sensitive mutants by chemical mutagenesis or cold passage of the wild-type RSV (Gharpure et al., 1969, J. Virol. 3:
30 414-21; Crowe et al., 1994, Vaccine 12: 691-9). However, earlier trials yielded discouraging results with these live attenuated temperature sensitive mutants. Virus candidates were either

underattenuated or overattenuated (Kim et al., 1973, *Pediatrics* 52:56-63; Wright et al., 1976, *J. Pediatrics* 88:931-6) and some of the vaccine candidates were genetically unstable which resulted in the loss of the attenuated phenotype (Hodes et al., 1974, *Proc. Soc. Exp. Biol. Med.* 145:1158-64).

5 Attempts have also been made to engineer recombinant vaccinia vectors which express RSV F or G envelope glycoproteins. However, the use of these vectors as vaccines to protect against RSV infection in animal studies has shown inconsistent results (Olmsted et al. 1986, *Proc. Natl. Acad. Sci.* 83:7462-7466; Collins et al., 1990, *Vaccine* 8:164-168).

10 Thus, efforts have turned to engineering recombinant RSV to generate vaccines. For a long time, negative-sense RNA viruses were refractory to study. Only recently has it been possible to recover negative strand RNA viruses using a recombinant reverse genetics approach (U.S. Patent No. 5,166,057 to Palese et al.). Although this method was originally applied to engineer influenza viral genomes (Luytjes et al. 1989, *Cell* 59:1107-1113; Enami et al. 1990, *Proc. Natl. Acad. Sci. USA* 92: 11563-11567), it has been successfully applied to a wide variety
15 of segmented and nonsegmented negative strand RNA viruses, including rabies (Schnell et al. 1994, *EMBO J.* 13: 4195-4203); VSV (Lawson et al., 1995, *Proc. Natl. Acad. Sci USA* 92: 4477-81); measles virus (Radecke et al., 1995, *EMBO J.* 14:5773-84); rinderpest virus (Baron & Barrett, 1997, *J. virol.* 71: 1265-71); human parainfluenza virus (Hoffman & Banerjee, 1997, *J. Virol.* 71:3272-7; Dubin et al., 1997, *Virology* 235:323-32); SV5 (He et al., 1997, *Virology*
20 237:249-60); respiratory syncytial virus (Collins et al. 1991, *Proc. Natl. Acad. Sci. USA* 88: 9663-9667) and Sendai virus (Park et al. 1991, *Proc. Natl. Acad. Sci. USA* 88:5537-5541; Kato et al. 1996, *Genes to Cells* 1:569-579). Although this approach has been used to successfully rescue RSV, a number of groups have reported that RSV is still refractory to study given several properties of RSV which distinguish it from the better characterized paramyxoviruses of the
25 genera Paramyxovirus, Rubulavirus, and Morbillivirus. These differences include a greater number of RNAs, an unusual gene order at the 3' end of the genome, extensive strain-to-strain sequence diversity, several proteins not found in other nonsegmented negative strand RNA viruses and a requirement for the M2 protein (ORF1) to proceed with full processing of full length transcripts and rescue of a full length genome (Collins et al. PCT WO97/12032; Collins,
30 P.L. et al. pp 1313-1357 of volume 1, Fields Virology, et al., Eds. (3rd ed., Raven Press, 1996).

3. SUMMARY OF THE INVENTION

The present invention relates to genetically engineered recombinant RS viruses and viral vectors which contain heterologous genes which for the use as vaccines. In accordance with the present invention, the recombinant RS viral vectors and viruses are engineered to contain
5 heterologous genes, including genes of other viruses, pathogens, cellular genes, tumor antigens, or to encode combinations of genes from different strains of RSV.

Recombinant negative-strand viral RNA templates are described which may be used to transfect transformed cell that express the RNA dependent RNA polymerase and allow for complementation. Alternatively, a plasmid expressing the components of the RNA polymerase
10 from an appropriate promoter can be used to transfect cells to allow for complementation of the negative-strand viral RNA templates. Complementation may also be achieved with the use of a helper virus or wild-type virus to provide the RNA dependent RNA polymerase. The RNA templates are prepared by transcription of appropriate DNA sequences with a DNA-directed RNA polymerase. The resulting RNA templates are of negative-or positive-polarity and contain
15 appropriate terminal sequences which enable the viral RNA-synthesizing apparatus to recognize the template. Bicistronic mRNAs can be constructed to permit internal initiation of translation of viral sequences and allow for the expression of foreign protein coding sequences from the regular terminal initiation site, or vice versa.

As demonstrated by the examples described herein, recombinant RSV genome in the
20 positive-sense or negative-sense orientation is co-transfected with expression vectors encoding the viral nucleocapsid (N) protein, the associated nucleocapsid phosphoprotein (P), the large (L) polymerase subunit protein, with or without the M2/ORF1 protein of RSV to generate infectious viral particles. Plasmids encoding RS virus polypeptides are used as the source of proteins which were able to replicate and transcribe synthetically derived RNPs. The minimum subset
25 of RSV proteins needed for specific replication and expression of the viral RNP was found to be the three polymerase complex proteins (N, P and L). This suggests that the entire M2-1 gene function, supplied by a separate plasmid expressing M2-1, may not be absolutely required for the replication, expression and rescue of infectious RSV.

The expression products and/or chimeric virions obtained may advantageously be
30 utilized in vaccine formulations. In particular, recombinant RSV genetically engineered to demonstrate an attenuated phenotype may be utilized as a live RSV vaccine. In another

embodiment of the invention, recombinant RSV may be engineered to express the antigenic polypeptides of another strain of RSV (e.g., RSV G and F proteins) or another virus (e.g., an immunogenic peptide from gp120 of HIV) to generate a chimeric RSV to serve as a vaccine, that is able to elicit both vertebrate humoral and cell-mediated immune responses. The use of recombinant influenza or recombinant RSV for this purpose is especially attractive since these viruses demonstrate tremendous strain variability allowing for the construction of a vast repertoire of vaccine formulations. The ability to select from thousands of virus variants for constructing chimeric viruses obviates the problem of host resistance encountered when using other viruses such as vaccinia.

The present invention further relates to the attenuation of human respiratory syncytial virus by deletion of viral accessory gene(s) either singly or in combination.

The present invention further relates to the attenuation of human respiratory syncytial virus by mutagenesis of the viral M2-1 gene.

3.1. DEFINITIONS

As used herein, the following terms will have the meanings indicated:

cRNA = anti-genomic RNA

HA = hemagglutinin (envelope glycoprotein)

HIV = human immunodeficiency virus

L = large polymerase subunit

M = matrix protein (lines inside of envelope)

MDCK = Madin Darby canine kidney cells

MDBK = Madin Darby bovine kidney cells

moi = multiplicity of infection

N = nucleocapsid protein

NA = neuraminidase (envelope glycoprotein)

NP = nucleoprotein (associated with RNA and required for polymerase activity)

NS = nonstructural protein (function unknown)

nt = nucleotide

P = nucleocapsid phosphoprotein

PA, PB1, PB2 = RNA-directed RNA polymerase components

RNP = ribonucleoprotein (RNA, PB2, PB1, PA and NP)

rRNP = recombinant RNP

RSV = respiratory syncytial virus

5 vRNA = genomic virus RNA

viral polymerase complex = PA, PB1, PB2 and NP

WSN = influenza A/WSN/33 virus

WSN-HK virus: reassortment virus containing seven genes from WSN virus and
the NA gene from influenza A/HK/8/68 virus

10

4. DESCRIPTION OF THE FIGURES

FIG. 1. Schematic representation of the RSV/CAT construct (pRSVA2CAT) used in
rescue experiments. The approximate 100 nt long leader and 200 nt long trailer regions of RSV
15 were constructed by the controlled annealing of synthetic oligonucleotides containing partial
overlapping complementarity. The overlapping leader oligonucleotides are indicated by the 1L -
5L shown in the construct. The overlapping trailer nucleotides are indicated by the 1T - 9T
shown in the construct. The nucleotide sequences of the leader and trailer DNAs were ligated
into purified CAT gene DNA at the indicate XbaI and PstI sites respectively. This entire
20 construct was then ligated into KpnI/HindIII digested pUC19. The inclusion of a T7 promoter
sequence and a HgaI site flanking the trailer and leader sequences, respectively, allowed in vitro
synthesis of RSV/CAT RNA transcripts containing the precise genomic sequence 3' and 5' ends.

FIG. 2. Thin layer chromatogram (TLC) showing the CAT activity present in 293 cell
extracts following infection and transfection with RNA transcribed from the RSV/CAT
25 construct shown in Figure 11. Confluent monolayers of 293 cells in six-well plates ($\sim 10^6$ cells)
were infected with either RSV A2 or B9320 at an m.o.i. of 0.1-1.0 pfu cell. At 1 hour post
infection cells were transfected with 5-10 μ g of CAT/RSV using the Transfect-Act™ protocol
of Life Technologies. At 24 hours post infection the infected/transfected monolayers were
harvested and processed for subsequence CAT assay according to Current Protocols in
30 Molecular Biology, Vol. 1, Chapter 9.6.2; Gorman, et al., (1982) Mol. Cell. Biol. 2:1044-1051.
Lanes 1, 2, 3 and 4 show the CAT activity present in (1) uninfected 293 cells, transfected with

CAT/RSV-A2 infected 293 cells, co-infected with supernatant from (2) above. The CAT activity observed in each lane was produced from 1/5 of the total cellular extract from 10^6 cells.

FIG. 3. Schematic representation of the RSV strain A2 genome showing the relative positions of the primer pairs used for the synthesis of cDNAs comprising the entire genome.

5 The endonuclease sites used to splice these clones together are indicated; these sites were present in the native RSV sequence and were included in the primers used for cDNA synthesis. Approximately 100 ng of viral genomic RNA was used in RT/PCR reactions for the separate synthesis of each of the seven cDNAs. The primers for the first and second strand cDNA synthesis from the genomic RNA template are also shown. For each cDNA, the primers for the first strand synthesis are nos. 1-7 and the primers for the second strand synthesis are nos. 1'-7'.

FIG. 4. Schematic representation of the RSV subgroup B strain B9320. BamH1 sites were created in the oligonucleotide primers used for RT/PCR in order to clone the G and F genes from the B9320 strain into RSV subgroup A2 antigenomic cDNA (FIG. 4A). A cDNA fragment which contained G and F genes from 4326 nucleotides to 9387 nucleotides of A2 strain was first subcloned into pUC19 (pUCRVH). Bgl II sites were created at positions of 4630 (SH/G intergenic junction) (FIG. 4B) and 7554 (F/M2 intergenic junction) (FIG. 4C). B93260 A-G and -F cDNA inserted into pUCR/H which is deleted of the A-G and F genes. The resulting antigenomic cDNA clone was termed as pRSVB-GF and was used to transfect Hep-2 cells to generate infectious RSVB-GF virus.

20 FIG. 5. Recombinant RSVB-GF virus was characterized by RT/PCR using RSV subgroup B specific primers. RSV subgroup B specific primers in the G region were incubated with aliquots of the recombinant RSV viral genomes and subjected to PCR. The PCR products were analyzed by electrophoresis on a 1% agarose gel and visualized by staining with ethidium bromide. As shown, no DNA product was produced in the RT/PCR reaction using RSV A2 as a template. However, a predicted product of 254 base pairs was seen in RT/PCR of RSVB-GF RNA and PCR control of plasmid pRSV-GF DNA as template, indicating the rescued virus contained G and F genes derived from B9320 virus.

FIG. 6. Identification of chimeric rRSVA2(B-G) by RT/PCR and Northern blot analysis of RNA expression. FIG. 6A. RT/PCT analysis of chimeric rRSV A2(B-G), in comparison with wild-type A2(A2). Virion RNA extracted from rRSVA2(B-G) (lanes 1, 2) and rRSVA2 (lanes 3,4) was reverse transcribed using a primer annealed to (-) sense vRNA in the RSV F

gene in the presence (+) or absence (-) of reverse transcriptase (RT), followed by PCR with a primer pair flanking the B-G insertion site. No DNA was detected in RT/PCR when reverse transcriptase (RT) was absent (lanes 2,4). A cDNA fragment, which is about 1kb bigger than the cDNA derived from A2, was produced from rRSVA(B-G). This longer PCR DNA product was digested by *Stu I* restriction enzyme unique to the inserted B-G gene (lane 5). 100 bp DNA size marker is indicated (M). FIG. 6B. Northern blot analysis of G mRNA expression. Hep-2 cells were infected with RSV B9320, rRSVA2 and chimeric rRSVA2(B-G). At 48 hr postinfection, total cellular RNA was extracted and electrophoresed on a 1.2% agarose gel containing formaldehyde. RNA was transferred to Hybond Nylon membrane and the filter was hybridized with a ³²P-labeled oligonucleotide probe specific for A2-G or specific for B9320-G mRNA. Both A2 G specific and B9320 G specific transcripts were detected in the rRSVA2 (B-G) infected cells. The run-off RNA transcript (G-M2) from rRSV A2 (B-G) infected cells is also indicated.

FIG. 7. Analysis of protein expression by rRSVA2 (B-G). Hep-2 cells were mock-infected (lanes 1, 5), infected with RSV B9320 (lanes 2, 6), rRSVA2 (lanes 3, 7) and rRSV A2 (B-G) (lanes 4, 8). At 14-18 hr postinfection, infected cells were labeled with ³⁵S-promix and polypeptides were immunoprecipitated by goat polyclonal antiserum against RSV A2 strain (lanes 1-5) or by mouse polyclonal antiserum against RSV B9320 strain (lanes 5-8). Immunoprecipitated polypeptides were separated on a 10% polyacrylamide gel. Both RSV A2 specific G protein and RSV B9320 specific G protein were produced in rRSV A2 (B-G) infected cells. The G protein migration is indicated by *. Mobility of the F1 glycoprotein, and N, P, and M is indicated. Molecular sizes are shown on the left in kilodaltons.

FIG. 8. Plaque morphology of rRSV, rRSVC4G, rRSVA2(B-G) and wild-type A2 virus (wt A2). Hep-2 cells were infected with each virus and incubated at 35°C for six days. The cell monolayers were fixed, visualized by immunostaining, and photographed.

FIG. 9. Growth curve of rRSV, rRSVC4G, wild-type A2 RSV (wt A2) and chimeric rRSVA2(B-G). Hep-2 cells were infected with either virus at a moi of 0.5 and the medium was harvested at 24 hr intervals. The titer of each virus was determined in duplicate by plaque assay on Hep-2 cells and visualized by immunostaining.

FIG. 10. RSV L protein charged residue clusters targeted for site-directed mutagenesis. Contiguous charged amino acid residues in clusters were converted to alanines by site-directed

mutagenesis of the RSV L gene using the QuikChange site-directed mutagenesis kit (Stratagene).

FIG. 11. RSV L protein cysteine residues targeted for site-directed mutagenesis. Cysteine residues were converted to alanine-residues by site-directed mutagenesis of the RSV L gene using the QuikChange site-directed mutagenesis kit (Stratagene).

FIG. 12. Identification RSV M2-2 and SH deletion mutants. Deletions in M2-2 were generated by Hind III digestion of pET(S/B) followed by recloning of a remaining Sac I to BamHI fragment into a full-length clone. Deletions in SH were generated by Sac I digestion of pET(A/S) followed by recloning of a remaining Avr II Sac I fragment into a full-length clone.

FIG. 12A. Identification of the recovered rRSV Δ SH and rRSV Δ M2-2 was performed by RT/PCR using primer pairs specific for the SH gene or M2-2 gene, respectively. Fig. 12B rRSV Δ SH Δ M2-2 was also detected by RT/PCR using primer pairs specific for the M2-2 and SH genes. RT/PCR products were run on an ethidium bromide agarose gel and bands were visualized by ultraviolet (UV) light.

FIG. 13. Structure of rA2 Δ M2-2 genome and recovery of rA2 Δ M2-2. (A). Sequences shown is the region of the M2 gene that M2-1 and M2-2 open reading frames overlap. Total of 234 nt that encode the C-terminal 78 amino acids of M2-2 was deleted through the introduced *Hind III* sites (underlined). The N-terminal 12 amino acid residues of the M2-2 open reading frame are maintained as it overlaps with the M2-1 gene. (B). RT/PCR products of rA2 Δ M2-2 and rA2 viral RNA using primers V1948 and V1581 in the presence (+) or absence (-) of reverse transcriptase (RT). The size of the DNA product derived from rA2 or rA2 Δ M2-2 is indicated.

FIG. 14. Viral RNA expression by rA2 Δ M2-2 and rA2. (A). Total RNA was extracted from rA2 or rA2 Δ M2-2 infected Vero cells at 48 hr postinfection, separated by electrophoresis on 1.2% agarose/2.2 M formaldehyde gels and transferred to nylon membranes. Each blot was hybridized with a Dig-labeled riboprobe specific for the M2-2, M2-1, F, SH, G or N gene. The size of the RNA marker is indicated on the left. (B). Hep-2 and Vero cells were infected with rA2 or rA2 Δ M2-2 for 24 hr and total cellular RNA was extracted. RNA Northern blot was hybridized with a ³²P-labeled riboprobe specific to the negative sense F gene to detect viral genomic RNA or a ³²P-labeled riboprobe specific to the positive sense F gene to detect viral antigenomic RNA and F mRNA. The top panel of the Northern blot on the right was taken from

the top portion of the gel shown in the lower panel and was exposed for 1 week to show antigenome. The lower panel of the Northern blot was exposed for 3 hr to show the F mRNA. The genome, antigenome, F mRNA and dicistronic F-M2 RNA are indicated.

FIG. 15. Viral protein expression in rA2ΔM2-2 and rA2 infected cells. (A). Mock-
5 infected, rA2ΔM2-2 and rA2 infected Vero cells were metabolically labeled with ³⁵S-promix (100 μCi/ml) between 14 to 18 hr postinfection. Cell lysates were prepared for immunoprecipitation with goat polyclonal anti-RSV or rabbit polyclonal anti-M2-2 antisera. Immunoprecipitated polypeptides were separated on a 17.5% polyacrylamide gel containing 4 M urea and processed for autoradiography. The positions of each viral protein are indicated on
10 the right and the molecular weight size markers are indicated on the left. (B). Protein synthesis kinetics in Hep-2 and Vero cells by Western blotting. Hep-2 and Vero cells were infected with rA2 or rA2ΔM2-2 and at 10 hr, 24 hr, or 48 hr postinfection, total infected cellular polypeptides were separated on a 17.5% polyacrylamide gel containing 4 M urea. Proteins were transferred to a nylon membrane and the blot probed with polyclonal antisera against M2-1, NS1 or SH as
15 indicated.

FIG. 16. Plaque morphology of rA2ΔM2-2 and rA2. Hep-2 or Vero cells were infected with rA2ΔM2-2 or rA2 under semisolid overlay composed of 1% methylcellulose and 1 X L15 medium containing 2% FBS for 5 days. Virus plaques were visualized by immunostaining with a goat polyclonal anti-RSV antiserum and photographed under microscope.

20 FIG. 17. Growth curves of rA2ΔM2-2 in Hep-2 and Vero cells. Vero cells (A) or Hep-2 cells (B) were infected with rA2ΔM2-2 or rA2 at m.o.i. of 0.5, and aliquots of medium were harvested at 24 hr intervals as indicated. The virus titers were determined by plaque assay in Vero cells. Virus titer at each time point is average of two experiments.

FIG. 18. Northern blot analysis of rA2ΔNS1, rA2ΔNS2 and rA2ΔNS1ΔNS2. Total
25 cellular RNA was extracted from rA2, rA2ΔNS1, rA2ΔNS2 and rA2ΔNS1ΔNS2 infected Vero cells at 24 hr postinfection, separated by electrophoresis on 1.2% agarose/2.2 M formaldehyde gels and transferred to nylon membranes. Each blot was hybridized with a Dig-labeled riboprobe specific for the NS1, NS2, or M2-2 gene as indicated.

FIG. 19. Plaque morphology of deletion mutants. Hep-2 or Vero cells were infected
30 with each deletion mutant as indicated under semisolid overlay composed of 1% methylcellulose and 1 X L15 medium containing 2% FBS for 6 days. Virus plaques were

visualized by immunostaining with a goat polyclonal anti-RSV antiserum and photographed under microscope.

FIG. 20. Growth curves of rA2 Δ NS1 in Vero cells. Vero cells were infected with rA2 Δ NS1 or rA2 at m.o.i. of 0.5, and aliquots of medium were harvested at 24 hr intervals as indicated. The virus titers were determined by plaque assay in Vero cells.

FIG. 21. Growth curves of rA2 Δ NS2 in Vero cells. Vero cells were infected with rA2 Δ NS2 or rA2 at m.o.i. of 0.5, and aliquots of medium were harvested at 24 hr intervals as indicated. The virus titers were determined by plaque assay in Vero cells.

FIG. 22. Growth curves of rA2 Δ SH Δ M2-2 in Vero cells. Vero cells were infected with rA2 Δ SH Δ M2-2 or rA2 at m.o.i. of 0.5, and aliquots of medium were harvested at 24 hr intervals as indicated. The virus titers were determined by plaque assay in Vero cells.

FIG. 23. Northern blot analysis of several deletion mutants. Total cellular RNA was extracted from Vero cells infected with each deletion mutant as indicated at 24 hr postinfection, separated by electrophoresis on 1.2% agarose/2.2 M formaldehyde gels and transferred to nylon membranes. Each blot was hybridized with a Dig-labeled riboprobe specific for the NS1, NS2, SH or M2-2 gene as indicated.

FIG. 24. Growth curves of rA2 Δ NS2 Δ M2-2 in Vero cells. Vero cells were infected with rA2 Δ NS2 Δ M2-2 or rA2 at m.o.i. of 0.5, and aliquots of medium were harvested at 24 hr intervals as indicated. The virus titers were determined by plaque assay in Vero cells.

FIG. 25. Growth curves of rA2 Δ NS1 Δ NS2 in Vero cells. Vero cells were infected with rA2 Δ NS1 Δ NS2 or rA2 at m.o.i. of 0.5, and aliquots of medium were harvested at 24 hr intervals as indicated. The virus titers were determined by plaque assay in Vero cells.

Fig.26. Insertion of the G and F genes of RSV B9320 strain into recombinant A2 strain. The G and F genes of B9320 were amplified by RT/PCR using primers that contained the BamH I restriction enzyme sites. A DNA cassette containing the G and F genes of B9320 was then introduced into the pRSV(R/H) subclone using the introduced Bgl II restriction enzyme sites that flanked the RSV G and F genes of the A2 strain. The cDNA fragment containing the G and F genes of B9320 was subsequently shuffled into the full-length A2 antigenomic cDNA by ligating at the

Xho I and BamH I sites. The gene start signal of the G gene and the gene end signal of the F gene of B9320 are underlined and the restriction enzyme sites used for cloning are indicated.

Fig. 27. Strain specific expression of the chimeric RSV rA-G_BF_B and rA-G_BF_BΔM2-

- 5 2. **A.** Viral RNA expression. Total cellular RNA were extracted from virus infected Vero cells and the Northern blots were hybridized with probes specific to the G or F gene of either subgroup A or subgroup B RSV. The M2-2 gene expression was examined by using a riboprobe specific to the M2-2 open reading frame. **B.** Viral protein expression. The infected Vero cells were labeled with ³⁵S-methionine and
- 10 ³⁵S-cysteine and the cell lysate immunoprecipitated with anti-RSV polyclonal antibody or anti-M2-2 antibody. To detect the G protein expression, the infected cell extracts were subjected to western blotting using subgroup specific monoclonal antibody against the G protein. Both rA-G_BF_B and rA-G_BF_BΔM2-2 expressed the subgroup B specific G and F proteins and retained normal expression of the other
- 15 genes derived from the subgroup A2 backbone. No M2-2 protein was expressed in rA-G_BF_BΔM2-2 infected cells. Lane 1: rA2, lane 2: rA2ΔM2-2, lane 3: B9320, lane 4: rA-G_BF_B, lane 5: rA-G_BF_BΔM2-2.

Fig. 28. Growth kinetics of the chimeric viruses in Hep-2 and Vero cells. Hep-2 or Vero cells were infected with viruses in duplicates at moi of either 0.1 or 0.01. At

20 24 hours intervals, the infected culture supernatants were harvested and virus titers determined by plaque assay in Vero cells.

25

30

5. DESCRIPTION OF THE INVENTION

5 The present invention relates to genetically engineered recombinant RS
viruses and viral vectors which express heterologous genes or mutated RS viral
genes or a combination of viral genes derived from different strains of RS virus.
The invention relates to the construction and use of recombinant negative strand
RS viral RNA templates which may be used with viral RNA-directed RNA
polymerase to express heterologous gene products in appropriate host cells
10 and/or to rescue the heterologous gene in virus particles. The RNA templates of
the present invention may be prepared by transcription of appropriate DNA
sequences using a DNA-directed RNA polymerase such as bacteriophage T7, T3
or Sp6 polymerase. The recombinant RNA templates may be used to transfect
continuous/transfected cell lines that express the RNA-directed RNA
15 polymerase proteins allowing for complementation.

The invention is demonstrated by way of working examples in which
infectious RSV is rescued from cDNA containing the RSV genome in the
genomic or antigenomic sense introduced into cells expressing the N, P, and L
proteins of the RSV polymerase complex. The working examples further
20 demonstrate that expression of M2-1 expression plasmid is not required for
recovery of infectious RSV from cDNA which is contrary to what has been
reported earlier (Collins et al., 1995, Proc. Natl. Acad. Sci. USA 92:11563-7).
Furthermore, the deletion of the M2-ORF2 from recombinant RSV cDNA
results in the rescue of attenuated RSV particles. M2-2-deleted-RSV is an
25 excellent vehicle to generate chimeric RSV encoding heterologous gene
products, these chimeric viral vectors and rescued virus particles have utility as
expression vectors for the expression of heterologous gene products and as live
attenuated RSV vaccines expressing either RSV antigenic polypeptides or
antigenic polypeptides of other viruses.

30 The invention is further demonstrated by way of working examples in
which a cDNA clone which contained the complete genome of RSV, in addition

to a T7 promoter, a hepatitis delta virus ribozyme and a T7 terminator, is used to generate an infectious viral particle when co-transfected with expression vectors encoding the N, P, L proteins of RSV. In addition, the working examples describe RNA transcripts of cloned DNA containing the coding region -- in negative sense orientation -- of the chloramphenicol-acetyl-transferase (CAT) gene or the green fluorescent protein (GFP) gene flanked by the 5' terminal and 3' terminal nucleotides of the RSV genome. The working examples further demonstrate that an RSV promoter mutated to have increased activity resulted in rescue of infectious RSV particles from a full length RSV cDNA with high efficiency. These results demonstrate the successful use of recombinant viral negative strand templates and RSV polymerase with increased activity to rescue RSV. This system is an excellent tool to engineer RSV viruses with defined biological properties, e.g. live-attenuated vaccines against RSV, and to use recombinant RSV as an expression vector for the expression of heterologous gene products.

This invention relates to the construction and use of recombinant negative strand viral RNA templates which may be used with viral RNA-directed RNA polymerase to express heterologous gene products in appropriate host cells, to rescue the heterologous gene in virus particles and/or express mutated or chimeric recombinant negative strand viral RNA templates (see U.S. Patent No. 5,166,057 to Palese *et al.*, incorporated herein by reference in its entirety). In a specific embodiment of the invention, the heterologous gene product is a peptide or protein derived from another strain of the virus or another virus. The RNA templates may be in the positive or negative-sense orientation and are prepared by transcription of appropriate DNA sequences using a DNA-directed RNA polymerase such as bacteriophage T7, T3 or the Sp6 polymerase.

The ability to reconstitute RNP's in vitro allows the design of novel chimeric influenza and RSV viruses which express foreign genes. One way to achieve this goal involves modifying existing viral genes. For example, the G or F gene may be modified to contain foreign sequences, such as the HA gene of influenza in its external domains. Where the heterologous sequence are epitopes

or antigens of pathogens, these chimeric viruses may be used to induce a protective immune response against the disease agent from which these determinants are derived. For example, a chimeric RNA may be constructed in which a coding sequence derived from the gp120 coding region of human immunodeficiency virus was inserted into the coding sequence of RSV, and chimeric virus produced from transfection of this chimeric RNA segment into a host cell infected with wild-type RSV.

In addition to modifying genes coding for surface proteins, genes coding for nonsurface proteins may be altered. The latter genes have been shown to be associated with most of the important cellular immune responses in the RS virus system. Thus, the inclusion of a foreign determinant in the G or F gene of RSV may - following infection - induce an effective cellular immune response against this determinant. Such an approach may be particularly helpful in situations in which protective immunity heavily depends on the induction of cellular immune responses (e.g., malaria, etc.).

The present invention also relates to attenuated recombinant RSV produced by introducing specific mutations in the genome of RSV which results in an amino acid change in an RSV protein, such as a polymerase protein, which results in an attenuated phenotype.

The present invention also further relates to the generation of attenuated recombinant RSV produced by introducing specific deletions of viral accessory gene(s) either singly or in combination. Specifically, the present invention relates to the generation of attenuated recombinant RSV bearing a deletion of either the M2-2, SH, NS1, or NS2 viral accessory gene. Additionally, the present invention specifically relates to the generation of attenuated recombinant RSV bearing a combination deletion of either the M2-2/SH viral accessory genes, the M2-2/NS2 viral accessory genes, the NS1/NS2 viral accessory genes, the NS1/NS2 viral accessory genes, the SH/NS1 viral accessory genes, the SH/NS2 viral accessory genes, or the SH/NS1/NS2 viral accessory genes.

The invention is demonstrated by way of the working examples presented herein in which infectious attenuated RSV is rescued from RSV

cDNA bearing deletions in the M2-2, SH, NS1, or NS2 viral accessory gene(s) either singly or in combination. Such M2-2, SH, NS1, NS2, M2-2/SH, M2-2/NS2, NS1/NS2, SH/NS1, SH/NS2, or SH/NS1/NS2-deleted RSV represent excellent vehicles for the generation of live attenuated RSV vaccines.

5 Additionally, such M2-2, SH, NS1, NS2, M2-2/SH, M2-2/NS2, NS1/NS2, SH/NS1, SH/NS2, or SH/NS1/NS2-deleted RSV represent excellent vehicles for the generation of chimeric RSV encoding heterologous gene products in place of either the M2-2, SH, NS1, NS2, M2-2/SH, M2-2/NS2, NS1/NS2, SH/NS1, SH/NS2, or SH/NS1/NS2 genes. These chimeric RSV-based viral vectors and
10 rescued infectious attenuated viral particles thus have utility as expression vectors for the expression of heterologous gene products and as live attenuated RSV vaccines expressing either RSV antigenic polypeptides or antigenic polypeptides of heterologous viruses.

The present invention further relates to the generation of attenuated
15 recombinant RSV produced by introducing specific mutations into the M2-1 gene. Specifically, the present invention relates to the generation of attenuated recombinant RSV bearing a mutation of the M2-1 gene introduced by one or more techniques, including, without limitation, cysteine scanning mutagenesis and C-terminal truncations of the M2-1 protein.

20 **5.1. CONSTRUCTION OF THE RECOMBINANT RNA TEMPLATES**

Heterologous gene coding sequences flanked by the complement of the viral polymerase binding site/promoter, e.g., the complement of the 3'-RSV
25 termini or the 3'- and 5'- RSV termini may be constructed using techniques known in the art. Heterologous gene coding sequences may also be flanked by the complement of the RSV polymerase binding site/promoter, e.g., the leader and trailer sequence of RSV using techniques known in the art. Recombinant DNA molecules containing these hybrid sequences can be cloned and
30 transcribed by a DNA-directed RNA polymerase, such as bacteriophage T7, T3 or the Sp6 polymerase and the like, to produce the recombinant RNA templates

which possess the appropriate viral sequences that allow for viral polymerase recognition and activity.

In a preferred embodiment of the present invention, the heterologous sequences are derived from the genome of another strain of RSV, e.g., the genome of RSV A strain is engineered to include the nucleotide sequences encoding the antigenic polypeptides G and F of RSV B strain, or fragments thereof. In such an embodiment of the invention, heterologous coding sequences from another strain of RSV can be used to substitute for nucleotide sequences encoding antigenic polypeptides of the starting strain, or be expressed in addition to the antigenic polypeptides of the parent strain, so that a recombinant RSV genome is engineered to express the antigenic polypeptides of one, two or more strains of RSV.

In yet another embodiment of the invention, the heterologous sequences are derived from the genome of any strain of influenza virus. In accordance with the present invention, the heterologous coding sequences of influenza may be inserted within a RSV coding sequence such that a chimeric gene product is expressed which contains the heterologous peptide sequence within the RSV viral protein. In either embodiment, the heterologous sequences derived from the genome of influenza may include, but are not limited to HA, NA, PB1, PB2, PA, NS1 or NS2.

In one specific embodiment of the invention, the heterologous sequences are derived from the genome of human immunodeficiency virus (HIV), preferably human immunodeficiency virus-1 or human immunodeficiency virus-2. In another embodiment of the invention, the heterologous coding sequences may be inserted within an RSV gene coding sequence such that a chimeric gene product is expressed which contains the heterologous peptide sequence within the influenza viral protein. In such an embodiment of the invention, the heterologous sequences may also be derived from the genome of a human immunodeficiency virus, preferably of human immunodeficiency virus-1 or human immunodeficiency virus-2.

In instances whereby the heterologous sequences are HIV-derived, such sequences may include, but are not limited to, sequences derived from the env gene (i.e., sequences encoding all or part of gp160, gp120, and/or gp41), the pol gene (i.e., sequences encoding all or part of reverse transcriptase, endonuclease, protease, and/or integrase), the gag gene (i.e., sequences encoding all or part of p7, p6, p55, p17/18, p24/25) tat, rev, nef, vif, vpu, vpr, and/or vpx.

One approach for constructing these hybrid molecules is to insert the heterologous coding sequence into a DNA complement of a RSV genomic RNA so that the heterologous sequence is flanked by the viral sequences required for viral polymerase activity; i.e., the viral polymerase binding site/promoter, hereinafter referred to as the viral polymerase binding site. In an alternative approach, oligonucleotides encoding the viral polymerase binding site, e.g., the complement of the 3'-terminus or both termini of the virus genomic segments can be ligated to the heterologous coding sequence to construct the hybrid molecule. The placement of a foreign gene or segment of a foreign gene within a target sequence was formerly dictated by the presence of appropriate restriction enzyme sites within the target sequence. However, recent advances in molecular biology have lessened this problem greatly. Restriction enzyme sites can readily be placed anywhere within a target sequence through the use of site-directed mutagenesis (e.g., see, for example, the techniques described by Kunkel, 1985, Proc. Natl. Acad. Sci. U.S.A. 82;488). Variations in polymerase chain reaction (PCR) technology, described infra, also allow for the specific insertion of sequences (i.e., restriction enzyme sites) and allow for the facile construction of hybrid molecules. Alternatively, PCR reactions could be used to prepare recombinant templates without the need of cloning. For example, PCR reactions could be used to prepare double-stranded DNA molecules containing a DNA-directed RNA polymerase promoter (e.g., bacteriophage T3, T7 or Sp6) and the hybrid sequence containing the heterologous gene and the influenza viral polymerase binding site. RNA templates could then be transcribed directly from this recombinant DNA. In yet another embodiment, the recombinant RNA templates may be prepared by ligating RNAs specifying the negative polarity of

the heterologous gene and the viral polymerase binding site using an RNA ligase. Sequence requirements for viral polymerase activity and constructs which may be used in accordance with the invention are described in the subsections below.

5

5.1.1. INSERTION OF THE HETEROLOGOUS GENES

The gene coding for the L protein contains a single open reading frame. The genes coding for M2 contain two open reading frames for ORF1 and 2, respectively. NS1 and NS2 are coded for by two genes, NS1 and NS2. The G and F proteins, coded for by separate genes, are the major surface glycoproteins of the virus. Consequently, these proteins are the major targets for the humoral immune response after infection. Insertion of a foreign gene sequence into any of these coding regions could be accomplished by either an addition of the foreign sequences to be expressed or by a complete replacement of the viral coding region with the foreign gene or by a partial replacement. The heterologous sequences inserted into the RSV genome may be any length up to approximately 5 kilobases. Complete replacement would probably best be accomplished through the use of PCR-directed mutagenesis.

Alternatively, a bicistronic mRNA could be constructed to permit internal initiation of translation of viral sequences and allow for the expression of foreign protein coding sequences from the regular terminal initiation site. Alternatively, a bicistronic mRNA sequence may be constructed wherein the viral sequence is translated from the regular terminal open reading frame, while the foreign sequence is initiated from an internal site. Certain internal ribosome entry site (IRES) sequences may be utilized. The IRES sequences which are chosen should be short enough to not interfere with RS virus packaging limitations. Thus, it is preferable that the IRES chosen for such a bicistronic approach be no more than 500 nucleotides in length, with less than 250 nucleotides being preferred. Further, it is preferable that the IRES utilized not share sequence or structural homology with picornaviral elements. Preferred

IRES elements include, but are not limited to the mammalian BiP IRES and the hepatitis C virus IRES.

5.2. EXPRESSION OF HETEROLOGOUS GENE PRODUCTS USING RECOMBINANT RNA TEMPLATE

The recombinant templates prepared as described above can be used in a variety of ways to express the heterologous gene products in appropriate host cells or to create chimeric viruses that express the heterologous gene products. In one embodiment, the recombinant template can be combined with viral polymerase complex purified *infra*, to produce rRNPs which are infectious. To this end, the recombinant template can be transcribed in the presence of the viral polymerase complex. Alternatively, the recombinant template may be mixed with or transcribed in the presence of viral polymerase complex prepared using recombinant DNA methods (e.g. see Kingsbury et al., 1987, Virology 156:396-403). In yet another embodiment, the recombinant template can be used to transfect appropriate host cells to direct the expression of the heterologous gene product at high levels. Host cell systems which provide for high levels of expression include continuous cell lines that supply viral functions such as cell lines superinfected with RSV, cell lines engineered to complement RSV viral functions, etc.

5.3. PREPARATION OF CHIMERIC NEGATIVE STRAND RNA VIRUS

In order to prepare chimeric virus, reconstituted RNPs containing modified RSV RNAs or RNA coding for foreign proteins may be used to transfect cells which are also infected with a "parent" RSV virus. Alternatively, the reconstituted RNP preparations may be mixed with the RNPs of wild type parent virus and used for transfection directly. Following transfection, the novel viruses may be isolated and their genomes identified through hybridization

analysis. In additional approaches described herein for the production of infectious chimeric virus, rRNPs may be replicated in host cell systems that express the RSV or influenza viral polymerase proteins (e.g., in virus/host cell expression systems; transformed cell lines engineered to express the polymerase proteins, etc.), so that infectious chimeric virus are rescued; in this instance, helper virus need not be utilized since this function is provided by the viral polymerase proteins expressed. In a particularly desirable approach, cells infected with rRNPs engineered for all eight influenza virus segments may result in the production of infectious chimeric virus which contain the desired genotype; thus eliminating the need for a selection system.

Theoretically, one can replace any one of the genes of RSV, or part of any one of the RSV genes, with the foreign sequence. However, a necessary part of this equation is the ability to propagate the defective virus (defective because a normal viral gene product is missing or altered). A number of possible approaches exist to circumvent this problem.

A third approach to propagating the recombinant virus may involve co-cultivation with wild-type virus. This could be done by simply taking recombinant virus and co-infecting cells with this and another wild-type virus (preferably a vaccine strain). The wild-type virus should complement for the defective virus gene product and allow growth of both the wild-type and recombinant virus. This would be an analogous situation to the propagation of defective-interfering particles of influenza virus (Nayak et al., 1983, In: Genetics of Influenza Viruses, P. Palese and D. W. Kingsbury, eds., Springer-Verlag, Vienna, pp. 255-279). In the case of defective-interfering viruses, conditions can be modified such that the majority of the propagated virus is the defective particle rather than the wild-type virus. Therefore this approach may be useful in generating high titer stocks of recombinant virus. However, these stocks would necessarily contain some wild-type virus.

Alternatively, synthetic RNPs may be replicated in cells co-infected with recombinant viruses that express the RS virus polymerase proteins. In fact, this method may be used to rescue recombinant infectious virus in accordance with

the invention. To this end, the RSV virus polymerase proteins may be expressed in any expression vector/host cell system, including, but not limited to, viral expression vectors (e.g., vaccinia virus, adenovirus, baculovirus, etc.) or cell lines that express the polymerase proteins (e.g., see Krystal et al., 1986, Proc. Natl. Acad. Sci. USA 83: 2709-2713).

5.4. GENERATION OF CHIMERIC VIRUSES WITH AN ATTENUATED PHENOTYPE

The methods of present invention may be used to introduce mutations or heterologous sequences to generate chimeric attenuated viruses which have many applications, including analysis of RSV molecular biology, pathogenesis, and growth and infection properties. In accordance with the present invention, mutations or heterologous sequences may be introduced for example into the F or G protein coding sequences, NS1, NS2, M1ORF1, M2ORF2, N, P, or L coding sequences. In yet another embodiment of the present invention, a particular viral gene, or the expression thereof, may be eliminated to generate an attenuated phenotype, e.g., the M ORF may be deleted from the RSV genome to generate a recombinant RSV with an attenuated phenotype. In yet another embodiment, the individual internal genes of human RSV can be replaced by another strains counterpart, or their bovine or murine counterpart. This may include part or all of one or more of the NS1, NS2, N, P, M, SH, M2(ORF1), M2(ORF2) and L genes or the G and F genes. The RSV genome contains ten mRNAs encoding three transmembrane proteins, G protein, fusion F protein required for penetration, and the small SH protein; the nucleocapsid proteins N, P and L; transcription elongation factor M2 ORF 1; the matrix M protein and two nonstructural proteins, NS1 and NS2. Any one of the proteins may be targeted to generate an attenuated phenotype. Other mutations which may be utilized to result in an attenuated phenotype are insertional, deletional and site directed mutations of the leader and trailer sequences.

In accordance with the present invention, an attenuated RSV exhibits a substantially lower degree of virulence as compared to a wild-type virus,

including a slower growth rate, such that the symptoms of viral infection do not occur in an immunized individual.

In accordance with the present invention attenuated recombinant RSV may be generated by incorporating a broad range of mutations including single nucleotide changes, site-specific mutations, insertions, substitutions, deletions, or rearrangements. These mutations may affect a small segment of the RSV genome, e.g., 15 to 30 nucleotides, or large segments of the RSV genome, e.g., 50 to 1000 nucleotides, depending on the nature of the mutation. In yet another embodiment, mutations are introduced upstream or downstream of an existing cis-acting regulatory element in order to ablate its activity, thus resulting in an attenuated phenotype.

In accordance with the invention, a non-coding regulatory region of a virus can be altered to down-regulate any viral gene, e.g. reduce transcription of its mRNA and/or reduce replication of vRNA (viral RNA), so that an attenuated virus is produced.

Alterations of non-coding regulatory regions of the viral genome which result in down-regulation of replication of a viral gene, and/or down-regulation of transcription of a viral gene will result in the production of defective particles in each round of replication; i.e. particles which package less than the full complement of viral segments required for a fully infectious, pathogenic virus. Therefore, the altered virus will demonstrate attenuated characteristics in that the virus will shed more defective particles than wild type particles in each round of replication. However, since the amount of protein synthesized in each round is similar for both wild type virus and the defective particles, such attenuated viruses are capable of inducing a good immune response.

The foregoing approach is equally applicable to both segmented and non-segmented viruses, where the down regulation of transcription of a viral gene will reduce the production of its mRNA and the encoded gene product. Where the viral gene encodes a structural protein, e.g., a capsid, matrix, surface or envelope protein, the number of particles produced during replication will be reduced so that the altered virus demonstrates attenuated characteristics; e.g., a

titer which results in subclinical levels of infection. For example, a decrease in viral capsid expression will reduce the number of nucleocapsids packaged during replication, whereas a decrease in expression of the envelope protein may reduce the number and/or infectivity of progeny virions. Alternatively, a decrease in expression of the viral enzymes required for replication, e.g., the polymerase, replicase, helicase, and the like, should decrease the number of progeny genomes generated during replication. Since the number of infectious particles produced during replication are reduced, the altered viruses demonstrated attenuated characteristics. However, the number of antigenic virus particles produced will be sufficient to induce a vigorous immune response.

An alternative way to engineer attenuated viruses involves the introduction of an alteration, including but not limited to an insertion, deletion or substitution of one or more amino acid residues and/or epitopes into one or more of the viral proteins. This may be readily accomplished by engineering the appropriate alteration into the corresponding viral gene sequence. Any change that alters the activity of the viral protein so that viral replication is modified or reduced may be accomplished in accordance with the invention.

For example, alterations that interfere with but do not completely abolish viral attachment to host cell receptors and ensuing infection can be engineered into viral surface antigens or viral proteases involved in processing to produce an attenuated strain. According to this embodiment, viral surface antigens can be modified to contain insertions, substitution or deletions of one or more amino acids or epitopes that interfere with or reduce the binding affinity of the viral antigen for the host cell receptors. This approach offers an added advantage in that a chimeric virus which expresses a foreign epitope may be produced which also demonstrates attenuated characteristics. Such viruses are ideal candidates for use as live recombinant vaccines. For example, heterologous gene sequences that can be engineered into the chimeric viruses of the invention include, but are not limited to, epitopes of human immunodeficiency virus (HIV) such as gp120; hepatitis B virus surface antigen (HBsAg); the glycoproteins of herpes virus

(e.g., gD, gE); VP1 of poliovirus; and antigenic determinants of nonviral pathogens such as bacteria and parasites, to name but a few.

In this regard, RSV is an ideal system in which to engineer foreign epitopes, because the ability to select from thousands of virus variants for constructing chimeric viruses obviates the problem of host resistance or immune tolerance encountered when using other virus vectors such as vaccinia. In another embodiment, alterations of viral proteases required for processing viral proteins can be engineered to produce attenuation. Alterations which affect enzyme activity and render the enzyme less efficient in processing, should affect viral infectivity, packaging, and/or release to produce an attenuated virus.

In another embodiment, viral enzymes involved in viral replication and transcription of viral genes, e.g., viral polymerases, replicases, helicases, etc. may be altered so that the enzyme is less efficient or active. Reduction in such enzyme activity may result in the production of fewer progeny genomes and/or viral transcripts so that fewer infectious particles are produced during replication.

The alterations engineered into any of the viral enzymes include but are not limited to insertions, deletions and substitutions in the amino acid sequence of the active site of the molecule. For example, the binding site of the enzyme could be altered so that its binding affinity for substrate is reduced, and as a result, the enzyme is less specific and/or efficient. For example, a target of choice is the viral polymerase complex since temperature sensitive mutations exist in all polymerase proteins. Thus, changes introduced into the amino acid positions associated with such temperature sensitivity can be engineered into the viral polymerase gene so that an attenuated strain is produced.

5.4.1. THE RSV L GENE AS A TARGET FOR ATTENUATION

In accordance with the present invention, the RSV L gene is an important target to generate recombinant RSV with an attenuated phenotype. The L gene represents 48% of the entire RSV genome. The present invention encompasses

generating L gene mutants with defined mutations or random mutations in the RSV L gene. Any number of techniques known to those skilled in the art may be used to generate both defined or random mutations into the RSV L gene. Once the mutations have been introduced, the functionality of the L gene cDNA mutants are screened in vitro using a minigenome replication system and the recovered L gene mutants are then further analyzed in vitro and in vivo.

The following strategies are exemplary of the approaches which may be used to generate mutants with an attenuated phenotype. Further, the following strategies as described below have been applied to the L gene only by way of example and may also be applied to any of the other RSV genes.

One approach to generate mutants with an attenuated phenotype utilizes a scanning mutagenesis approach to mutate clusters of charged amino acids to alanines. This approach is particularly effective in targeting functional domains, since the clusters of charged amino acids generally are not found buried within the protein structure. Replacing the charged amino acids with conservative substitutions, such as neutral amino acids, e.g., alanine, should not grossly alter the structure of the protein but rather, should alter the activity of the functional domain of the protein. Thus, disruption of charged clusters should interfere with the ability of that protein to interact with other proteins, thus making the mutated protein's activity thermosensitive which can yield temperature sensitive mutants.

A cluster of charged amino acids may be arbitrarily defined as a stretch of five amino acids in which at least two or more residues are charged residues. In accordance with the scanning mutagenesis approach all of the charged residues in the cluster are mutated to alanines using site-directed mutagenesis. Due to the large size of the RSV L gene, there are many clustered charged residues. Within the L gene, there are at least two clusters of four contiguous charged residues and at least seventeen clusters of three contiguous charged residues. At least two to four of the charged residues in each cluster may be substituted with a neutral amino acid, e.g., alanine.

In yet another approach to generate mutants with an attenuated phenotype utilizes a scanning mutagenesis approach to mutate cysteines to

amino acids, such as glycines or alanines. Such an approach takes advantage of the frequent role of cysteines in intramolecular and intermolecular bond formations, thus by mutating cysteines to another residue, such as a conservative substitution e.g., valine or alanine, or a drastic substitution e.g., aspartic acid, the stability and function of a protein may be altered due to disruption of the protein's tertiary structure. There are approximately thirty-nine cysteine residues present in the RSV L gene.

In yet another approach random mutagenesis of the RSV L gene will cover residues other than charged or cysteines. Since the RSV L gene is very large, such an approach may be accomplished by mutagenizing large cDNA fragments of the L gene by PCR mutagenesis. The functionality of such mutants may be screened by a minigenome replication system and the recovered mutants are then further analyzed in vitro and in vivo.

5.5. VACCINE FORMULATIONS USING THE CHIMERIC VIRUSES

Virtually any heterologous gene sequence may be constructed into the chimeric viruses of the invention for use in vaccines. In a preferred embodiment, the present invention relates to bivalent RSV vaccines which confers protection against RSV-A and RSV-B. To formulate such a vaccine, a chimeric RS virus is used which expresses the antigenic polypeptides of both RSV-A and RSV-B subtypes. In yet another preferred embodiment, the present invention relates to a bivalent vaccine which confers protection against both RSV and influenza. To formulate such a vaccine, a chimeric RS virus is used which expresses the antigenic polypeptides of both RSV and influenza.

Preferably, epitopes that induce a protective immune response to any of a variety of pathogens, or antigens that bind neutralizing antibodies may be expressed by or as part of the chimeric viruses. For example, heterologous gene sequences that can be constructed into the chimeric viruses of the invention for use in vaccines include but are not limited to sequences derived from a human immunodeficiency virus (HIV), preferably type 1 or type 2. In a preferred

embodiment, an immunogenic HIV-derived peptide which may be the source of an antigen may be constructed into a chimeric influenza virus that may then be used to elicit a vertebrate immune response.

Such HIV-derived peptides may include, but are not limited to sequences derived from the env gene (i.e., sequences encoding all or part of gp160, gp120, and/or gp41), the pol gene (i.e., sequences encoding all or part of reverse transcriptase, endonuclease, protease, and/or integrase), the gag gene (i.e., sequences encoding all or part of p7, p6, p55, p17/18, p24/25), tat, rev, nef, vif, vpu, vpr, and/or vpx.

Other heterologous sequences may be derived from hepatitis B virus surface antigen (HBsAg); the glycoproteins of herpes virus (e.g. gD, gE); VP1 of poliovirus; antigenic determinants of non-viral pathogens such as bacteria and parasites, to name but a few. In another embodiment, all or portions of immunoglobulin genes may be expressed. For example, variable regions of anti-idiotypic immunoglobulins that mimic such epitopes may be constructed into the chimeric viruses of the invention.

Either a live recombinant viral vaccine or an inactivated recombinant viral vaccine can be formulated. A live vaccine may be preferred because multiplication in the host leads to a prolonged stimulus of similar kind and magnitude to that occurring in natural infections, and therefore, confers substantial, long-lasting immunity. Production of such live recombinant virus vaccine formulations may be accomplished using conventional methods involving propagation of the virus in cell culture or in the allantois of the chick embryo followed by purification.

In this regard, the use of genetically engineered RSV (vectors) for vaccine purposes may require the presence of attenuation characteristics in these strains. Current live influenza virus vaccine candidates for use in humans are either cold adapted, temperature sensitive, or passaged so that they derive several (six) genes from avian influenza viruses, which results in attenuation. The introduction of appropriate mutations (e.g., deletions) into the templates used for transfection may provide the novel viruses with attenuation

characteristics. For example, specific missense mutations which are associated with temperature sensitivity or cold adaption can be made into deletion mutations. These mutations should be more stable than the point mutations associated with cold or temperature-sensitive mutants and reversion frequencies should be extremely low.

Alternatively, chimeric viruses with "suicide" characteristics may be constructed. Such viruses would go through only one or a few rounds of replication in the host. When used as a vaccine, the recombinant virus would go through a single replication cycle and induce a sufficient level of immune response but it would not go further in the human host and cause disease. Recombinant viruses lacking one or more of the essential RS virus genes would not be able to undergo successive rounds of replication. Such defective viruses can be produced by co-transfecting reconstituted RNPs lacking a specific gene(s) into cell lines which permanently express this gene(s). Viruses lacking an essential gene(s) will be replicated in these cell lines but when administered to the human host will not be able to complete a round of replication. Such preparations may transcribe and translate --in this abortive cycle -- a sufficient number of genes to induce an immune response. Alternatively, larger quantities of the strains could be administered, so that these preparations serve as inactivated (killed) virus vaccines. For inactivated vaccines, it is preferred that the heterologous gene product be expressed as a viral component, so that the gene product is associated with the virion. The advantage of such preparations is that they contain native proteins and do not undergo inactivation by treatment with formalin or other agents used in the manufacturing of killed virus vaccines.

In another embodiment of this aspect of the invention, inactivated vaccine formulations may be prepared using conventional techniques to "kill" the chimeric viruses. Inactivated vaccines are "dead" in the sense that their infectivity has been destroyed. Ideally, the infectivity of the virus is destroyed without affecting its immunogenicity. In order to prepare inactivated vaccines, the chimeric virus may be grown in cell culture or in the allantois of the chick embryo, purified by zonal ultracentrifugation, inactivated by formaldehyde or β -

propiolactone, and pooled. The resulting vaccine is usually inoculated intramuscularly.

Inactivated viruses may be formulated with a suitable adjuvant in order to enhance the immunological response. Such adjuvants may include but are not limited to mineral gels, e.g., aluminum hydroxide; surface active substances such as lysolecithin, pluronic polyols, polyanions; peptides; oil emulsions; and potentially useful human adjuvants such as BCG and Corynebacterium parvum.

Many methods may be used to introduce the vaccine formulations described above, these include but are not limited to oral, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, and intranasal routes. It may be preferable to introduce the chimeric virus vaccine formulation via the natural route of infection of the pathogen for which the vaccine is designed. Where a live chimeric virus vaccine preparation is used, it may be preferable to introduce the formulation via the natural route of infection for influenza virus. The ability of RSV and influenza virus to induce a vigorous secretory and cellular immune response can be used advantageously. For example, infection of the respiratory tract by chimeric RSV or influenza viruses may induce a strong secretory immune response, for example in the urogenital system, with concomitant protection against a particular disease causing agent.

The following sections describe by way of example, and not by limitation, the manipulation of the negative strand RNA viral genomes using RSV as an example to demonstrate the applicability of the methods of the present invention to generate chimeric viruses for the purposes of heterologous gene expression, generating infectious viral particles and attenuated viral particles for the purposes of vaccination.

6. RESCUE OF INFECTIOUS RESPIRATORY SYNCYTIALVIRUSES (RSV) USING RNA DERIVED FROM SPECIFIC RECOMBINANT DNAS

This example describes a process for the rescue of infectious respiratory syncytial virus (RSV), derived from recombinant cDNAs encoding the entire

RSV RNA genome into stable and infectious RSVs, as noted in Section 5 above. The method described may be applied to both segmented and non-segmented RNA viruses, including orthomyxovirus, paramyxovirus, e.g., Sendai virus, parainfluenza virus types 1-4, mumps, newcastle disease virus; morbillivirus, e.g., measles, canine distemper virus, rinderpest virus; pneumovirus, e.g., respiratory syncytial virus; rhabdovirus, e.g., rabies, vesiculovirus, vesicular stomatitis virus; but is described by way of example in terms of RSV. This process can be used in the production of chimeric RSV viruses which can express foreign genes, i.e., genes non-native to RSV, including other viral proteins such as the HIV env protein. Another exemplary way to achieve the production of chimeric RSV involves modifying existing, native RSV genes, as is further described. Accordingly, this example also describes the utility of this process in the directed attenuation of RSV pathogenicity, resulting in production of a vaccine with defined, engineered biological properties for use in humans.

The first step of the rescue process involving the entire RSV RNA genome requires synthesis of a full length copy of the 15 kilobase (Kb) genome of RSV strain A2. This is accomplished by splicing together subgenomic double strand cDNAs (using standard procedures for genetic manipulation) ranging in size from 1 kb-3.5 kb, to form the complete genomic cDNA. Determination of the nucleotide sequence of the genomic cDNA allows identification of errors introduced during the assembly process; errors can be corrected by site directed mutagenesis, or by substitution of the error region with a piece of chemically synthesized double strand DNA. Following assembly, the genomic cDNA is positioned adjacent to a transcriptional promoter (e.g., the T7 promoter) at one end and DNA sequence which allows transcriptional termination at the other end, e.g., a specific endonuclease or a ribozyme, to allow synthesis of a plus or minus sense RNA copy of the complete virus genome *in vitro* or in cultured cells. The leader or trailer sequences may contain additional sequences as desired, such as flanking ribozyme and tandem T7 transcriptional terminators. The ribozyme can be a hepatitis delta virus

ribozyme or a hammerhead ribozyme and functions to yield an exact 3' end free of non-viral nucleotides.

In accordance with this aspect of the invention, mutations, substitutions or deletions can be made to the native RSV genomic sequence which results in an increase in RSV promoter activity. Applicants have demonstrated that even an increase in RSV promoter activity greatly enhances the efficiency of rescue of RSV, allowing for the rescue of infectious RSV particles from a full-length RSV cDNA carrying the mutation. In particular, a point mutation at position 4 of the genome (C to G) results in a several fold increase in promoter activity and the rescue of infectious viral particles from a full-length RSV cDNA clone carrying the mutation.

The rescue process utilizes the interaction of full-length RSV strain A2 genome RNA, which is transcribed from the constructed cDNA, with helper RSV subgroup B virus proteins inside cultured cells. This can be accomplished in a number of ways. For example, full-length virus genomic RNA from RSV strain A2 can be transcribed *in vitro* and transfected into RSV strain B9320 infected cells, such as 293 cells using standard transfection protocols. In addition, *in vitro* transcribed genomic RNA from RSV strain A2 can be transfected into a cell line expressing the essential RSV strain A2 proteins (in the absence of helper virus) from stably integrated virus genes.

Alternatively, *in vitro* transcribed virus genome RNA (RSV strain A2) can also be transfected into cells infected with a heterologous virus (e.g., in particular vaccinia virus) expressing the essential helper RSV strain A2 proteins, specifically the N, P, L and/or M2-ORF1 proteins. In addition the *in vitro* transcribed genomic RNA may be transfected into cells infected with a heterologous virus, for example vaccinia virus, expressing T7 polymerase, which enables expression of helper proteins from transfected plasmid DNAs containing the helper N, P, and L genes.

As an alternative to transfection of *in vitro* transcribed genomic RNA, plasmid DNA containing the entire RSV cDNA construct may be transfected into cells infected with a heterologous virus, for example vaccinia virus,

expressing the essential helper RSV strain A2 proteins and T7 polymerase, thereby enabling transcription of the entire RSV genomic RNA from the plasmid DNA containing the RSV cDNA construct. The vaccinia virus need not however, supply the helper proteins themselves but only the T7 polymerase; then helper proteins may be expressed from transfected plasmids containing the RSV N, P, and L genes, appropriately positioned adjacent to their own T7 promoters.

When replicating virus is providing the helper function during rescue experiments, the B9320 strain of RSV is used, allowing differentiation of progeny rescue directed against RSV B9320. Rescued RSV strain A2 is positively identified by the presence of specific nucleotide 'marker' sequences inserted in the cDNA copy of the RSV genome prior to rescue.

The establishment of a rescue system for native, i.e., 'wild-type' RSV strain A2 allows modifications to be introduced into the cDNA copy of the RSV genome to construct chimeric RSV containing sequences heterologous in some manner to that of native RSV, such that the resulting rescued virus may be attenuated in pathogenicity to provide a safe and efficacious human vaccine as discussed in Section 5.4 above. The genetic alterations required to cause virus attenuation may be gross (e.g., translocation of whole genes and/or regulatory sequences within the virus genome), or minor (e.g., single or multiple nucleotide substitution(s), addition(s) and/or deletion(s) in key regulatory or functional domains within the virus genome), as further described in detail.

In addition to alteration(s) (including alteration resulting from translocation) of the RSV genetic material to provide heterologous sequence, this process permits the insertion of 'foreign' genes (i.e., genes non-native to RSV) or genetic components thereof exhibiting biological function or antigenicity in such a way as to give expression of these genetic elements; in this way the modified, chimeric RSV can act as an expression system for other heterologous proteins or genetic elements, such as ribozymes, anti-sense RNA, specific oligoribonucleotides, with prophylactic or therapeutic potential, or other viral proteins for vaccine purposes.

6.1. RESCUE OF THE LEADER AND TRAILER SEQUENCES OF RSV STRAIN A2 USING RSV STRAIN B9320 AS HELPER VIRUS

6.1.1. VIRUSES AND CELLS

Although RSV strain A2 and RSV strain B9320 were used in this Example, they are exemplary. It is within the skill in the art to use other strains of RSV subgroup A and RSV subgroup B viruses in accordance with the teachings of this Example. Methods which employ such other strains are encompassed by the invention.

RSV strain A2 and RSV strain B9320 were grown in Hep-2 cells and Vero cells respectively, and 293 cells were used as host during transfection/rescue experiments. All three cell lines were obtained from the ATCC (Rockville, Maryland).

6.1.2. CONSTRUCTION AND FUNCTIONAL ANALYSIS OF REPORTER PLASMIDS

Plasmid pRSVA2CAT (Fig. 1) was constructed as described below.

The cDNAs of the 44 nucleotide leader and 155 nucleotide trailer components of RSV strain A2 (see Mink et al., Virology 185:615-624 (1991); Collins et al., Proc. Natl. Acad. Sci. 88:9663-9667 (1991)), the trailer component also including the promoter consensus sequence of bacteriophage T7 polymerase, were separately assembled by controlled annealing of oligonucleotides with partial overlapping complementarity (see Fig. 1). The oligonucleotides used in the annealing were synthesized on an Applied Biosystems DNA synthesizer (Foster City, CA). The separate oligonucleotides and their relative positions in the leader and trailer sequences are indicated in Fig. 1. The oligonucleotides used to construct the leader were:

1. 5'CGA CGC ATA TTA CGC GAA AAA ATG CGT
ACA ACA AAC TTG CAT AAA C

2. 5'CAA AAA AAT GGG GCA AAT AAG AAT TTG
ATA AGT ACC ACT TAA ATT TAA CT
3. 5'CTA GAG TTA AAT TTA AGT GGT ACT
4. 5'TAT CAA ATT CTT ATT TGC CCC ATT TTT TTG
GTT TAT GCA AGT TTG TTG TA
5. 5'CGC ATT TTT TCG CGT AAT ATG CGT CGG TAC

The oligonucleotides used to construct the trailer were:

1. 5'GTA TTC AAT TAT AGT TAT TAA AAA TTA AAA
ATC ATA TAA TTT TTT AAA TA
2. 5'ACT TTT AGT GAA CTA ATC CTA AAG TTA TCA
TTT TAA TCT TGG AGG AAT AA
3. 5'ATT TAA ACC CTA ATC TAA TTG GTT TAT ATG
TGT ATT AAC TAA ATT ACG AG
4. 5'ATA TTA GTT TTT GAC ACT TTT TTT CTC GTT
ATA GTG AGT CGT ATT A
5. 5'AGC TTA ATA CGA CTC ACT ATA ACG A
6. 5'GAA AAA AAG TGT CAA AAA CTA ATA TCT
CGT AAT TTA GTT AAT ACA CAT AT
7. 5'AAA CCA ATT AGA TTA GGG TTT AAA TTT ATT
CCT CCA AGA TTA AAA TGA TA
8. 5'ACT TTA GGA TTA GTT CAC TAA AAG TTA TTT
AAA AAA TTA TAT GAT TTT TA
9. 5'ATT TTT AAT AAC TAT AAT TGA ATA CTG CA

The complete leader and trailer cDNAs were then ligated to the chloramphenicol-acetyl-transferase (CAT) reporter gene XbaI and PstI sites respectively to form a linear - 1 kb RSV/CAT cDNA construct. This cDNA construct was then ligated into the Kpn I and Hind II sites of pUC19. The integrity of the final pRSVA2CAT construct was checked by gel analysis for the size of the Xba I/Pst I and Kpn I/Hind II digestion products. The complete leader and trailer cDNAs were also ligated to the green fluorescent protein (GFP) gene using appropriate restriction enzyme sites to form a linear cDNA

construct. The resulting RSV-GFP-CAT is a bicistronic reporter construct which expresses both CAT and GFP.

In vitro transcription of Hga I linearized pRSVA2CAT with bacteriophage T7 polymerase was performed according to the T7 supplier protocol (Promega Corporation, Madison, Wisconsin). Confluent 293 cells in six-well dishes ($\sim 1 \times 10^6$ cells per well) were infected with RSV strain B9320 at 1 plaque forming units (p.f.u.) per cell and 1 hour later were transfected with 5-10 μ g of the *in vitro* transcribed RNA from the pRSVA2CAT construct. The transfection procedure followed the transfection procedure of Collins et al., Virology 195:252-256 (1993) and employed Transect/ACT™ and Opti-MEM reagents according to the manufacturers specifications (Gibco-BRL, Bethesda, Maryland). At 24 hours post-infection the 293 cells were assayed for CAT activity using a standard protocol (Current Protocols in Molecular Biology, Vol. 1, Chapter 9.6.2; Gorman, et al., 1982) Mol. Cell Biol. 2: 1044-1051). The detection of high levels of CAT activity indicated that *in vitro* transcribed negative sense RNA containing the 'leader' and 'trailer' regions of the RSV A2 strain genome and the CAT gene can be encapsidated, replicated and expressed using proteins supplied by RSV strain B9320 (See Fig. 2). The level of CAT activity observed in these experiments was at least as high as that observed in similar rescue experiments where homologous RSV strain A2 was used as helper virus. The ability of an antigenically distinct subgroup B RSV strain B9320 to support the encapsidation, replication and transcription of a subgroup A RSV strain A2 RNA has to our knowledge hitherto not been formally reported.

6.2. CONSTRUCTION OF A cDNA REPRESENTING THE COMPLETE GENOME OF RSV

To obtain a template for cDNA synthesis, RSV genomic RNA, comprising 15,222 nucleotides, was purified from infected Hep-2 cells according to the method described by Ward et al., J. Gen. Virol. 64:167-1876 (1983). Based on the published nucleotide sequence of RSV, oligonucleotides

construct. The resulting RSV-GFP-CAT is a bicistronic reporter construct which expresses both CAT and GFP.

In vitro transcription of Hga I linearized pRSVA2CAT with bacteriophage T7 polymerase was performed according to the T7 supplier protocol (Promega Corporation, Madison, Wisconsin). Confluent 293 cells in six-well dishes (-1×10^6 cells per well) were infected with RSV strain B9320 at 1 plaque forming units (p.f.u.) per cell and 1 hour later were transfected with 5-10 μ g of the *in vitro* transcribed RNA from the pRSVA2CAT construct. The transfection procedure followed the transfection procedure of Collins et al., Virology 195:252-256 (1993) and employed Transect/ACT™ and Opti-MEM reagents according to the manufacturers specifications (Gibco-BRL, Bethesda, Maryland). At 24 hours post-infection the 293 cells were assayed for CAT activity using a standard protocol (Current Protocols in Molecular Biology, Vol. 1, Chapter 9.6.2; Gorman, et al., 1982) Mol. Cell Biol. 2: 1044-1051). The detection of high levels of CAT activity indicated that *in vitro* transcribed negative sense RNA containing the 'leader' and 'trailer' regions of the RSV A2 strain genome and the CAT gene can be encapsidated, replicated and expressed using proteins supplied by RSV strain B9320 (See Fig. 2). The level of CAT activity observed in these experiments was at least as high as that observed in similar rescue experiments where homologous RSV strain A2 was used as helper virus. The ability of an antigenically distinct subgroup B RSV strain B9320 to support the encapsidation, replication and transcription of a subgroup A RSV strain A2 RNA has to our knowledge hitherto not been formally reported.

6.2. CONSTRUCTION OF A cDNA REPRESENTING THE COMPLETE GENOME OF RSV

To obtain a template for cDNA synthesis, RSV genomic RNA, comprising 15,222 nucleotides, was purified from infected Hep-2 cells according to the method described by Ward et al., J. Gen. Virol. 64:167-1876 (1983). Based on the published nucleotide sequence of RSV, oligonucleotides

were synthesized using an Applied Biosystems DNA synthesizer (Applied Biosystems, Foster City, CA) to act as primers for first and second strand cDNA synthesis from the genomic RNA template. The nucleotide sequences and the relative positions of the cDNA primers and key endonuclease sites within the RSV genome are indicated in Fig. 3. The production of cDNAs from virus genomic RNA was carried out according to the reverse transcription/polymerase chain reaction (RT/PCR) protocol of Perkin Elmer Corporation, Norwalk, Connecticut (see also Wang et al., (1989) Proc. Natl. Acad. Sci. 86:9717-9721); the amplified cDNAs were purified by electroelution of the appropriate DNA band from agarose gels. Purified DNA was ligated directly into the pCRII plasmid vector (Invitrogen Corp. San Diego), and transformed into either 'One Shot *E. coli* cells (Invitrogen) or 'SURE' *E. coli* cells (Stratagene, San Diego). The resulting, cloned, virus specific, cDNAs were assembled by standard cloning techniques (Sambrook et al., Molecular Cloning - A Laboratory Manual, Cold Spring Harbor laboratory Press (Cold Spring Harbor, NY, 1989) to produce a cDNA spanning the complete RSV genome. The entire cDNA genome was sequenced, and incorrect sequences were replaced by either site-directed mutagenesis or chemically synthesized DNA. Nucleotide substitutions were introduced at bases 7291 and 7294 (with base number 1 being at the start of the genomic RNA 3' end) in the 'F' gene, to produce a novel Stu I endonuclease site, and at positions 7423, 7424, and 7425 (also in the F gene) to produce a novel Pme I site. These changes were designed to act as definitive markers for rescue events. The bacteriophage T7 polymerase and the Hga I endonuclease site were placed at opposite ends of the virus genome cDNA such that either negative or positive sense virus genome RNA can be synthesized *in vitro*. The cDNAs representing the T7 polymerase promoter sequence and the recognition sequence for Hga I were synthesized on an Applied Biosystems DNA synthesizer and were separately ligated to the ends of the virus genome cDNA, or were added as an integral part of PCR primers during amplification of the terminal portion of the genome cDNA, where appropriate; the latter procedure was used when suitable endonuclease sites near the genome cDNA

termini were absent, preventing direct ligation of chemically synthesized T7 promoter/Hga I site cDNA to the genome cDNA. This complete construct (genome cDNA and flanking T7 promoter/Hga I recognition sequence) was then cloned into the Kpn I/Not I sites of the Bluescript II SK phagemid (Stratagene, San Diego) from which the endogenous T7 promoter has been removed by site-directed mutagenesis. RNA transcribed from this complete genome construct may be rescued using RSV subgroup B helper virus to give infectious RSV in accordance with Example 6.1. This basic rescue system for the complete native, i.e., 'wild-type' RSV A2 strain genomic RNA can be employed to introduce a variety of modifications into the cDNA copy of the genome resulting in the introduction of heterologous sequences into the genome. Such changes can be designed to reduce viral pathogenicity without restricting virus replication to a point where rescue becomes impossible or where virus gene expression is insufficient to stimulate adequate immunity.

The following oligonucleotides were used to construct the ribozyme/T7 terminator sequence:

5'GGT*GGCCGGCATGGTCCCAGC
3'CCA CCGGCCGTACCAGGGTCG

CTCGCTGGCGCCGGCTGGGCAACA
GAGCGACCGCGGCCGACCCGTGTG

TTCCGAGGGGACCGTCCCCTCGGT
AAGGCTCCCCTGGCAGGGGAGCCA

AATGGCGAATGGGACGTCGACAGC
TTACCGCTTACCCTGCAGCTGTG

TAACAAAGCCCGAAGGAAGCT
ATTGTTTCGGGCTTCCTTCGA

GAGTTGCTGCTGCCACCGTTG
CTCAACGACGACGGAGGCAAC

5 AGCAATAACTAGATAACCTTGGG
TCGTTATTGATCTATTGGAACCC

CCTCTAAACGGGTCTTGAGGGTCT
GGAGATTTGCCCAGAACTCCCAGA

10 TTTTGCTGAAAGGAGGAACTA
AAAACGACTTTCCTCCTTGAT

TATGCGGCCGCGTCGACGGTA
ATACGCCGGCGEAGCTGCCAT

15 CCGGGCCCGCCTTCGAAG3'
GGCCCGGGCGGAAGCTTC5'

20 A cDNA clone containing the complete genome of RSV a T7 promoter,
a hepatitis delta virus ribozyme and a T7 terminator was generated. This
construct can be used to generate antigenomic RNA or RSV *in vivo* in the
presence of T7 polymerase. Sequence analysis indicated that the plasmid
contained few mutations in RSV genome.

25 6.2.1. MODIFICATIONS OF THE RSV GENOME

Modifications of the RSV RNA genome can comprise gross alterations
of the genetic structure of RSV, such as gene shuffling. For example, the RSV
M2 gene can be translocated to a position closer to the 5' end of the genome, in
30 order to take advantage of the known 3' to 5' gradient in virus gene expression,
resulting in reduced levels of M2 protein expression in infected cells and thereby

reducing the rate of virus assembly and maturation. Other genes and/or regulatory regions may also be translocated appropriately, in some cases from other strains of RSV of human or animal origin. For example, the F gene (and possibly the 'G' gene) of the human subgroup B RSV could be inserted into an otherwise RSV strain A genome (in place of, or in addition to the F and G genes of RSV strain A).

In another approach, the RNA sequence of the RSV viruses N protein can be translocated from its 3' proximal site to a position closer to the 5' end of the genome, again taking advantage of the 3' to 5' gradient in gene transcription to reduce the level of N protein produced. By reducing the level of N protein produced, there would result a concomitant increase in the relative rates of transcription of genes involved in stimulating host immunity to RSV and a concomitant reduction in the relative rate of genome replication. Thus, by translocating the RSV RNA sequence coding for RSV N protein, a chimeric RSV virus having attenuated pathogenicity relative to native RSV will be produced.

Another exemplary translocation modification resulting in the production of attenuated chimeric RSV comprises the translocation of the RSV RNA sequence coding for the L protein of RSV. This sequence of the RS virus is believed responsible for viral polymerase protein production. By translocating the RSV sequence coding for L protein from its native 5' terminal location in the native RSV genome to a location at or near the 3' terminus of the genome, a chimeric RSV virus exhibiting attenuated pathogenicity will be produced. Yet another exemplary translocation comprises the switching the locations of the RSV RNA sequences coding for the RSV G and F proteins (i.e., relative to each other in the genome) to achieve a chimeric RSV having attenuated pathogenicity resulting from the slight modification in the amount of the G and F proteins produced. Such gene shuffling modifications as are exemplified and discussed above are believed to result in a chimeric, modified RSV having attenuated pathogenicity in comparison to the native RSV starting material. The nucleotide sequences for the foregoing encoded proteins are known, as is the nucleotide sequence for the entire RSV genome. See McIntosh, Respiratory Syncytial

Virus in Virology, 2d Ed. edited by B.N. Fields, D.M. Knipe et al., Raven Press, Ltd. New York, 1990 Chapter 38, pp 1045-1073, and references cited therein.

These modifications can additionally or alternatively comprise localized, site specific, single or multiple, nucleotide substitutions, deletions or additions within genes and/or regulatory domains of the RSV genome. Such site specific, single or multiple, substitutions, deletions or additions can reduce virus pathogenicity without overly attenuating it, for example, by reducing the number of lysine or arginine residues at the cleavage site in the F protein to reduce efficiency of its cleavage by host cell protease (which cleavage is believed to be an essential step in functional activation of the F protein), and thereby possibly reduce virulence. Site specific modifications in the 3' or 5' regulatory regions of the RSV genome may also be used to increase transcription at the expense of genome replication. In addition, localized manipulation of domains within the N protein, which is believed to control the switch between transcription and replication can be made to reduce genome replication but still allow high levels of transcription. Further, the cytoplasmic domain(s) of the G and F glycoproteins can be altered in order to reduce their rate of migration through the endoplasmic reticulum and golgi of infected cells, thereby slowing virus maturation. In such cases, it may be sufficient to modify the migration of G protein only, which would then allow additional up-regulation of 'F' production, the main antigen involved in stimulating neutralizing antibody production during RSV infections. Such localized substitutions, deletions or additions within genes and/or regulatory domains of the RSV genome are believed to result in chimeric, modified RSV also having reduced pathogenicity relative to the native RSV genome.

6.3. RESCUE OF A cDNA REPRESENTING THE COMPLETE GENOME OF RSV

6.3.1. THE CONSTRUCTION AND FUNCTIONAL ANALYSIS OF EXPRESSION PLASMIDS

5 The RSV, N, P, and L genes encode the viral polymerase of RSV. The
function of the RSV M genes is unknown. The ability of RSV, N, P, M, and L
expression plasmids to serve the function of helper RSV strain A2 proteins was
assessed as described below. The RSV, N, P, L, and M2-1 genes were cloned
10 into the modified PCITE 2a(+) vector (Novagen, Madison, WI) under the
control of the T7 promoter and flanked by a T7 terminator at its 3' end. PCITE-
2a(+) was modified by insertion of a T7 terminator sequence from PCITE-3a(+)
into the AlwI and Bgl II sites of pCITE-2a(+). The functionality of the N, P,
and L expression plasmids was determined by their ability to replicate the
15 transfected pRSVA2CAT. At approximately 80% confluency, Hep-2 cells in
six-well plates were infected with MVA at a moi of 5. After 1 hour, the infected
cells were transfected with pRSVA₂CAT (0.5mg), and plasmids encoding the N
(0.4mg), P (0.4mg), and L (0.2mg) genes using lipofectACE (Life
Technologies, Gaithersburg, M.D.). The transfection proceeded for 5 hours or
20 overnight and then the transfection medium was replaced with fresh MEM
containing 2% (fetal bovine serum) FBS. Two days post-infection, the cells
were lysed and the lysates were analyzed for CAT activity using Boehringer
Mannheim's CAT ELISA kit. CAT activity was detected in cells that had been
transfected with N, P, and L plasmids together with pRSVA₂CAT. However, no
25 CAT activity was detected when any one of the expression plasmids was
omitted. Furthermore, co-transfection of RSV-GFP-CAT with the N, P, and L
expression plasmids resulted in expression of both GFP and CAT proteins. The
ratios of different expression plasmids and moi of the recombinant vaccinia
virus were optimized in the reporter gene expression system.

6.3.2. RECOVERY OF INFECTIOUS RSV FROM THE COMPLETE RSV cDNA

Hep-2 cells were infected with MVA (recombinant vaccinia virus expressing T7 polymerase) at an moi of one. Fifty minutes later, transfection mixture was added onto the cells. The transfection mixture consisted of 2 µg of N expression vector, 2 µg of P expression vector, 1 µg of L expression vector, 1.25 µg of M2/ORF1 expression vector, 2 µg of RSV genome clone with enhanced promoter, 50 µl of LipofecTACE (Life Technologies, Gaithersburg, M.D.) and 1 ml OPTI-MEM. One day later, the transfection mixture was replaced by MEM containing 2% FCS. The cells were incubated at 37°C for 2 days. The transfection supernatant was harvested and used to infect fresh Hep-2 cells in the presence of 40 µg/ml arabinoside (drug against vaccinia virus). The infected Hep2 cells were incubated for 7 days. After harvesting the P1 supernatant, cells were used for immunostaining using antibodies directed against F protein of RSV A2 strain. Six positively stained loci with visible cell-cell-fusion (typical for RSV infection) were identified. The RNA was extracted from P1 supernatant, and used as template for RT-PCR analysis. PCR products corresponding to F and M2 regions were generated. both products contained the introduced markers. In control, PCR products derived from natural RSV virus lacked the markers.

A point mutation was created at position 4 of the leader sequence of the RSV genome clone (C residue to G) and this genome clone was designated pRSVC4GLwt. This clone has been shown in a reporter gene context to increase the promoter activity by several fold compared to wild-type. After introduction of this mutation into the full-length genome, infectious virus was rescued from the cDNA clone. The rescued recombinant RSV virus formed smaller plaques than the wild-type RSV virus (Figure 8).

This system allows the rescue mutated RSV. Therefore, it may be an excellent tool to engineer live-attenuated vaccines against RSV and to use RSV vector and viruses to achieve heterologous gene expression. It may be possible to express G protein of type B RSV into the type A background, so the vaccine

is capable of protect both type A and type B RSV infection. It may also be possible to achieve attenuation and temperature sensitive mutations into the RSV genome, by changing the gene order or by site-directed mutagenesis of the L protein.

5

6.4. USE OF MONOCLONAL ANTIBODIES TO DIFFERENTIATE RESCUED VIRUS FROM HELPER VIRUS

10 In order to neutralize the RSV strain B9320 helper virus and facilitate identification of rescued A2 strain RSV, monoclonal antibodies against RSV strain B9320 were made as follows.

15 Six BALB/c female mice were infected intranasally (i.n.) with 10^5 plaque forming units (p.f.u.) of RSV B9320, followed 5 weeks later by intraperitoneal (i.p.) inoculation with 10^6 - 10^7 pfu of RSV B9320 in a mixture containing 50% complete Freund's adjuvant. Two weeks after i.p. inoculation, a blood sample from each mouse was tested for the presence of RSV specific antibody using a standard neutralization assay (Beeler and Coelingh, J. Virol. 63:2941-2950 (1988)). Mice producing the highest level of neutralizing antibody were then further boosted with 10^6 p.f.u. of RSV strain B9320 in phosphate buffered saline (PBS), injected intravenously at the base of the tail. Three days later, the mice were sacrificed and their spleens collected as a source of monoclonal antibody producing B-cells. Splenocytes (including B-cells) were teased from the mouse spleen through incisions made in the spleen capsule into 5 ml of Dulbecco's Modified Eagle's Medium (DME). Clumps of cells were allowed to settle out, 20 and the remaining suspended cells were separately collected by centrifugation at 2000xg for 5 minutes at room temperature. These cell pellets were resuspended in 15 ml 0.83 (W/V) NH_4Cl , and allowed to stand for 5 minutes to lyse red blood cells. Splenocytes were then collected by centrifugation as before through a 10 ml; cushion of fetal calf serum. The splenocytes were then rinsed in DME, 25 repelleted and finally resuspended in 20 ml of fresh DME. These splenocytes were then mixed with Sp2/0 cells (a mouse myeloma cell line used as fusion 30

partners for the immortalization of splenocytes) in a ratio of 10:1, spleen cells: Sp2/0 cells. Sp2/0 cells were obtained from the ATCC and maintained in DME supplemented with 10% fetal bovine serum. The cell mixture was then centrifuged for 8 minutes at 2000xg at room temperature. The cell pellet was resuspended in 1 ml of 50% polyethylene glycol 1000 mol. wt. (PEG 1000), followed by addition of equal volumes of DME at 1 minute intervals until a final volume of 25 ml was attained. The fused cells were then pelleted as before and resuspended at 3.5×10^6 spleen cells ml^{-1} in growth medium (50% conditioned medium from SP2/0 cells, 50% HA medium containing 100 ml RPMI 25 ml F.C.S., 100 μgm1 gentamicin, 4 ml 50X Hypoxanthine, Thymidine, Aminopterin (HAT) medium supplied as a prepared mixture of Sigma Chem. Co., St. Louis, MO). The cell suspension was distributed over well plates (200 μl well⁻¹) and incubated at 37°C, 95 humidity and 5% CO₂. Colonies of hybridoma cells (fused splenocytes and Sp2/0 cells) were then subcultured into 24 well plates and grown until nearly confluent; the supernatant growth medium was then sampled for the presence of RSV strain B9320 neutralizing monoclonal antibody, using a standard neutralization assay (Beeler and Coelingh, J. Virol. 63:2941-50 (1988)). Hybridoma cells from wells with neutralizing activity were resuspended in growth medium and diluted to give a cell density of 0.5 cells per 100 μl and plated out in 96 well plates, 200 μl per well. This procedure ensured the production of monoclones (i.e. hybridoma cell lines derived from a single cell) which were then reassayed for the production of neutralizing monoclonal antibody. Those hybridoma cell lines which produced monoclonal antibody capable of neutralizing RSV strain B9320 but not RSV strain A2 were subsequently infected into mice, i.p. (10^6 cells per mouse). Two weeks after the i.p. injection mouse ascites fluid containing neutralizing monoclonal antibody for RSV strain B9320 was tapped with a 19 gauge needle, and stored at -20°C.

This monoclonal antibody was used to neutralize the RSV strain B9320 helper virus following rescue of RSV strain A2 as described in Section 9.1. This was carried out by diluting neutralizing monoclonal antibody 1 in 50 with

molten 0.4% (w/v) agar in Eagle's Minimal Essential Medium (EMEM) containing 1% F.C.S. This mixture was then added to Hep-2 cell monolayers, which had been infected with the progeny of rescue experiments at an m.o.i. of 0.1-0.01 p.f.u. per cell. The monoclonal antibody in the agar overlay inhibited the growth of RSV strain B9320, but allowed the growth of RSV strain A2, resulting in plaque formation by the A2 strain. These plaques were picked using a pasteur pipette to remove a plug a agar above the plaque and the infected cells within the plaque; the cells and agar plug were resuspended in 2 ml of EMEM, 1% FCS, and released virus was plaqued again in the presence of monoclonal antibody on a fresh Hep-2 cell monolayer to further purify from helper virus. The twice plaqued virus was then used to infect Hep-2 cells in 24 well plates, and the progeny from that were used to infect six-well plates at an m.o.i. of 0.1 p.f.u. per cell. Finally, total infected cell RNA from one well of a six-well plates was used in a RT/PCR reaction using first and second strand primers on either side of the 'marker sequences' (introduced into the RSV strain A2 genome to act as a means of recognizing rescue events) as described in Section 6.2 above. The DNA produced from the RT/PCR reaction was subsequently digested with Stu I and Pme I to positively identify the 'marker sequences' introduced into RSV strain A2 cDNA, and hence to establish the validity of the rescue process.

7. RESCUE OF INFECTIOUS RSV PARTICLES IN THE ABSENCE OF M2 EXPRESSION

The following experiments were conducted to compare the efficiencies of rescue of RS virions in the presence and absence of the M2/ORF1 gene. If the M2/ORF1 gene function is not required to achieve rescue of RSV infectious particles, it should be possible to rescue RS virions in the absence of the expression of the M2/ORF1 gene function. In the present analysis, Hep-2 cells which are susceptible to RSV replication, were co-transfected with plasmids encoding the 'N', 'P' and 'L' genes of the viral polymerase of RSV and the cDNA corresponding to the full-length antigenome of RSV, in the presence or absence of plasmid DNA encoding the M2/ORF1 gene, and the number of RSV

infectious units were measured in order to determine whether or not the M2/ORF1 gene product was required to rescue infectious RSV particles.

The following plasmids were used in the experiments described below: a cDNA clone encoding the full-length antigenome of RSV strain A2, designated pRSVC4GLwt; and plasmids encoding the N, P, and L polymerase proteins, and plasmid encoding the M2/ORF1 elongation factor, each downstream of a T7 RNA promoter, designated by the name of the viral protein encoded.

pRSVC4GLwt was transfected, together with plasmids encoding proteins N, P and L, into Hep-2 cells which had been pre-infected with a recombinant vaccinia virus expressing the T7 RNA polymerase (designated MVA). In another set of Hep-2 cells, pRSVC4GLwt was co-transfected with plasmids encoding the N, P and L polymerase proteins, and in addition a plasmid encoding the M2 function. Transfection and recovery of recombinant RSV were performed as follows: Hep-2 cells were split in six-well dishes (35mm per well) 5 hours or 24 hours prior to transfection. Each well contained approximately 1×10^6 cells which were grown in MEM (minimum essential medium) containing 10% FBS (fetal bovine serum). Monolayers of Hep-2 cells at 70% - 80% confluence were infected with MVA at a multiplicity of infection (moi) of 5 and incubated at 35°C for 60 minutes. The cells were then washed once with OPTI-MEM (Life Technologies) and the medium of each dish replaced with 1 ml of OPTI-MEM and 0.2 ml of the transfection mixture. The transfection mixture was prepared by mixing the four plasmids, pRSVC4GLwt, N, P and L plasmids in a final volume of 0.1 ml OPTI-MEM at amounts of 0.5 - 0.6 µg of pRSVC4GLwt, 0.4 µg of N plasmid, 0.4 µg of P plasmid, and 0.2 µg of L plasmid. A second mixture was prepared which additionally included 0.4 µg M2/ORFI plasmid. The plasmid mixtures of 0.1 ml were combined with 0.1 ml of OPTI-MEM containing 10 µl of lipofecTACE (Life Technologies, Gaithersburg, M.D.) to constitute the complete transfection mixture. After a 15 minute incubation at room temperature, the transfection mixture was added to the cells, and one day later this was replaced by MEM containing 2% FBS. Cultures were incubated at 35°C for 3 days at which time the supernatants were

harvested. Cells were incubated at 35°C since the MVA virus is slightly temperature sensitive and is much more efficient at 35°C.

Three days post-transfection, the transfected cell supernatants were assayed for the presence of RSV infectious units by an immunoassay which would indicate the presence of RSV packaged particles (see Table 1). In this assay, 0.3 - 0.4 ml of the culture supernatants were passaged onto fresh (uninfected) Hep-2 cells and overlaid with 1% methylcellulose and 1 x L15 medium containing 2% FBS. After incubation for 6 days, the supernatant was harvested and the cells were fixed and stained by an indirect horseradish peroxidase method, using a goat anti-RSV antibody which recognizes the RSV viral particle (Biogenesis, Sandown, NH) followed by a rabbit anti-goat antibody conjugated to horseradish peroxidase. The antibody complexes that bound to RSV-infected cells were detected by the addition of a AEC-(3-amino-9-ethylcarbazole) chromogen substrate (DAKO) according to the manufacturer's instructions. The RSV plaques were indicated by a black-brown coloration resulting from the reaction between the chromogen substrate and the RSV-antibody complexes bound to the plaques. The number of RSV plaques is expressed as the number of plaque forming units (p.f.u.) per 0.5 ml of transfection supernatant (see Table 1).

Comparisons of the amount of RS virions recovered from the supernatants of transfection dishes in the presence or absence of M2/ORF1 are shown in Table 1. The results of four separate experiments demonstrated that the absence of M2/ORF1 from the transfection assay did not diminish the number of infectious units of RSV observed. Thus, the results of these experiments clearly indicate that RSV can be rescued in the absence of the M2/ORF1 from cells transfected only with plasmids encoding the three polymerase proteins, N, P and L, and the cDNA encoding the full-length RSV antigenome. The rescue of true RS virions in the absence of M2/ORF1 was further indicated by the ability to passage the rescued recombinant RSV for up to six passages. Therefore, the production of RSV virions is not dependent on

the expression of the M2/ORF1 gene, nor does the inclusion of the M2/ORF1 gene in the transfection assay increase the efficiency of true RSV rescue.

5

Table 1. Production of infectious RSV through plasmid transfection is not dependent on expression of M2ORF1

Expt.	Production of infectious RSV (pfu from 0.5ml transfection supernatants)	
	+M2 ORF1	-M2 ORF1
1.	6,10(8)	16,9(13)
2.	120,46,428(198)	100,122,105(109)
3.	160,180(170)	150,133(142)
4.	588,253,725(522)	300,1000,110(470)

15

Each experiment was done singly, in duplicates or triplicates. The average number of plaque forming units (pfu) from 0.5 ml transfected cell supernatants is shown in the brackets.

8. EXAMPLE: EXPRESSION OF RSV SUBGROUP B-G AND -F PROTEINS BY RSV A2 STRAIN

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The following experiments were conducted to generate a chimeric RSV which expresses the antigenic polypeptides of more than one strain of RSV. Two main antigenic subgroups (A and B) of respiratory syncytial virus (RSV) cause human diseases. Glycoproteins F and G are the two major antigenic determinants of RSV. The F glycoproteins of subgroup A and B viruses are estimated to be 50% related, while the relationship of G glycoproteins is considerably less, about 1-5%. Infection of RSV subgroup A induces either partial or no resistance to replication of a subgroup B strain and vice versa. Both subgroup A and subgroup B RSV virus vaccines are needed to protect from RSV infection.

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The first approach described herein is to make an infectious chimeric RSV cDNA clone expressing subgroup B antigens by replacing the current

infectious RSV A2 cDNA clone G and F region with subgroup B-G and -
F genes. The chimeric RSV would be subgroup B antigenic specific. The
second approach described herein is to insert subgroup B-G gene in the current
A2 cDNA clone so that one virus would express both subgroup A and B specific
antigens.

8.1. SUBSTITUTION OF A2 G AND F BY B9320 G AND F GENES

RSV subgroup B strain B9320 G and F genes were amplified from
B9320 vRNA by RT/PCR and cloned into pCRII vector for sequence
determination. BamH I site was created in the oligonucleotide primers used for
RT/PCR in order to clone the G and F genes from B9320 strain into A2
antigenomic cDNA (Fig. 4A). A cDNA fragment which contained G and F
genes from 4326 nt to 9387 nt of A2 strain was first subcloned into pUC19
(pUCR/H). Bgl II sites were created at positions of 4630 (SH/G intergenic
junction) and 7554 (F/M2 intergenic junction), respectively by Quickchange
site-directed mutagenesis kit (Stratagene, Lo Jolla, CA). B9320 G and F cDNA
inserted in pCR.II vector was digested with BamH I restriction enzyme and then
subcloned into Bgl II digested pUCR/H which had the A2 G and F genes
removed. The cDNA clone with A2 G and F genes replaced by B9320 G and F
was used to replace the Xho I to Msc I region of the full-length A2 antigenomic
cDNA. The resulting antigenomic cDNA clone was termed pRSVB-GF and
was used to transfect Hep-2 cells to generate infectious RSVB-GF virus.

Generation of chimeric RSVB-GF virus was as follows, pRSVB-GF was
transfected, together with plasmids encoding proteins N, P, and L, into Hep-2
cells which had been infected with MVA, a recombinant vaccinia virus which
expresses the T7 RNA polymerase. Hep-2 cells were split a day before
transfection in six-well dishes. Monolayers of Hep-2 cells at 60% - 70%
confluence were infected with MVA at moi of 5 and incubated at 35°C for
60 min. The cells were then washed once with OPTI-MEM (Life Technologies,

Gaithersburg, MD). Each dish was replaced with 1 ml of OPTI-MEM and added with 0.2 ml of transfection medium. The transfection medium was prepared by mixing five plasmids in a final volume of 0.1 ml of OPTI-MEM medium, namely 0.6 µg of RSV antigenome pRSVB-GF, 0.4 µg of N plasmid, 0.4 µg of P plasmid, and 0.2 µg of L plasmid. This was combined with 0.1 ml of OPTI-MEM containing 10 µl lipofecTACE (Life Technologies, Gaithersburg, MD U.S.A.). After a 15 minute incubation at room temperature, the DNA/lipofecTACE was added to the cells and the medium was replaced one day later by MEM containing 2% FBS. Cultures were further incubated at 35°C for 3 days and the supernatants harvested. Aliquots of culture supernatants were then used to infect fresh Hep-2 cells. After incubation for 6 days at 35°C, the supernatant was harvested and the cells were fixed and stained by an indirect horseradish peroxidase method using goat anti-RSV antibody (Biogenesis, Sandown, NH) followed by a rabbit anti-goat antibody linked to horseradish peroxidase. The virus infected cells were then detected by addition of substrate chromogen (DAKO, Carpinteria, CA, U.S.A.) according to the manufacturer's instructions. RSV-like plaques were detected in the cells which were infected with the supernatants from cells transfected with pRSVB-GF. The virus was further plaque purified twice and amplified in Hep-2 cells.

Recombinant RSVB-GF virus was characterized by RT/PCR using RSV subgroup B specific primers. Two independently purified recombinant RSVB-GF virus isolates were extracted with an RNA extraction kit (Tel-Test, Friendswood, TX) and RNA was precipitated by isopropanol. Virion RNAs were annealed with a primer spanning the RSV region from nt 4468 to 4492 and incubated for 1 hr under standard RT conditions (10 µl reactions) using superscript reverse transcriptase (Life Technologies, Gaithersburg, MD). Aliquots of each reaction were subjected to PCR (30 cycles at 94°C for 30 s, 55°C for 30 s and 72°C for 2 min) using subgroup B specific primers in G region (CACCACCTACCTTACTCAAGT and TTTGTTTGTGGGTTTGATGGTTGG). The PCR products were analyzed by electrophoresis on 1% agarose gel and visualized by staining with ethidium

bromide. As shown in Fig. 5, no DNA product was produced in RT/PCR reactions using RSV A2 strain as template. However, a predicted product of 254 bp was detected in RT/PCR reactions utilizing RSVB-GF RNA or the PCR control plasmid, pRSVB-GF DNA, as template, indicating the rescued virus contained G and F genes derived from B9320 virus.

8.2. EXPRESSION OF B9320G BY RSV A2 VIRUS

RSV subgroup B strain B9320 G gene was amplified from B9320 vRNA by RT/PCR and cloned into pCRII vector for sequence determination. Two Bgl II sites were incorporated into the PCR primers which also contained gene start and gene end signals (GATATCAAGATCTACAATAACATTGGGGCAAATGC and GCTAAGAGATCTTTTT GAATAACTAAGCATG). B9320G cDNA insert was digested with Bgl II and cloned into the SH/G (4630 nt) or F/M2 (7552 nt) intergenic junction of a A2 cDNA subclone (Fig. 4B and Fig. 4C). The Xho I to Msc I fragment containing B9320G insertion either at SH/G or F/M2 intergenic region was used to replace the corresponding Xho I to Msc I region of the A2 antigenomic cDNA. The resulting RSV antigenomic cDNA clone was termed as pRSVB9320G-SH/G or pRSVB9320G-F/M2.

Generation of RSV A2 virus which had B9320 G gene inserted at F/M2 intergenic region was performed similar to what has described for generation of RSVB-GF virus. Briefly, pRSVB9320G-F/M2 together with plasmids encoding proteins N, P and L were transfected, into Hep-2 cells, infected with a MVA vaccinia virus recombinant, which expresses the T7 RNA polymerase (Life Technologies, Gaithersburg, M.D.). The transfected cell medium was replaced by MEM containing 2% fetal bovine serum (FBS) one day after transfection and further incubated for 3 days at 35°C. Aliquots of culture supernatants (PO) were then used to infect fresh Hep-2 cells. After incubation for 6 days at 35°C, the supernatant was harvested and the cells were fixed and stained by an indirect horseradish peroxidase method using goat anti-RSV antibody (Biogenesis)

followed by a rabbit anti-goat antibody linked to horseradish peroxidase. The virus infected cells were then detected by addition of substrate chromogen (Dako). RSV-like plaques were detected in the cells which were infected with the supernatants from cells transfected with pRSVB9320G-F/M2.

5 Characterization of pRSVB9320G-F/M2 virus was performed by RT/PCR using B9320G specific primers. A predicted PCR product of 410bp was seen in RT/PCR sample using pRSVB9320G-F/M2 RNA as template, indicating the rescued virus contained G gene derived from B9320. (Figure 6)

10 Expression of the inserted RSV B9320 G gene was analyzed by Northern blot using a ³²P-labeled oligonucleotide specific to A2-G or B-G mRNA. Total cellular RNA was extracted from Hep-2 cells infected with wild-type RSVB 9320, rRSVA2, or rRSVB9320G-F/M2 48 hours postinfection using an RNA extraction kit (RNA stat-60, Tel-Test). RNA was electrophoresed on a 1.2% agarose gel containing formaldehyde and transferred to a nylon membrane
15 (Amersham). An oligonucleotide specific to the G gene of the A2 strain (5'TCTTGACTGTTGTGGATTGCAGGGTTGACTTGACTCCGATCGATCC-3') and an oligonucleotide specific to the B9320 G gene (5'CTTGTGTTGTTGTTGTATGGTGT
20 GTTTCTGATTTTGTATTGATCGATCC-3') were labeled with ³²P-ATP by a kinasing reaction known to those of ordinary skill in the art. Hybridization of the membrane with one of the ³²P-labeled G gene specific oligonucleotides was performed at 65 °C and washed according to standard procedure. Both A2-G and B9320-G specific RNA were detected in the rRSVB9320G-F/M2 infected Hep-2 Cells. (Figure 6B) These results demonstrate subtype specific RNA
25 expression.

30 Protein expression of the chimeric rRSVA2(B-G) was compared to that of RSV B9320 and rRSV by immunoprecipitation of ³⁵S-labeled infected Hep-2 cell lysates. Briefly, the virus infected cells were labeled with ³⁵S-promix (100 µCi/ml ³⁵S-Cys and ³⁵S-Met, Amersham, Arlington Heights, IL) at 14 hours to 18 hours post-infection according to a protocol known to those of ordinary skill in the art. The cell monolayers were lysed by RIPA buffer and the polypeptides

were immunoprecipitated with either polyclonal antiserum raised in goat against detergent disrupted RSV A2 virus (Fig. 7, lanes 1-4) or antiserum raised in mice against undisrupted B9320 virions (Fig. 7, lanes 5-8). The radio labeled immunoprecipitated polypeptides were electrophoresed on 10% polyacrylamide gels containing 0.1% SDS and detected by autoradiography. Anti-RSV A2 serum immunoprecipitated the major polypeptides of the RSV A2 strain, whereas anti-B9320 serum mainly reacted with RSV B9320 G protein and the conserved F protein of both A and B subgroups. As shown in Fig. 7, a protein which is identical to the A2-G protein (lane 3), was immunoprecipitated from the rRSVA2(B-G) infected cells (lane 4) by using an antiserum against RSV A2. The G protein of RSV B9320 strain was not recognized by the anti-A2 antiserum. A protein species, smaller than A2-G protein, was immunoprecipitated from both B9320 (lane 6) and rRSVA2(B-G)(lane 9) infected cells using the antiserum raised in mice against B9320 virions. This polypeptide was not present in the uninfected and RSV A2 infected cells and likely is to represent the G protein specific to the RSV B 9320 strain. Amino acid sequence comparison of both A2 and B9320 RSV G proteins indicated that two additional potential N-glycosylation sites (N-X-S/t) are present in the RSV A2G protein, which may contribute to slower migration of the A2 G protein under the conditions used. The F protein of RSV B9320 also migrated slightly faster than RSV A2 F protein. The P and M proteins also showed mobility differences between the two virus subtypes. The identity of the polypeptide near the top of the protein gel present in FSV B9320 and rRSVA2(B-G) infected cells is not known. Antisera raised in mice against the RSV B9320 virions poorly recognized the N, P and M proteins are compared to the goat antiserum raised against the RSV A2 strain. The data described above clearly indicate that chimeric rRSV A2(B-G) expresses both the RSV A2 and B9320 specific G proteins.

8.2.1 REPLICATION OF RECOMBINANT RSV IN TISSUE CULTURE

Recombinant RS viruses were plaque purified three times and amplified in Hep-2 cells. Plaque assays were performed in Hep-2 cells in 12-well plates using an overlay of 1% methylcellulose and 1 x L15 medium containing 2% fetal bovine serum (FBS). After incubation at 35°C for 6 days, the monolayers were fixed with methanol and plaques were identified by immunostaining. Plaque size and morphology of rRSV was very similar to that of wild-type A2 RSV (Fig. 8). However, the plaques formed by rRSVC4G were smaller than rRSV and wild-type A2 virus. The only genetic difference between rRSV and rRSVC4 was a single nucleotide substitution in the RSV leader region. Therefore, the smaller plaque size of rRSV A2(B-G) was not distinguishable from that of rRSVC4G.

The growth curves of rRSV, rRSVC4G and rRSV A2 (B-G) were compared to that of the biologically derived wild-type A2 virus. Hep-2 cells were grown in T25 culture flasks and infected with rRSV, rRSVC4G, rRSVA2(B-G), or wild-type RSV A2 strain at a moi of 0.5. After 1 hour adsorption at 37°C, the cells were washed three times with MEM containing 2% FBS and incubated at 37°C in 5% CO₂. At 4 hour intervals post-infection, 250 µl of the culture supernatant was collected, and stored at -70°C until virus titration. Each aliquot taken was replaced with an equal amount of fresh medium. The titer of each virus was determined by plaque assay on Hep-2 cells and visualized by immunostaining (*vide supra*). As shown in Fig. 9, the growth kinetics of rRSV is very similar to that of wild-type A2 virus. Maximum virus titer for all the viruses were achieved between 48 hr to 72 hr. The virus titer of rRSVC4G was about 2.4-fold (at 48 hr) and 6.6-fold (at 72 hr) lower than rRSV and wild-type A2 RSV. The poor growth of rRSVC4G may also be due to the single nucleotide change in the leader region. The chimeric rRSV A2(B-G) showed slower kinetics and lower peak titer (Fig. 9).

9. EXAMPLE: GENERATION OF RSV L GENE MUTANTS

The strategy for generating L gene mutants is to introduce defined mutations or random mutations into the RSV L gene. The functionality of the L gene cDNA mutants can be screened *in vitro* by a minigenome replication system. The recovered L gene mutants are then further analyzed *in vitro* and *in vivo*.

9.1 MUTAGENESIS STRATEGIES

9.1.1 SCANNING MUTAGENESIS TO CHANGE THE CLUSTERED CHARGED AMINO ACIDS TO ALANINE

This mutagenesis strategy has been shown to be particularly effective in systematically targeting functional domains exposed on protein surfaces. The rationale is that clusters of charged residues generally do not lie buried in the protein structure. Making conservative substitutions of these charged residues with alanines will therefore remove the charges without grossly changing the structure of the protein. Disruption of charged clusters may interfere with the interaction of RSV L protein with other proteins and make its activity thermosensitive, thereby yielding temperature-sensitive mutants.

A cluster was originally defined arbitrarily as a stretch of 5 amino acids in which two or more residues are charged residues. For scanning mutagenesis, all the charged residues in the clusters can be changed to alanines by site directed mutagenesis. Because of the large size of the RSV L gene, there are many clustered charged residues in the L protein. Therefore, only contiguous charged residues of 3 to 5 amino acids throughout the entire L gene were targeted (Fig. 10). The RSV L protein contains 2 clusters of five contiguous charged residues, 2 clusters of four contiguous charged residues and 17 clusters

of three contiguous charge residues. Two to four of the charged residues in each cluster were substituted with alanines.

The first step of the invention was to introduce the changes into pCITE-L which contains the entire RSV L-gene, using a QuikChange site-directed mutagenesis kit (Stratagene). The introduced mutations were then confirmed by sequence analysis.

9.1.2. CYSTEINE SCANNING MUTAGENESIS

Cysteines are good targets for mutagenesis as they are frequently involved in intramolecular and intermolecular bond formations. By changing cysteines to glycines or alanines, the stability and function of a protein may be altered because of disruption of its tertiary structure. Thirty-nine cysteine residues are present in the RSV L protein (Fig. 11). Comparison of the RSV L protein with other members of paramyxoviruses indicates that some of the cysteine residues are conserved.

Five conserved cysteine residues were changed to either valine (conservative change) or to aspartic acids (nonconservative change) using a QuikChange site-directed mutagenesis kit (Stratagene) degenerate mutagenic oligonucleotides. It will be apparent to one skilled in the art that the sequence of the mutagenic oligonucleotides is determined by the protein sequence desired. The introduced mutations were confirmed by sequence analysis.

9.1.3. RANDOM MUTAGENESIS

Random mutagenesis may change any residue, not simply charged residues or cysteines. Because of the size of the RSV L gene, several L gene cDNA fragments were mutagenized by PCR mutagenesis. This was accomplished by PCR using *exo*⁻ Pfu polymerase obtained from Stratagene. Mutagenized PCR fragments were then cloned into a pCITE-L vector. Sequencing analysis of 20 mutagenized cDNA fragments indicated that 80% -

90% mutation rates were achieved. The functionality of these mutants was then screened by a minigenome replication system. Any mutants showing altered polymerase function were then further cloned into the full-length RSV cDNA clone and virus recovered from transfected cells.

5

9.2. FUNCTIONAL ANALYSIS OF RSV L PROTEIN MUTANTS BY MINIGENOME REPLICATION SYSTEM

10 The functionality of the L-genes mutants were tested by their ability to replicate a RSV minigenome containing a CAT gene in its antisense and flanked by RSV leader and trailer sequences. Hep-2 cells were infected with MVA vaccinia recombinants expressing T7 RNA polymerase. After one hour, the cells were transfected with plasmids expressing mutated L protein together with plasmids expressing N protein and P protein, and pRSV/CAT plasmid
15 containing CAT gene (minigenome). CAT gene expression from the transfected cells was determined by a CAT ELISA assay (Boehringer Mannheim) according to the manufacturer's instruction. The amount of CAT activity produced by the L gene mutant was then compared to that of wild-type L protein.

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9.3. RECOVERY OF MUTANT RECOMBINANT RSV

To recover or rescue mutant recombinant RSV, mutations in the L-gene were engineered into plasmids encoding the entire RSV genome in the positive sense (antigenome). The L gene cDNA restriction fragments (BamH I and Not
25 I) containing mutations in the L-gene were removed from pCITE vector and cloned into the full-length RSV cDNA clone. The cDNA clones were sequenced to confirm that each contained the introduced mutations.

Each RSV L gene mutant virus was rescued by co-transfection of the following plasmids into subconfluent Hep-2 cells grown in six-well plates.
30 Prior to transfection, the Hep-2 cells were infected with MVA, a recombinant

vaccinia virus which expresses T7 RNA polymerase. One hour later, cells were transfected with the following plasmids:

- pCITE-N: encoding wild-type RSV N gene, 0.4 µg
- pCITE-P: encoding wild-type RSV P gene, 0.4 µg
- pCITE-Lmutant: encoding mutant RSV L gene, 0.2 µg
- pRSVL mutant: full-length genomic RSV of the positive sense (antigenome) containing the same L-gene mutations as pCITE-L mutant, 0.6 µg

DNA was introduced into cells by lipofectACE (Life Technologies) in OPTI-MEM. After five hours or overnight transfection, the transfection medium was removed and replaced with 2% MEM. Following incubation at 35°C for three days, the media supernatants from the transfected cells were used to infect Vero cells. The virus was recovered from the infected Vero cells and the introduced mutations in the recovered recombinant viruses confirmed by sequencing of the RT/PCR DNA derived from viral RNA.

Examples of the L gene mutants obtained by charged to alanine scanning mutagenesis are shown in the Table 2. Mutants were assayed by determining the expression of CAT by pRSV/CAT minigenome following co-transfection of plasmids expressing N, P and either wild-type or mutant L. Cells were harvested and lysed 40 hours post-transfection after incubation at 33°C or 39°C. The CAT activity was monitored by CAT ELISA assay (Boehringer Mannheim). Each sample represents the average of duplicate transfections. The amount of CAT produced for each sample was determined from a linear standard curve.

From the above preliminary studies, different types of mutations have been found.

9.3.1. DETRIMENTAL MUTATIONS

Seven L protein mutants displayed a greater than 99% reduction in the amount of CAT produced compared to that of wild-type L protein. These

mutations drastically reduced the activity of the RSV polymerase and are not expected to be viable.

Table 2 CAT Expression levels of Mutant RSV L-gene

	Mut.	Conc. of CAT (ng/mL)		Charge Cluster	Charge to Alanine Change	Rescued Virus
		33°C	39°C			
5	A33	0.246	Bkg	5	135E, 136K	No
	A73	3.700	0.318	3	146D, 147E, 148 D	Yes
	A171	3.020	Bkg	3	157K, 158D	Yes
	A81	1.000	0.280	3	255H, 256K	Yes
	A185	Bkg	Bkg	3	348E, 349E	No
10	A91	Bkg	Bkg	3	353R, 355R	No
	A101	Bkg	Bkg	3	435D, 436E, 437R	No
	A192	1.960	Bkg	3	510E, 511R	Yes
	A11	0.452	Bkg	1	520R	Yes
	A111	2.320	0.267	4	568H, 569E	Yes
15	A121	0.772	Bkg	2	587L, 588R	No
	A133	Bkg	Bkg	4	620E, 621R	No
	A141	2.800	Bkg	3	1025K, 1026D	Yes
	A25	0.169	Bkg	3	1033D, 1034D	Yes
	A45	5.640	0.478	5	1187D, 1188K	Yes
20	A153	4.080	0.254	5	1187D, 1188K, 1189R, 1190E	Yes
	A162	10.680	Bkg	3	1208E, 1209R	No
	A201	Bkg	Bkg	3	1269E, 1270K	No
	A211	2.440	0.047	3	1306D, 1307E	Yes
	A221	0.321	Bkg	3	1378D, 1379E	No
25	A231	Bkg	Bkg	3	1515E, 1516K	No
	A241	1.800	0.308	3	1662H, 1663K	Yes
	A57	5.660	0.706	3	1725D, 1726K	Yes

Mut.	Conc. of CAT (ng/mL)		Charge Cluster	Charge to Alanine Change	Rescued Virus
	33°C	39°C			
A65	3.560	0.168	2	1957R, 1958K	Yes
A251	0.030	Bkg.	3	2043D, 2044K	Yes
A261	Bkg	Bkg	3	2102K, 2103H	No
AD11	2.800	0.456	5 and 3	1187D, 1188K, 1725D, 1726K	No
AD21	2.640	0.226	5 and 2	1187D, 1188K, 1957R, 1958K	No
AD31	1.280	0.192	3 and 2	1725D, 1726K, 1957R, 1958K	No
F1	Bkg	Bkg	-	521 F to L	Yes
F13	0.13	Bkg	-	521 F to L	Yes
Lwt	3.16	-	-	no amino acid changes	Yes

9.3.2. INTERMEDIATE MUTATIONS

Several L mutants showed an intermediate level of CAT production which ranged from 1% to 50% of that wild-type L protein. A subset of these mutants were introduced into virus and found to be viable. Preliminary data indicated that mutant A2 showed 10-to 20-fold reduction in virus titer when grown at 40°C compared 33°C. Mutant A25 exhibited a smaller plaque formation phenotype when grown at both 33°C and 39°C. This mutant also had a 10-fold reduction in virus titer at 40°C compared to 33°C.

9.3.3 **MUTANTS WITH L PROTEIN FUNCTION SIMILAR OR HIGHER THAN WILD TYPE L PROTEIN**

Some L gene mutants produced CAT gene expression levels similar to or greater than the wild-type L protein *in vitro* and the recovered virus mutants have phenotypes indistinguishable from wild-type viruses in tissue culture.

Once mutations in L that confer temperature sensitivity and attenuation have been identified, the mutations will be combined to test for the cumulative effect of multiple temperature-sensitivity markers. The L mutants bearing more than one temperature sensitive marker are expected to have lower permissive temperature and to be genetically more stable than single-marker mutants.

The generated L gene mutants may also be combined with mutations present in other RSV genes and/or with non-essential RSV gene deletion mutants (e.g., SH, NS1 and NS2 deletion). This will enable the selection of safe, stable and effective live attenuated RSV vaccine candidates.

10. **GENERATION OF HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL SH AND M2ORF2 GENES**

10.1. **M2-2 DELETION MUTANT**

To delete M2-2 genes, two Hind III restriction enzyme sites were introduced at RSV nucleotides 8196 and 8430, respectively, in a cDNA subclone pET(S/B) which contained an RSV restriction fragment from 4478 to 8505. The RSV restriction fragment had been previously prepared by Quikchange site-directed mutagenesis (Stratagene, La Jolla, CA). Digestion of pET(S/B) with Hind III restriction enzyme removed a 234 nucleotide sequence which contained the majority of the M2-2 open reading frame. The nucleotides encoding the first 13 amino acids at the N-terminus of the M2-2 gene product were not removed because this sequence overlaps M2-1. The cDNA fragment

which contained M2-2 gene deletion was digested with SacI and BamHI and cloned back into a full-length RSV cDNA clone, designated pRSVΔM2-2

Infectious RSV with this M2-2 deletion was generated by transfecting pRSVΔM2-2 plasmid into MVA-infected Hep-2 cells expressing N, P and L genes. Briefly, pRSVΔM2-2 was transfected, together with plasmids encoding proteins N, P and L, into Hep-2 cells which had been infected with a recombinant vaccinia virus (MVA) expressing the T7 RNA polymerase. Transfection and recovery of recombinant RSV was performed as follows. Hep-2 cells were split five hours or a day before the transfection in six-well dishes. Monolayers of Hep-2 cells at 70% - 80% confluence were infected with MVA at a multiplicity of infection (moi) of 5 and incubated at 35°C for 60 min. The cells were then washed once with OPTI-MEM (Life Technologies, Gaithersburg, M.D.). Each dish was replaced with 1 ml of OPTI-MEM and 0.2 ml transfection medium was added. The transfection medium was prepared by mixing 0.5 - 0.6 µg of RSV antigenome, 0.4 µg of N plasmid, 0.4 µg of P plasmid, and 0.2 µg of L plasmid in a final volume of 0.1 ml OPTI-MEM medium. This was combined with 0.1 ml of OPTI-MEM containing 10 µl lipofecTACE (Life Technologies). After a 15 minute incubation at room temperature, the DNA/lipofecTACE mixture was added to the cells. The medium was replaced one day later with MEM containing 2% FBS. Cultures were further incubated at 35°C for 3 days and the supernatants harvested. Three days post-transfection, 0.3 - 0.4 ml culture supernatants were passaged onto fresh Hep-2 cells and incubated with MEM containing 2% FBS. After incubation for six days, the supernatant was harvested and the cells were fixed and stained by an indirect horseradish peroxidase method using goat anti-RSV antibody (Biogenesis) followed by a rabbit anti-goat antibody linked to horseradish peroxidase. The virus infected cells were then detected by addition of substrate chromogen (DAKO) according to the manufacturer's instructions. Recombinant RSV which contained M2-2 gene deletion was recovered from the transfected cells. Identification of rRSVΔM2-2 was performed by RT/PCR using primers flanking the deleted region. As shown in Fig. 12A, a cDNA

fragment which is 234 nucleotides shorter than the wild-type RSV was detected in rRSVΔM2-2 infected cells. No cDNA was detected in the RT/PCR reaction which did not contain reverse transcriptase in the RT reaction. This indicated that the DNA product was derived from viral RNA and not from contamination.

5 The properties of the M2-2 deletion RSV are currently being evaluated.

10.2. SH DELETION MUTANT.

To delete the SH gene from RSV, a Sac I restriction enzyme site was introduced at the gene start signal of SH gene at position of nt 4220. A unique SacI site also exists at the C-terminus of the SH gene. Site-directed mutagenesis was performed in subclone pET(A/S), which contains an AvrII(2129) SacI (4478) restriction fragment. Digestion of pET(A/S) mutant with SacI removed a 258 nucleotide fragment of the SH gene. Digestion of the pET(A/S) subclone containing the SH deletion was digested with Avr II and Sac I and the resulting restriction fragment was then cloned into a full-length RSV cDNA clone. The full-length cDNA clone containing the SH deletion was designated pRSVΔSH.

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Generation of the pRSVΔSH mutant was performed as described above (see 10.1). SH-minus RSV (rRSVΔSH) was recovered from MVA-infected cells that had been co-transfected with pRSVΔSH together with N, P and L expression plasmids. Identification of the recovered rRSVΔSH was performed by RT/PCR using a pair of primers which flanked the SH gene. As shown in Fig. 12A, a cDNA band which is about 258 nucleotides shorter than the wild-type virus was detected in the rRSVΔSH infected cells. No DNA was detected in the RT/PCR reaction which did not have reverse transcriptase in the RT reaction. This indicated that the PCR DNA was derived from viral RNA and was not artifact, and that the virus obtained was truly SH-minus RSV.

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10.3 GENERATION OF BOTH SH AND M2-2 DELETION MUTANT.

Both SH and M2-2 genes were deleted from the full-length RSV cDNA construct by cDNA subcloning. A Sac I to Bam HI fragment containing M2-2 deletion removed from cDNA subclone pET(S/B) Δ M2-2RSV was cloned into pRSV Δ SH cDNA clone. The double gene deletion plasmid pRSV Δ SH Δ M2-2 was confirmed by restriction enzyme mapping. As shown in Fig. 12B, the SH/M2-2 double deletion mutant is shorter than the wild-type pRSV cDNA.

Recovery of infectious RSV containing both the SH and M2-2 deletion was performed as described earlier. Infectious virus with both SH and M2-2 deleted was obtained from transfected Hep-2 cells.

11. EXAMPLE: GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING A VIRAL ACCESSORY GENE(S) EITHER SINGLY OR IN COMBINATION

Rationale:

Human respiratory syncytial virus is the major cause of pneumonia and bronchiolitis in infants under one year of age. RSV is responsible for more than one in five pediatric hospital admissions due to respiratory tract disease and causes 4,500 deaths yearly in the USA alone. Despite decades of investigation to develop an effective vaccine against RSV, no safe and effective vaccine has been achieved to prevent the severe morbidity and significant mortality associated with RSV infection. Various approaches have been used to develop RSV vaccine candidates: formalin-inactivated virus, recombinant subunit vaccine of expressed RSV glycoproteins, and live attenuated virus. Recently, generation of live attenuated RSV mutants has been the focus for the RSV vaccine development. In the past, generation of live attenuated RSV mutant can only be achieved by in vitro passage and/or chemical mutagenesis. Virus was either underattenuated or overattenuated and was not genetically stable. The

present investigation provides an immediate approach to generate genetically stable live attenuated RSV vaccines by deleting an accessory gene(s) individually or in combination. Gene deletions are considered to be a very powerful strategy for attenuating RSV because such deletions will not revert and the recombinant RSV deletion mutants are thus expected to be genetically very stable.

RSV is unique among the paramyxoviruses in its gene organization. In addition to the N, P, L, M, G and F genes which are common to all the paramyxoviruses, RSV contains four additional genes which encode five proteins: NS1, NS2, SH, M2-1 and M2-2. M2-1 and M2-2 are translated from two open reading frames that overlap in the middle of the M2 mRNA. M2-1 enhances mRNA transcriptional processivity and also functions as an antitermination factor by increasing transcriptional readthrough at the intergenic junctions (Collins, P. L. *et al. Proc. Natl. Acad. Sci. U S A* 93, 81-85 (1996); Hardy, R. W. *et al. J. Virol.* 72, 520-526 (1998)). However, the M2-2 protein was found to inhibit RSV RNA transcription and replication in vitro (Collins, P. L. *et al. Proc. Natl. Acad. Sci. U S A* 93, 81-85 (1996)). The accessory protein NS1 was reported to be a potent transcription inhibitor (Atreya, P. L. *et al., J. Virol.* 72, 1452-1461 (1998)). The SH gene has been shown to be dispensable for RSV growth in tissue culture in a naturally occurring virus and in a recombinant RSV harboring an engineered SH deletion (Bukreyev, A. *et al., J Virol* 71(12), 8973-82 (1997); Karron, R. A. *et al. J. Infect. Dis.* 176, 1428-1436 (1997)). SH minus RSV replicates as well as the wild type RSV *in vitro*. Recently, it was reported that the NS2 gene could also be deleted (Teng, M. N., *et al J Virol* 73(1), 466-73 (1999); Buchholz, U. J. *et al. J Virol* 73(1), 251-9 (1999)). Deletion of M2-1, M2-2, and NS1 has not been reported, neither was deletion of more than two nonessential genes reported. Traditionally, live attenuated virus mutants were generated by passaging of RSV at lower temperature for many times and/or mutagenized by chemical reagents. The mutations are introduced randomly and the virus phenotype is difficult to maintain because revertants may develop. The ability to produce virus from an

infectious cDNA makes it possible to delete gene or genes that are associated with virus pathogenesis. Gene deletion alone or in combination with mutations in the other viral genes (G, F, M, N, P and L) may yield a stably attenuated RSV vaccine to effectively protect RSV infection.

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11.1 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL M2-2 GENE

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This example describes production of a recombinant RSV in which expression of the M2-2 gene has been ablated by removal of a polynucleotide sequence encoding the M2-2 gene and its encoded protein. The RSV M2-2 gene is encoded by M2-2 gene and its open reading frame is partially overlapped with the 5'-proximal M2-1 open reading frame by 12 amino acids (Collins, P. L. *et al. Proc. Natl. Acad. Sci. U S A* 93, 81-85 (1996)). The predicted M2-2 polypeptide contains 90 amino acids, but the M2-2 protein has not yet been identified intracellularly. The M2-2 protein down-regulates RSV RNA transcription and replication in a minigenome model system (Collins, P. L. *et al. Proc. Natl. Acad. Sci. USA* 92, 11563-11567 (1995)). The significance of this negative effect on RSV RNA transcription and replication in the viral replication cycle is not known.

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11.1.1 RECOVERY OF RECOMBINANT RSV THAT LACKS THE M2-2 GENE

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To produce a recombinant RSV that no longer expresses the M2-2 protein, the M2-2 gene was deleted from a parental RSV cDNA clone (Jin, H. *et al. Virology* 251, 206-214 (1998)). The antigenomic cDNA clone encodes a complete antigenomic RNA of strain A2 of RSV, which was used successfully to recover recombinant RSV. This antigenomic cDNA contains a single

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nucleotide change in the leader region at position 4 from C to G in its antigenomic sense. The construction of plasmid pA2ΔM2-2 involved a two step cloning procedure. Two *Hind III* restriction enzyme sites were introduced at RSV sequence of 8196 nt and 8430 nt respectively in a cDNA subclone (pET-S/B) that contained RSV *Sac I* (4477nt) to *BamH I* (8504nt) cDNA fragment using Quickchange mutagenesis kit (Stratagene). Digestion of this cDNA clone with *Hind III* restriction enzyme removed the 234 nt *Hind III* cDNA fragment that contained the M2-2 gene. The remaining *Sac I* to *BamH I* fragment that did not contain the M2-2 gene was then cloned into a RSV antigenomic cDNA pRSVC4G. The resulting plasmid was designated as pA2ΔM2-2.

To recover recombinant RSV with the M2-2 open reading frame deleted, pA2ΔM2-2 was transfected, together with plasmids encoding the RSV N, P, and L proteins under the control of T7 promoter, into Hep-2 cells which had been infected with a modified vaccinia virus encoding the T7 RNA polymerase (MVA-T7). After 5 hours incubation of the transfected Hep-2 cells at 35°C, the medium was replaced with MEM containing 2% FBS and the cells were further incubated at 35°C for 3 days. Culture supernatants from the transfected Hep-2 cells were used to infect the fresh Hep-2 or Vero cells to amplify the rescued virus. Recovery of rA2ΔM2-2 was indicated by syncytial formation and confirmed by positive staining of the infected cells using polyclonal anti-RSV A2 serum. Recovered rA2ΔM2-2 was plaque purified three times and amplified in Vero cells. To confirm that rA2ΔM2-2 contained the M2-2 gene deletion, viral RNA was extracted from virus and subjected to RT/PCR using a pair of primers spanning the M2-2 gene. Viral RNA was extracted from rA2ΔM2-2 and rA2 infected cell culture supernatant by an RNA extraction kit (RNA STAT-50, Tel-Test, Friendswood, TX). Viral RNA was reverse transcribed with reverse transcriptase using a primer complementary to viral genome from 7430 nt to 7449 nt. The cDNA fragment spanning the M2-2 gene was amplified by PCR with primer V1948 (7486 nt to 7515 nt at positive-sense) and primer V1581 (8544 nt to 8525 nt at negative sense). The PCR product was analyzed on a 1.2 % agarose gel and visualized by EtBr staining. As shown in Fig. 13B, wild type

rA2 yielded a PCR DNA product corresponding to the predicted 1029 nt fragment, whereas rA2ΔM2-2 yielded a PCR product of 795 nt, 234 nt shorter. Generation of RT/PCR product was dependent on the RT step, indicating that they were derived from RNA rather than from DNA contamination.

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11.1.2 RNA SYNTHESIS OF rA2ΔM2-2

mRNA expression from cells infected with rA2ΔM2-2 or rA2 was analyzed by Northern blot hybridization analyses. Total cellular RNA was extracted from rA2ΔM2-2 or rA2 infected cells by an RNA extraction kit (RNA STAT-60, Tel-Test, Friendswood, TX). RNA was electrophoresed on a 1.2% agarose gel containing formaldehyde and transferred to a nylon membrane (Amersham Pharmacia Biotech, Piscataway, NJ). The membrane was hybridized with a RSV gene specific riboprobe labeled with digoxigenin (Dig). The hybridized RNA bands were visualized by using Dig-Luminescent Detection Kit for Nucleic Acids (Boehringer Mannheim, Indianapolis, IN). Hybridization of the membranes with riboprobes was done at 65°C, membrane washing and signal detection were performed according to the standard procedures. To examine mRNA synthesis from rA2ΔM2-2 and rA2, accumulation of the M2 mRNA and the other viral mRNA products in infected Vero cells was analyzed by Northern blot hybridization. Hybridization of the blot with a probe specific to the M2-2 open reading frame did not detect any signal in rA2ΔM2-2 infected cells. Instead, a shorter M2 mRNA was detected in rA2ΔM2-2 infected cells using a riboprobe specific to the M2-1 gene (Fig. 14A). These observations confirmed that the M2-2 gene was deleted from rA2ΔM2-2. Accumulation of the other nine RSV mRNA transcripts was also examined and the amounts of each mRNA were found to be comparable between rA2ΔM2-2 and rA2 infected cells. Examples of Northern blots probed with N, SH, G and F are also shown in Fig. 14A. Slightly faster migration of F-M2 bicistronic mRNA was also discernible due to the deletion of the M2-2 region.

The M2-2 protein was previously reported to be a potent transcriptional negative regulator in a minigenome replication assay. However, deletion of the M2-2 gene from virus did not appear to affect viral mRNA production in infected cells. To determine if levels of viral antigenome and genome RNA of rA2ΔM2-2 were also similar to rA2, the amount of viral genomic and antigenomic RNA produced in infected Vero and Hep-2 cells was examined by Northern hybridization. Hybridization of the infected total cellular RNA with a ³²P-labeled F gene riboprobe specific to the negative genomic sense RNA indicated that much less genomic RNA was produced in cells infected with rA2ΔM2-2 compared to rA2 (Fig. 14B). A duplicate membrane was hybridized with a ³²P-labeled F gene riboprobe specific to the positive sense RNA. Very little antigenomic RNA was detected in cells infected with rA2ΔM2-2, although the amount of the F mRNA in rA2ΔM2-2 infected cells was comparable to rA2. Therefore, it appears that RSV genome and antigenome synthesis was down-regulated due to deletion of the M2-2 gene. This down-regulation was seen in both Vero and Hep-2 cells and thus was not cell type dependent.

11.1.3 PROTEIN SYNTHESIS OF rA2ΔM2-2

Since the putative M2-2 protein has not been identified in RSV infected cells previously, it was thus necessary to demonstrate that the M2-2 protein is indeed encoded by RSV and produced in infected cells. A polyclonal antiserum was produced against the M2-2 fusion protein that was expressed in a bacterial expression system. To produce antiserum against the M2-2 protein of RSV, a cDNA fragment encoding the M2-2 open reading frame from 8155nt to 8430nt was amplified by PCR and cloned into the pRSETA vector (Invitrogen, Carlsbad, CA). pRSETA/M2-2 was transformed into BL21-Gold(DE3)plysS cells (Stratagene, La Jolla, CA) and expression of His-tagged M2-2 protein was induced by IPTG. The M2-2 fusion protein was purified through HiTrap affinity columns (Amersham Pharmacia Biotech, Piscataway, NJ) and was used to

immunize rabbits. Two weeks after a booster immunization, rabbits were bled and the serum collected.

Viral specific proteins produced from infected cells were analyzed by immunoprecipitation of the infected cell extracts or by Western blotting. For immunoprecipitation analysis, the infected Vero cells were labeled with ³⁵S-promix (100 µCi/ml ³⁵S-Cys and ³⁵S-Met, Amersham, Arlington Heights, IL) at 14 hr to 18 hr postinfection. The labeled cell monolayers were lysed by RIPA buffer and the polypeptides immunoprecipitated by polyclonal anti-RSV A2 serum (Biogenesis, Sandown, NH) or anti-M2-2 serum. Immunoprecipitation of rA2 infected Vero cell lysates with anti-M2-2 antibody produced a protein band of approximately 10 kDa, which is the predicated size for the M2-2 polypeptide. This polypeptide was not detected in rA2ΔM2-2 infected cells (Fig.15A), confirming that M2-2 is a protein product produced by RSV and its expression was ablated from rA2ΔM2-2. The overall polypeptide pattern of rA2ΔM2-2 was indistinguishable from that of rA2. However, it was noted that slightly more P and SH proteins were produced in rA2ΔM2-2 infected Vero cells by immunoprecipitation. Nevertheless, by Western blotting analysis, a comparable amount of SH was produced in cells infected with rA2ΔM2-2 or rA2 (Fig.15B).

Immunoprecipitated polypeptides were electrophoresed on 17.5% polyacrylamide gels containing 0.1% SDS and 4 M urea, and detected by autoradiography. For Western blotting analysis, Hep-2 and Vero cells were infected with rA2ΔM2-2 or rA2. At various times postinfection, virus infected cells were lysed in protein lysis buffer and the cell lysates were electrophoresed on 17.5% polyacrylamide gels containing 0.1% SDS and 4 M urea. The proteins were transferred to a nylon membrane. Immunoblotting was performed as described in Jin *et al.* (Jin, H. *et al. Embo J* 16(6), 1236-47 (1997)), using polyclonal antiserum against M2-1, NS1, or SH.

Western blotting was used to determine the protein synthesis kinetics of rA2ΔM2-2 in both Vero and Hep-2 cell lines. Hep-2 or Vero cells were infected with rA2ΔM2-2 or rA2 at moi of 0.5 and at various times of postinfection, the

infected cells were harvested and the polypeptides separated on a 17.5% polyacrylamide gel containing 4 M urea. The proteins were transferred to a nylon membrane and probed with polyclonal antisera against the three accessory proteins: M2-1, NS1 and SH. Protein expression kinetics for all three viral proteins were very similar for rA2ΔM2-2 and rA2 in both Hep-2 and Vero cells (Fig.15B). Synthesis of the NS1 protein was detected at 10 hr postinfection, which was slightly earlier than M2-1 and SH because the NS1 protein is the first gene translated and is a very abundant protein product in infected cells. Similar protein synthesis kinetics was also observed when the membrane was probed with a polyclonal antiserum against RSV (data not shown). Comparable M2-1 was detected in rA2ΔM2-2 infected cells, indicating that deletion of the M2-2 open reading frame did not affect the level of the M2-1 protein that is translated by the same M2 mRNA.

11.1.4 GROWTH ANALYSIS OF RECOMBINANT RSV IN TISSUE CULTURE

To compare plaque morphology of rA2ΔM2-2 with rA2, Hep-2 or Vero cells were infected with each virus and overlaid with semisolid medium composed of 1% methylcellulose and 1 X L15 medium with 2% FBS. Five days after infection, infected cells were immunostained with antisera against RSV A2 strain. Plaque size was determined by measuring plaques from photographed microscopic images. Plaque formation of rA2ΔM2-2 in Hep-2 and Vero cells was compared with rA2. As shown in Fig. 16, rA2ΔM2-2 formed pin point sized plaques in Hep-2 cells, with a reduction of about 5-fold in virus plaque size observed for rA2ΔM2-2 compared to rA2. However, only a slight reduction in plaque size (30%) was seen in Vero cells infected with rA2ΔM2-2.

A growth kinetics study of rA2ΔM2-2 in comparison with rA2 was performed in both Hep-2 and Vero cells. Cells grown in 6-cm dishes were infected with rA2 or rA2ΔM2-2 at a moi of 0.5. After 1 hr adsorption at room

temperature, infected cells were washed three times with PBS, replaced with 4 ml of OPTI-MEM and incubated at 35°C incubator containing 5% CO₂. At various times post-infection, 200µl culture supernatant was collected, and stored at -70°C until virus titration. Each aliquot taken was replaced with an equal amount of fresh medium. Virus titer was determined by plaque assay in Vero cells on 12-well plates using an overlay of 1% methylcellulose and 1 X L15 medium containing 2% FBS. As seen in Fig. 17, rA2ΔM2-2 showed very slow growth kinetics and the peak titer of rA2ΔM2-2 was about 2.5-3 log lower than that of rA2 in Hep-2 cells. In Vero cells, rA2ΔM2-2 reached a peak titer similar to rA2. To analyze virus replication in different host cells, each cell line grown in 6-well plates was infected with rA2ΔM2-2 or rA2 at moi of 0.2. Three days postinfection, the culture supernatants were collected and virus was quantitated by plaque assay. rA2ΔM2-2 was examined for its growth properties in various cell lines that derived from different hosts with different tissue origins (Table 3). Significantly reduced replication of rA2ΔM2-2, two orders of magnitude less than rA2, was observed in infected Hep-2, MRC-5, and Hela cells, all of human origin. However, replication of rA2ΔM2-2 was only slightly reduced in MDBK and LLC-MK2 cells that are derived from bovine and rhesus monkey kidney cells, respectively.

Table 3. Replication levels of rA2 M2-2 and rA2 in various cell lines

Cell lines	Host	Tissue origin	Virus titer [log ₁₀ (pfu/ml)]	
			rA2	rA2ΔM2-2
Vero	Monkey	Kidney	6.1	6.1
Hep-2	Human	Larynx	6.2	4.3
MDBK	Bovine	Kidney	6.1	5.5
MRC-5	Human	Lung	5.5	3.0
Hela	Human	Cervix	6.6	4.5
LLC-MK2	Monkey	Kidney	6.7	6.1

11.1.5 REPLICATION OF rA2ΔM2-2

IN MICE AND COTTON RATS

Virus replication *in vivo* was determined in respiratory pathogen-free 12-week-old Balb/c mice (Simonsen Lab., Gilroy, CA) and *S. Hispidus* cotton rats (Virion Systems, Rockville, MD). Mice or cotton rats in groups of 6 were inoculated intranasally under light methoxyflurane anesthesia with 10^6 pfu per animal in a 0.1-ml inoculum of rA2 or rA2ΔM2-2. On day 4 postinoculation, animals were sacrificed by CO₂ asphyxiation and their nasal turbinates and lungs were obtained separately. Tissues were homogenized and virus titers were determined by plaque assay in Vero cells. To evaluate immunogenicity and protective efficacy, three groups of mice were inoculated intranasally with rA2, rA2ΔM2-2 or medium only at day 0. Three weeks later, mice were anesthetized, serum samples were collected, and a challenge inoculation of 10^6 pfu of biologically derived wild type RSV strain A2 was administered intranasally. Four days post-challenge, the animals were sacrificed and both nasal turbinates and lungs were harvested and virus titer determined by plaque assay. Serum antibodies against RSV A2 strain were determined by 60% plaque reduction assay (Coates, H.V. *et al.*, *AM. J. Epid.* 83:299-313 (1965)) and by immunostaining of RSV infected cells.

Table 4. Replication of rA2ΔM2-2 and rA2 in cotton rats

Virus	Virus titer (mean log ₁₀ pfu/g tissue ± SE) ^a	
	Nasal turbinates	Lung
rA2	4.0 ± 0.33	5.5 ± 0.12
rA2ΔM2-2	< 1.4	<1.4

^a Groups of six cotton rats were immunized intranasally with 10^6 pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

To evaluate levels of attenuation and immunogenicity of rA2ΔM2-2, replication of rA2ΔM2-2 in the upper and lower respiratory tract of mice and

cotton rats was examined. Cotton rats in groups of 6 were inoculated with 10^6 pfu of rA2ΔM2-2 or rA2 intranasally. Animals were sacrificed at 4 days postinoculation, their nasal turbinates and lung tissues were harvested, homogenized, and levels of virus replication in these tissues were determined by plaque assay. rA2ΔM2-2 exhibited at least 2 log reduction of replication in both nasal turbinates and lungs of the infected cotton rats (Table 4). No virus replication was detected in cotton rats infected with rA2ΔM2-2, whereas a high level of wild type rA2 virus replication was detected in both the upper and lower respiratory tract of cotton rats. Attenuation of rA2ΔM2-2 was also observed in mice. Geometric mean titer of virus replication and standard error obtained from two experiments are shown in Table 5. rA2ΔM2-2 replication was only detected in one or two of 12 infected mice. The replication was limited, only a few plaques were observed at 10^{-1} dilution of the tissue homogenates. Despite its restricted replication in mice, rA2ΔM2-2 induced significant resistance to challenge with wild type A2 RSV (Table 5). When mice previously inoculated with rA2ΔM2-2 or rA2 were inoculated intranasally with 10^6 pfu dose of wild type A2 strain, no wild type A2 virus replication was detected in the upper and lower respiratory tract of mice. Therefore, rA2ΔM2-2 was fully protective against wild type A2 virus challenge.

The immunogenicity of rA2ΔM2-2 was also examined. Two groups of mice were infected with rA2ΔM2-2 or rA2, and three weeks later, serum samples were collected. The serum neutralization titer was determined by 50% plaque reduction titer. The neutralization titer from rA2ΔM2-2 infected mice was comparable to that of rA2, both had 60% plaque reduction titer at 1:16 dilution. The serum obtained from rA2ΔM2-2 infected mice was also able to immunostain RSV plaques, confirming that RSV-specific antibodies were produced in rA2ΔM2-2 infected mice.

Table 5. Replication of rA2ΔM2-2 and rA2 in mice, and protection against wild type A2 RSV challenge

Immunizing Virus	Virus replication ^a		RSV A2 replication after challenge ^b	
	(Mean log ₁₀ pfu/g tissue ± SE)		(Mean log ₁₀ pfu/g tissue ± SE)	
	Nasal turbinates	Lung	Nasal turbinates	Lung
rA2	3.72 ± 0.33	4.0 ± 0.13	< 1.4	< 1.4
rA2ΔM2-2	< 1.4	< 1.4	< 1.4	< 1.4
Control	< 1.4	< 1.4	3.53 ± 0.17	4.10 ± 0.13

^a Groups of 12 Balb/c mice were immunized intranasally with 10⁶ pfu of the indicated virus on day 0. The level of infected virus in indicated tissues was determined by plaque assay at day 4, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

^b Groups of 6 Balb/c Mice were intranasally administered with 10⁶ pfu of RSV A2 on day 21 and sacrificed 4 days later. Replication of wild type RSV A2 in tissues as indicated was determined by plaques assay, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

The two RSV antigenic subgroups, A and B, exhibit a relatively high degree of conservation in M2-2 proteins, suggesting functional importance for the M2-2 protein. Transcriptional analysis for rA2 and rA2ΔM2-2 yielded important findings within the present example. Although overall mRNA transcriptional levels were substantially the same for both viruses, Northern blot analysis revealed dramatic reduction of virus genome and antigenome RNA for rA2ΔM2-2 compared to wild type rA2. This finding is contradictory with what has been reported for the negative transcriptional regulation of the M2-2 protein in a minigenome system. It thus appears that the functional role of M2-2 in the virus life cycle is more complicated than previously thought. Nevertheless, the reduction in the level of genome and antigenome of rA2ΔM2-2 did not appear to affect virus yields in infected Vero cells.

The finding that rA2ΔM2-2 exhibited host range restricted replication in different cell lines provided a good indication that deletion of a nonessential gene is a good means to create a host range mutant, which can be a very important feature for vaccine strains. rA2ΔM2-2 did not replicate well in

several cell lines that are derived from human origin, lower virus yield was produced from these cell lines. However, the levels of protein synthesis in Hep-2 cells were similar to Vero cells that produced high levels of rA2ΔM2-2. This indicated that the defect in virus release was probably due to a defect in a later stage, probably during the virus assembly process.

The finding that the M2-2 minus virus grows well in Vero cells and exhibits attenuation in the upper and lower respiratory tracts of mice and cotton rats presents novel advantages for vaccine development. The reduced replication in respiratory tracts of rodents did not affect immunogenicity and protection against challenging wild type virus replication, indicating that this M2-2 minus virus may serve as a good vaccine for human use. The nature of the M2-2 deletion mutation, involving a 234 nt deletion, represents a type of mutation that will be highly refractory to reversion.

11.2 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL SH GENE

This example describes production of a recombinant RSV in which expression of the SH gene has been ablated by removal of a polynucleotide sequence encoding the SH gene and its encoded protein. The RSV SH protein is encoded by the SH mRNA which is the 5th gene translated by RSV. The SH protein contains 64 amino acids in the strain A2 and contains a putative transmembrane domain at amino acid positions 14-41. The SH protein only has counterparts in simian virus 5 (Hiebert, S. W. *et al.* 5. *J Virol* 55(3), 744-51 (1985)) and mumps virus (Elango, N. *et al.* *J Virol* 63(3), 1413-5 (1989)). The function of the SH protein has not been defined. This example demonstrated that the entire SH gene can be removed from RSV. Thus, SH gene deletion may provide an additional method for attenuating RSV by itself or in combination with other gene deletions or mutations.

To produce a recombinant RSV having deletion in the RSV, the entire SH open reading frame was deleted from an infectious cDNA clone that derived from the RSV A2 strain. A two step cloning procedure was performed to delete the SH gene (from 4220 nt to 4477 nt) from a cDNA subclone. A *Sac I* restriction enzyme site was introduced at the gene start signal of the SH gene at position of 4220 nt. A unique *Sac I* site also exists at the C-terminal of the SH gene at position of 4477nt. Site-directed mutagenesis to introduce a *Sac I* site at the 5' of the SH gene was performed in pET(A/S) subclone, which contained *Avr II* (2129nt) to *Sac I* (4477nt) restriction fragment of RSV sequence. Digestion of pET(A/S) plasmid that contained the introduced *Sac I* site with *Sac I* restriction enzyme removed 258 nt fragment of the SH gene. pET(A/S) which had the SH gene deletion was digested with *Avr II* and *Sac I* and the released RSV restriction fragment was then cloned into a full length RSV cDNA clone. The full-length cDNA clone containing the SH gene deletion was designated pA2ΔSH.

Generation of pA2ΔSH mutant was performed as described above (see Section 7). SH-minus RSV (rA2ΔSH) was recovered from MVA-infected cells that had been co-transfected with pA2 ΔSH together with three plasmids that expressed the N, P and L proteins, respectively. Identification of the recovered rA2ΔSH was performed by RT/PCR using a pair of primers which flanked the SH gene. A cDNA band that is about 258 nucleotide shorter than the wild-type RSV (rA2) was detected in the rA2ΔSH infected cells. No PCR product was seen in the RT/PCR reaction that did not have reverse transcriptase in the RT reaction. This indicated that the PCR DNA was derived from viral RNA and is not artifact, and the virus obtained is truly SH-minus RSV.

To compare plaque morphology of rA2ΔSH with rA2, Hep-2 or Vero cells were infected with each virus and overlayed with semisolid medium composed of 1% methylcellulose and 1 X L15 medium with 2% FBS. Five days after infection, infected cells were immunostained with antisera against RSV A2

strain. The plaque size of rA2ΔSH is similar to that of rA2 in both Hep-2 and Vero cells.

To analyze virus replication in different cell lines that were derived from various hosts with different tissue origin, each cell line grown in 6-well plates was infected with rA2ΔSH or rA2 at moi of 0.2. Three days postinfection, the culture supernatants were collected and virus was quantitated by plaque assay. As shown in Table 6, replication of rA2ΔSH was very similar to rA2 in all the cell lines examined, indicating that the growth of SH-minus RSV was not substantially affected by host range effects.

Table 6. Growth comparison of rA2ΔSH and rA2 in different cell lines

Cell lines	Host	Tissue origin	Virus titer [$\log_{10}(\text{pfu/ml})$]	
			rA2	rA2ΔSH
Vero	Monkey	Kidney	5.8	5.7
Hep-2	Human	Larynx	6.5	6.1
MDBK	Bovine	Kidney	6.3	6.6
MRC-5	Human	Lung	5.5	5.3
Hela	Human	Cervix	6.5	6.0

Virus replication *in vivo* was determined in respiratory pathogen-free 12-week-old Balb/c mice (Simonsen Lab., Gilroy, CA). Mice in groups of 6 were inoculated intranasally under light methoxyflurane anesthesia with 10^6 pfu per animal in a 0.1-ml inoculum of rA2 or rA2ΔSH. On day 4 postinoculation, animals were sacrificed by CO₂ asphyxiation and their nasal turbinates and lungs were obtained separately. Tissues were homogenized and virus titers were determined by plaque assay in Vero cells. As shown in Table 7, level of rA2ΔSH replication in lower respiratory tract was only slightly lower than rA2, indicating that SH deletion alone may not be sufficient to attenuate RSV.

Table 7. Replication of rA2ΔSH and rA2 in mice

Virus	Virus titer in lung (mean log ₁₀ pfu/g tissue ± SE) ^a
rA2	3.75 ± 0.07
rA2ΔSH	3.21 ± 0.25

^a Groups of mice were immunized intranasally with 10⁶ pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

11.3 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS1 GENE

This example describes production of a recombinant RSV in which expression of the NS1 gene has been ablated by removal of a polynucleotide sequence encoding the NS1 gene and its encoded protein. The RSV NS1 is encoded by the 3' proximal NS1 gene in the 3' to 5' direction of the RSV gene map. The NS1 protein is a small 139-amino acid polypeptide and its mRNA is most abundant of the RSV mRNA. The function of the NS1 protein has not yet been clearly identified. In the reconstituted RSV minigenome system, the NS1 protein appeared to be a negative regulatory protein for both transcription and RNA replication of a RSV minigenome (Grosfeld, H. *et al.*, *J. Virol.* 69, 5677-5686 (1995)). The NS1 protein does not have a known counterpart in other paramyxoviruses and its function in virus replication is not known. This example demonstrated that the entire NS1 gene can be removed from RSV and NS1 deletion may provide an additional method for attenuating RSV or in combination with other RSV gene deletions or mutations.

To delete the NS1 gene from RSV, two restriction enzyme sites were inserted at positions of the NS1 gene start signal and downstream of the NS1 gene end signal. A two step cloning procedure was performed to delete the entire NS1 gene from RSV. A *Pst* I restriction enzyme site was introduced at position of 45 nt and at position of 577 nt of RSV sequence by site-directed mutagenesis. Mutagenesis was performed in pET(X/A) cDNA subclone, which contained the first 2128 nucleotides of RSV sequences that encode the NS1,

NS2 and part of the N gene of RSV. The 2128 nucleotide RSV sequence was cloned into the pET vector through the *Xma I* and *Avr II* restriction enzyme sites. Digestion of pET(X/A) plasmid that contained the introduced two *Pst I* restriction enzyme sites removed the 532 nucleotide fragment that contained the NS1 gene. The deletion included the NS1 gene start signal, the NS1 coding region, and the NS1 gene end signal. pET(X/A) which contained the NS1 deletion was digested with *Avr II* and *Sac I* and the released restriction fragment was then cloned into a full length RSV cDNA clone. The full-length RSV antigenomic cDNA clone containing the NS1 gene deletion was designated pA2ΔNS1.

Generation of pA2ΔNS1 mutant was performed as described above (see Section 7). NS1-minus RSV (rA2ΔNS1) was recovered from MVA-infected cells that had been co-transfected with pA2ΔNS1 together with three plasmids that expressed the N, P and L proteins, respectively. Recovery of infectious RSV was indicated by syncytial formation and confirmed by immunostaining with an antibody against RSV. Identification of the recovered rA2ΔNS1 was performed by RT/PCR using a pair of primers flanking the NS1 gene. A cDNA band that is about 532 nt shorter than the wild-type RSV (rA2) was detected in the rA2ΔNS1 infected cells. No PCR product was seen in the RT/PCR reaction that did not have reverse transcriptase in the RT reaction. This indicated that the PCR DNA was derived from viral RNA and is not artifact, and the virus obtained is truly NS1-minus RSV.

mRNA expression from cells infected with rA2ΔNS1 or rA2 was analyzed by Northern blot hybridization analyses. Total cellular RNA was extracted from rA2ΔNS1 or rA2 infected cells by an RNA extraction kit (RNA STAT-60, Tel-Test, Friendswood, TX). RNA was electrophoresed on a 1.2% agarose gel containing formaldehyde and transferred to a nylon membrane (Amersham Pharmacia Biotech, Piscataway, NJ). The membrane was hybridized with a riboprobe specific to the NS1, NS2 or M2-2 gene. As shown in Fig.18, no NS1 mRNA was detected in cells infected with rA2ΔNS1 using a probe that was

specific to the NS1 gene. The fact that the NS1 gene can be deleted from RSV identifies that the NS1 protein is an accessory protein product that is not essential for RSV replication. rA2ΔNS1 formed very small plaques in infected Hep-2 cells, but only slight plaque size reduction was seen in Vero cells (Fig.19). The small plaque phenotype is commonly associated with attenuating mutations.

A growth kinetics study of rA2ΔNS1 in comparison with rA2 was performed in Vero cells. Cells grown in 6-cm dishes were infected with rA2 or rA2ΔNS1 at a moi of 0.2. As seen in Fig. 20, rA2ΔNS1 showed very slow growth kinetics and its peak titer was about 1.5 log lower than that of rA2. To analyze virus replication in different host cells, each cell line grown in 6-well plates was infected with rA2ΔNS1 or rA2 at moi of 0.2. Three days postinfection, the culture supernatants were collected and virus was quantitated by plaque assay. rA2ΔNS1 had about 1-1.5 log reduction in virus titer compared to rA2 in Vero, Hep-2 and MDBK cells. About 2 log reduction in virus titer was observed in Hela and MRC5 cells (Table 8). Replication of rA2ΔNS1 in a small animal model is currently being investigated. Preliminary data indicated that rA2ΔNS1 is attenuated in cotton rats. The NS1 deletion mutant therefore provides an additional method for attenuating RSV.

Table 8. Growth comparison of rA2ΔNS1 and rA2 in different cell lines

Cell lines	Virus titer [$\log_{10}(\text{pfu/ml})$]	
	rA2	rA2ΔNS1
Vero	6.4	5.5
Hep-2	6.7	5.1
MDBK	6.7	5.2
MRC-5	5.9	3.6
Hela	6.5	4.5

11.4 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS2 GENE

This example describes production of a recombinant RSV in which expression of the NS2 gene has been ablated by removal of a polynucleotide sequence encoding the NS2 gene and its encoded protein. The NS2 is a small protein that is encoded by the second 3' proximal NS2 gene in the 3' to 5' order of RSV genome. The NS2 protein might be the second most abundant RSV protein of RSV, but its function remains to be identified.

To delete the NS2 gene from RSV, two restriction enzyme sites were inserted at positions of upstream of the NS2 gene start signal and downstream of the NS2 gene end signal. A two step cloning procedure was performed to delete the entire NS1 gene from RSV. A *Pst I* restriction enzyme site was introduced at position of 577 nt and at position of 1110 nt of RSV sequence by site-directed mutagenesis. Mutagenesis was performed in pET(X/A) cDNA subclone, which contained the first 2128 nt of RSV sequences at antigenomic sense that encode the NS1, NS2 and part of the N gene of RSV. The 2128 nt RSV sequences were cloned into the pET vector through the *Xma I* and *Avr II* restriction enzyme sites. Digestion of pET(X/A) plasmid that contained the introduced two *Pst I* restriction enzyme sites removed 533 nucleotide fragment of the NS2 gene. The 533 nt fragment contained the gene start signal of NS2, NS2 coding region and the gene end signal of NS2. pET(X/S) plasmid that contained the NS2 gene deletion was digested with *Avr II* and *Sac I* restriction enzymes and the released RSV restriction fragment was then cloned into a full length RSV cDNA clone. The full-length cDNA clone containing the NS2 gene deletion was designated pA2ΔNS2.

Generation of rA2ΔNS2 mutant was performed as described above (see Section 7). NS2-minus RSV (rA2ΔNS2) was recovered from MVA-infected cells that had been co-transfected with pA2ΔNS2 together with three plasmids that expressed the N, P and L proteins, respectively. Recovery of infectious RSV was indicated by syncytial formation and confirmed by immunostaining with an

antibody against RSV. Identification of the recovered rRSV Δ NS2 was performed by RT/PCR using a pair of primers that flanked the NS2 gene. A cDNA band that is about 533 nucleotide shorter than the wild-type RSV (rA2) was detected in the rA2 Δ NS2 infected cells. No PCR product was seen in the RT/PCR reaction that did not have reverse transcriptase in the RT reaction. This indicated that the PCR DNA was derived from viral RNA and is not artifact, and the virus obtained is truly NS2-minus RSV.

mRNA expression from cells infected with rA2 Δ NS2 or rA2 was analyzed by Northern blot hybridization analyses as described earlier. The blot was hybridized with a riboprobe specific to the NS1, NS2 or M2-2 gene. As shown in Fig.18, no NS2 mRNA was detected in cells infected with rA2 Δ NS2 using a probe that was specific to the NS2 gene. Comparable level of NS1 and M2 mRNA was detected in rA2 Δ NS2-infected cells. The fact that the NS2 gene can be deleted from RSV indicates that the NS2 protein is an accessory protein product that is not essential for RSV replication. rA2 Δ NS2 formed very small plaques in infected Hep-2 cells, but plaque size similar to rA2 was seen in rA2 Δ NS2 infected Vero cells (Fig.19). The small plaque phenotype is commonly associated with attenuating mutations.

A growth kinetics study of rA2 Δ NS2 in comparison with rA2 was performed in Vero cells. Cells grown in 6-cm dishes were infected with rA2 or rA2 Δ NS2 at a moi of 0.2. As seen in Fig. 21, rA2 Δ NS2 showed slower growth kinetics and its peak titer was about 5-fold lower than that of rA2. To analyze virus replication in different host cells, each cell line grown in 6-well plates was infected with rA2 Δ NS2 or rA2 at moi of 0.2. Three days postinfection, the culture supernatants were collected and virus was quantitated by plaque assay. rA2 Δ NS2 had only slight reduction in virus titer compared to rA2 in Vero cells. About a 1 log reduction in virus titer was observed in Hep-2, MDBK, Hela and MRC5 cells (Table 9). Replication of rA2 Δ NS2 in a small animal model is currently being investigated. rA2 Δ NS2 exhibited about 10-fold reduction of replication in the lower respiratory tract of cotton rats (Table 10). The NS2 deletion mutant therefore provides a method to obtain attenuated RSV.

Table 9. Growth comparison of rA2ΔNS2 and rA2 in different cell lines

Cell lines	Virus titer [$\log_{10}(\text{pfu/ml})$]	
	rA2	rA2 NS2
Vero	6.4	6.2
Hep-2	6.7	5.9
MDBK	6.7	5.2
MRC-5	5.9	4.7
Hela	6.5	5.5

Table 10. Replication of rA2ΔNS2 and rA2 in cotton rats

Virus	Virus titer in lung (mean \log_{10} pfu/g tissue \pm SE) ^a
rA2	3.93 ± 0.13
RA2ΔNS2	2.79 ± 0.47

^a Groups of five cotton rats were immunized intranasally with 10^5 pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on the indicated specimens, and the mean \log_{10} titer \pm standard error (SE) per gram tissue was determined.

11.5 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL M2-2 AND SH GENES

This example describes production of a recombinant RSV in which expression of two RSV genes, M2-2 and SH, has been ablated by removal of polynucleotide sequences encoding the M2-2 and SH genes and their encoded proteins. As described earlier, the M2-2 or SH gene is dispensable for RSV replication *in vitro*. It is possible that deletion of two accessory genes will produce a recombinant RSV with a different attenuation phenotype. The degree of attenuation from deletion of two genes can be increased or decreased.

SH and M2-2 genes were deleted from the full-length RSV cDNA construct through cDNA cloning. A *Sac I* to *BamH I* fragment that contained M2-2 deletion in the pET(S/B) subclone as described earlier was removed by digestion with *Sac I* and *BamH I* restriction enzymes and was cloned into the full-length RSV antigenomic cDNA clone that contained the SH gene deletion (pA2ΔSH). The resulting plasmid that contained deletion of SH and M2-2 was designated pA2ΔSHΔM2-2. Deletion of SH and M2-2 in pA2ΔSHΔM2-2 plasmid was confirmed by restriction enzyme mapping.

Generation of rA2ΔSHΔM2-2 mutant was performed as described above (see Section 7). Recombinant RSV that contained a deletion of the SH and M2-2 genes (rA2ΔSHΔM2-2) was recovered from MVA-infected cells that had been co-transfected with pA2ΔSHΔM2-2 together with three plasmids that expressed the N, P and L proteins, respectively. Recovery of infectious RSV deletion mutant was indicated by syncytial formation and confirmed by immunostaining with an antibody against RSV.

Deletion of the SH and M2-2 genes in rA2ΔSHΔM2-2 was confirmed by RT/PCR using two sets of primers that flanked the SH gene and the M2-2 gene, respectively. mRNA expression from cells infected with rA2ΔSHΔM2-2 or rA2 was analyzed by Northern blot hybridization analyses as described earlier. Both SH and M2-2 mRNAs were not detected in cells infected with rA2ΔSHΔM2-2 using a probe that was specific to the SH gene or M2-2 gene. The fact that two RSV genes (SH and M2-2) can be deleted from RSV indicates that the SH and M2-2 proteins are dispensable for RSV replication. In contrast to rA2ΔM2-2 that formed very small plaques in Hep-2 cells, rA2ΔSHΔM2-2 had a plaque size larger than rA2ΔM2-2 (Fig.19).

A growth kinetics study of rA2ΔSHΔM2-2 in comparison with rA2 was performed in Vero cells. Cells grown in 6-cm dishes were infected with rA2 or rA2ΔSHΔM2-2 at a moi of 0.2. As seen in Fig. 22, rA2ΔSHΔM2-2 showed slower growth kinetics and its peak titer was about 1.5 log lower than that of rA2. This indicated that rA2ΔSHΔM2-2 is attenuated in tissue culture.

To evaluate the level of attenuation of rA2ΔSHΔM2-2, replication of rA2ΔSHΔM2-2 in the lower respiratory tracts of mice was examined. Mice in groups of 6 were inoculated with 10⁶ pfu of rA2ΔSHΔM2-2 or rA2 intranasally. Animals were sacrificed at 4 days postinoculation, their nasal turbinates and lung tissues were harvested, homogenized, and levels of virus replication in these tissues were determined by plaque assay. rA2ΔSHΔM2-2 exhibited about a 2 log reduction of replication in lungs of the infected mice (Table 11). This data indicated that rA2ΔSHΔM2-2 is attenuated in mice, although the degree of attenuation is not as significant as rA2ΔM2-2.

Table 11. Replication of rA2ΔSHΔM2-2 and rA2 in mice

Virus	Virus titer in lung (mean log ₁₀ pfu/g tissue ± SE) ^a
rA2	4.2 ± 0.08
rA2ΔSHΔM2-2	2.4 ± 1.2

^a Groups of six mice were immunized intranasally with 10⁶ pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

11.6 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL M2-2 AND NS1 GENES

This example describes production of a recombinant RSV in which expression of two different RSV genes, NS1 and M2-2, has been ablated by removal of polynucleotide sequences encoding the NS1 and M2-2 genes and their encoded proteins. As described earlier, NS1 and M2-2 gene alone is dispensable for RSV replication *in vitro*. This example provided a different attenuating method by deletion of two accessory genes from RSV.

NS1 and M2-2 genes were deleted from the full-length RSV cDNA construct through cDNA cloning. A *Xma I* to *Avr II* fragment that contained NS1 deletion in pET(X/A) subclone was removed by digestion with *Xma I* and

Avr II restriction enzymes and was cloned into the full-length RSV antigenomic cDNA clone that contained the M2-2 gene deletion (pA2ΔM2-2). The resulting plasmid that contained deletion of both NS1 and M2-2 was designated pA2ΔNS1 ΔM2-2. Deletion of NS1 and M2-2 in pA2ΔNS1ΔM2-2 plasmid was confirmed by restriction enzyme mapping.

Generation rA2ΔNS1ΔM2-2 mutant was performed as described above (see section 11.2). Recombinant RSV that contained deletion of NS1 and M2-2 genes was recovered from MVA-infected cells that had been co-transfected with pA2ΔNS1ΔM2-2 together with three plasmids that expressed the N, P and L proteins, respectively. Recovery of infectious RSV was indicated by syncytial formation and confirmed by immunostaining with an antibody against RSV. Identification of the recovered rA2ΔNS1ΔM2-2 was confirmed by RT/PCR using a pair of primers flanking the NS1 gene and the M2-2 gene.

Replication of rA2ΔNS1ΔM2-2 in tissue culture cell lines and in small animal models is being studied. Preliminary *in vitro* data indicated that rA2ΔNS1ΔM2-2 is very attenuated in tissue culture cells and recombinant RSV containing deletion of NS1 and M2-2 genes is more attenuated than rA2ΔSHΔM2-2.

11.7 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS2 AND M2-2 GENES

This example describes production of a recombinant RSV in which expression of two different RSV genes, NS2 and M2-2, has been ablated by removal of polynucleotide sequences encoding the NS2 and M2-2 genes and their encoded proteins. As described earlier, NS2 or M2-2 gene is dispensable for RSV replication *in vitro*. It is possible that deletion of two accessory genes from RSV will produce a recombinant RSV with a different attenuation phenotype.

NS2 and M2-2 genes were deleted from the full-length RSV cDNA construct through cDNA cloning. A *Xma I* to *Avr II* fragment that contained NS2 deletion in pET(X/A) subclone was removed by digestion with *Xma I* and *Avr II* restriction enzymes and was cloned into the full-length RSV antigenomic cDNA clone that contained the M2-2 gene deletion (pA2ΔM2-2). The resulting plasmid that contained deletion of both NS2 and M2-2 was designated pA2ΔNS2 ΔM2-2. Deletion of NS2 and M2-2 in pA2ΔNS2ΔM2-2 plasmid was confirmed by restriction enzyme mapping.

Generation of rA2ΔNS2ΔM2-2 mutant was performed as described above (see Section 7). Recombinant RSV that contained deletion in the NS2 and M2-2 genes (rA2ΔNS2ΔM2-2) was recovered from MVA-infected cells that had been co-transfected with pA2ΔNS2ΔM2-2 together with three plasmids that expressed the N, P and L proteins, respectively. Recovery of infectious RSV was indicated by syncytial formation and confirmed by immunostaining with an antibody against RSV. Identification of the recovered rA2ΔNS2ΔM2-2 was confirmed by RT/PCR using two pairs of primers flanking the NS2 or M2-2 gene, respectively.

mRNA expression from cells infected with rA2ΔNS2ΔM2-2 or rA2 was analyzed by Northern blot hybridization analyses. As shown in Fig. 23, neither NS2 nor M2-2 mRNA was detected in cells infected with rA2ΔNS2ΔM2-2 using a probe that was specific to the NS2 gene or to the M2-2 gene. Comparable levels of NS1 and SH mRNA expression was observed in cells infected with rA2ΔNS2ΔM2-2. Northern blot data confirmed that expression of both NS2 and M2-2 was ablated in rA2ΔNS2ΔM2-2.

A growth kinetics study of rA2ΔNS2ΔM2-2 in comparison with rA2 was performed in Vero cells. Cells grown in 6-cm dishes were infected with rA2 or rA2ΔNS2ΔM2-2 at a moi of 0.2. As seen in Fig. 24, rA2ΔNS2ΔM2-2 showed very slow growth kinetics and its peak titer was about 10-fold lower than that of rA2. To analyze virus replication in different host cells, each cell line grown in 6-well plates was infected with rA2ΔNS2ΔM2-2 or rA2 at moi of 0.2. Three days postinfection, the culture supernatants were collected and virus was

quantitated by plaque assay. rA2ΔNS2ΔM2-2 had about a few fold reduction in virus titer compared to rA2 in Vero cells. However, a 2-3 log reduction in virus titer was observed in Hep-2, MDBK, Hela, MRC5 and LLC-MK2 cells (Table 12). Therefore, replication of rA2ΔNS2ΔM2-2 exhibits a substantial host range effect, which is an indication of attenuation.

Table 12. Growth comparison of rA2ΔNS2/M2-2 and rA2 in different cell lines

Cell lines	Virus titer [$\log_{10}(\text{pfu/ml})$]	
	rA2	rA2ΔNS2/M2-2
Vero	6.4	5.7
Hep-2	6.7	3.5
MDBK	6.7	3.7
MRC-5	5.9	2.0
Hela	6.5	2.9
LLC-MK2	6.7	4.8

Replication of rA2ΔNS2/M2-2 *in vivo* was determined in respiratory pathogen-free 4-week old cotton rats. Cotton rats in groups of 5 were inoculated intranasally under light methoxyflurane anesthesia with 10^5 pfu per animal in a 0.1-ml inoculum of rA2 or rA2 ΔNS2ΔM2-2. On day 4 postinoculation, animals were sacrificed by CO₂ asphyxiation and their nasal turbinates and lungs were obtained separately. Tissues were homogenized and virus titers were determined by plaque assay in Vero cells. As shown in Table 13, no virus replication was detected in the upper and lower respiratory tracts of cotton rats that were infected with rA2 ΔNS2ΔM2-2. This indicated that deletion of the NS2 and M2-2 genes severely attenuated RSV. Thus, this recombinant RSV with an NS2 and M2-2 deletion might serve as a good vaccine candidate for human use.

Table 13. Replication of rA2ΔNS2/M2-2 and rA2 in cotton rats

Virus	Virus titer (mean log ₁₀ pfu/g tissue ± SE)	
	Nasal turbinates	Lung
rA2	2.30 ± 0.26	4.23 ± 0.10
rA2ΔNS2/M2-2	<1.4	<1.4

^a Groups of five cotton rats were immunized intranasally with 10⁵ pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

11.8 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS1 AND NS2 GENES

This example describes production of a recombinant RSV in which expression of two RSV genes, NS1 and NS2, has been ablated by removal of polynucleotide sequences encoding the NS1 and NS2 genes and their encoded proteins. As described earlier, NS1 or NS2 gene is dispensable for RSV replication *in vitro*. It is possible that deletion of two accessory genes from RSV will produce a recombinant RSV with alternative attenuation phenotype.

To delete the NS1 and NS2 gene from RSV, two restriction enzyme sites were inserted at positions of the gene start signal of NS1 and downstream of the gene end signal of NS2. A two step cloning procedure was performed to delete the entire NS1 and NS2 genes from RSV. A *Pst I* restriction enzyme site was introduced at position of 45 nt and at position of 1110 nt of RSV sequence by site-directed mutagenesis. Site-directed mutagenesis was performed in pET(X/A) cDNA subclone, which contained the first 2128 nucleotides of RSV sequence that encode the NS1, NS2 and part of the N gene of RSV. The 2128 nucleotide RSV cDNA fragment was cloned into the pET vector through the *Xma I* and *Avr II* restriction sites. Digestion of pET(X/A) plasmid that contained the introduced two *Pst I* restriction enzyme sites removed a 1065-nt fragment that included the NS1 and NS2 genes. pET(X/S) plasmid containing NS1 and NS2 deletion was digested with *Avr II* and *Sac I* restriction enzymes and the

remaining 1063 nucleotide RSV cDNA fragment was then cloned into a full length RSV antigenomic cDNA clone. The resulting plasmid that contained deletion of both NS1 and NS2 was designated pA2 Δ NS1 Δ NS2. Deletion of NS1 and NS2 in pA2 Δ NS1 Δ NS2 plasmid was confirmed by restriction enzyme mapping.

Recovery of infectious RSV that contained both NS1 and NS2 deletion (rA2 Δ NS1 Δ NS2) was performed as described earlier. Infectious virus with both NS1 and NS2 deleted was obtained from transfected Hep-2 cells. RT/PCR was performed to confirm that both NS1 and NS2 genes were deleted from rA2 Δ NS1 Δ NS2 using a pair of primers flanking the NS1 and NS2 genes. Deletion of NS1 and NS2 from rA2 Δ NS1 Δ NS2 was further confirmed by Northern blot. As shown in Fig. 18, neither NS1 nor NS2 mRNAs was detected in cells infected with rA2 Δ NS1 Δ NS2 using a riboprobe specific to the NS1 or NS2 gene. This indicated that expression of NS1 and NS2 was ablated from rA2 Δ NS1 Δ NS2.

rA2 Δ NS1 Δ NS2 formed very small plaques in infected Hep-2 cells, but only slight plaque size reduction was seen in Vero cells (Fig.19). The small plaque phenotype is commonly associated with attenuating mutations.

A growth kinetics study of rA2 Δ NS1 Δ NS2 in comparison with rA2 was performed in Vero cells. Cells grown in 6-cm dishes were infected with rA2 or rA2 Δ NS1 Δ NS2 at a moi of 0.2. As seen in Fig. 25, rA2 Δ NS1 Δ NS2 exhibited slower growth kinetics and its peak titer was about 5-fold lower than that of rA2. To analyze virus replication in different host cells, each cell line grown in 6-well plates was infected with rA2 Δ NS1 Δ NS2 or rA2 at moi of 0.2. Three days postinfection, the culture supernatants were collected and virus was quantitated by plaque assay. rA2 Δ NS1 Δ NS2 had only slight reduction in virus titer compared to rA2 in Vero cells. About 1.5 log reduction in virus titer was observed in Hep-2, MDBK and LLC-MK2 cells. More reduction in virus (about 3 log) was seen in Hela and MRC5 cells (Table 14). Replication of rA2 Δ NS1 Δ NS2 in a small animal model is currently being investigated. Preliminary data indicated that rA2 Δ NS1 Δ NS2 is attenuated in cotton rats. As replication of

rA2 ΔNS1ΔNS2 was not detected in cotton rats, it appears that the rA2ΔNS1ΔNS2 deletion mutant is very attenuated. The NS1 and NS2 deletion mutant therefore provides an alternative method for attenuating RSV.

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Table 14. Growth comparison of rA2ΔNS1ΔNS2 and rA2 in different cell lines

Cell lines	Virus titer [$\log_{10}(\text{pfu/ml})$]	
	rA2	rA2ΔNS1ΔNS2
Vero	6.4	6.2
Hep-2	6.7	5.1
MDBK	6.7	5.2
MRC-5	5.9	3.1
Hela	6.5	3.8
LLC-MK2	6.7	5.1

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11.9 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS1 AND SH GENES

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This example describes production of a recombinant RSV in which expression of two different RSV genes, NS1 and SH, has been ablated by removal of polynucleotide sequences encoding the NS1 and SH genes and their encoded proteins. As described earlier, NS1 or SH genes is dispensable for RSV replication *in vitro*. It is possible that deletion of two accessory genes from RSV will produce a recombinant RSV with increased attenuation phenotype.

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NS1 and SH genes were deleted from the full-length RSV cDNA construct through cDNA cloning. A *Xma I* to *Avr II* fragment that contained NS1 deletion in pET(X/A) subclone was removed by digestion with *Xma I* and *Avr II* restriction enzymes and was cloned into the full-length RSV antigenomic cDNA clone that contained the SH gene deletion (pA2 SH). The resulting plasmid that contained deletion of both NS1 and SH was designated

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pA2ΔNS1ΔSH. Deletion of NS1 and SH in pA2ΔNS1ΔSH plasmid was confirmed by restriction enzyme mapping.

Recovery of infectious RSV that contained both NS1 and SH deletion (rA2ΔNS1ΔSH) was performed as described earlier. Infectious virus with both NS1 and SH deleted was obtained from transfected Hep-2 cells. Virus was plaque purified 3 times and amplified in Vero cells. Deletion of both the NS1 and SH genes in rA2ΔNS1ΔSH was confirmed by RT/PCR using two sets of primers that flanked the NS1 or SH gene, respectively. Northern blot of rA2ΔNS1ΔSH infected total cellular RNA was performed using a riboprobe specific to the NS1 or SH gene. As shown in Fig. 23, expression of NS1 and SH mRNA was ablated in cells infected with rA2ΔNS1ΔSH.

Replication of rA2ΔNS1ΔSH *in vitro* and *in vivo* is currently being studied. The fact that the rA2ΔNS1ΔSH virus can grow, albeit with reduced efficiency, indicates that the NS1 and SH genes are dispensable for RSV replication. This mutant will therefore likely serve as an additional potential recombinant RSV vaccine agent.

11.10 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS2 AND SH GENES

This example describes production of a recombinant RSV in which expression of two different RSV genes, NS2 and SH, has been ablated by removal of polynucleotide sequences encoding the NS2 and SH genes and their encoded proteins. As described earlier, NS2 or SH gene is dispensable for RSV replication *in vitro*. It is possible that deletion of two accessory genes from RSV will produce a recombinant RSV with different attenuation phenotype.

NS2 and SH genes were deleted from the full-length RSV cDNA construct through cDNA cloning. A *Xma I* to *Avr II* fragment that contained NS2 deletion in pET(X/A) subclone was removed by digestion with *Xma I* and *Avr II* restriction enzymes and was cloned into the full-length RSV antigenomic cDNA clone that contained the SH gene deletion (pA2ΔSH). The resulting plasmid that contained deletion of both NS2 and SH was designated

pA2ΔNS2ΔSH. Deletion of NS2 and SH in pA2ΔNS2ΔSH plasmid was confirmed by restriction enzyme mapping.

Recovery of infectious RSV that contained both NS2 and SH deletion (rA2ΔNS2ΔSH) was performed as described earlier. Infectious virus with both NS2 and SH deleted was obtained from transfected Hep-2 cells. Virus was plaque purified 3 times and amplified in Vero cells. Deletion of both NS2 and SH gene in rA2ΔNS2ΔSH was confirmed by RT/PCR using two sets of primers that flanked the NS2 or SH gene, respectively. Northern blot of rA2ΔNS2ΔSH infected total cellular RNA was performed using a riboprobe specific to the NS2 or SH gene. As shown in Fig. 23, expression of NS2 and SH mRNA was ablated in cells infected with rA2 ΔNS2ΔSH.

Replication of rA2ΔNS2ΔSH *in vivo* was determined in respiratory pathogen-free 4-week old cotton rats. Cotton rats in groups of 5 were inoculated intranasally under light methoxyflurane anesthesia with 10⁵ pfu per animal in a 0.1-ml inoculum of rA2 or rA2ΔNS2ΔSH. On day 4 postinoculation, animals were sacrificed by CO₂ asphyxiation and their nasal turbinates and lungs were obtained separately. Tissues were homogenized and virus titers were determined by plaque assay in Vero cells. As shown in Table 15, reduced virus replication was observed in the upper and lower respiratory tracts of cotton rats that were infected with rA2ΔNS2ΔSH. This indicated that deletion of the NS2 and SH genes attenuated RSV and this recombinant RSV with NS2 and SH deletion might serve as a good vaccine candidate for human use.

Table 15. Replication of rA2ΔNS2ΔSH and rA2 in cotton rats

Virus	Virus titer (mean log ₁₀ pfu/g tissue ± SE)	
	Nasal turbinats	Lung
rA2	2.30 ± 0.26	4.23 ± 0.10
rA2ΔNS2ΔSH	1.11 ± 1.34	2.76 ± 0.06

^a Groups of five cotton rats were immunized intranasally with 10⁶ pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on

indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

11.11 GENERATION OF A HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE (RSV) CANDIDATE BY DELETING THE VIRAL NS1, NS2, AND SH GENES

This example describes production of a recombinant RSV in which expression of three RSV genes, NS1, NS2 and SH, has been ablated by removal of polynucleotide sequences encoding three RSV genes (NS1, NS2 and SH) and their encoded proteins. As described earlier, NS1, NS2 or SH alone is dispensable for RSV replication *in vitro*. It is possible that deletion of three accessory genes from RSV will produce a recombinant RSV with a different attenuation phenotype.

NS1, NS2 and SH genes were deleted from the full-length RSV cDNA construct through cDNA cloning. A *Xma I* to *Avr II* fragment that contained NS1 and NS2 deletion in pET(X/A) subclone as described earlier was removed by digestion with *Xma I* and *Avr II* restriction enzymes and was cloned into the full-length RSV antigenomic cDNA clone that contained the SH gene deletion (pA2ΔSH). The resulting plasmid that contained deletion of three genes (NS1, NS2 and SH) was designated pA2ΔNS1ΔNS2ΔSH. Deletion of NS1, NS2 and SH in pA2ΔNS1ΔNS2ΔSH plasmid was confirmed by restriction enzyme mapping.

Recovery of infectious RSV that contained three genes deletion (NS1, NS2 and SH), rA2ΔNS1ΔNS2ΔSH, was performed as described earlier. Infectious virus was obtained from transfected Hep-2 cells. Virus was plaque purified 3 times and amplified in Vero cells. Deletion of NS1, NS2 and SH genes in rA2ΔNS1ΔNS2ΔSH was confirmed by RT/PCR using two sets of primers that flanked the NS1 and NS2 genes or the SH gene, respectively. Northern blot of infected total cellular RNA of rA2ΔNS1ΔNS2ΔSH was performed using a riboprobe specific to the NS1, NS2 or SH gene. As shown in Fig. 23, expression of NS1, NS2 and SH mRNA was ablated in cells infected

with rA2ΔNS1ΔNS2ΔSH. This indicated that these three genes were indeed deleted from RSV.

Replication of rA2ΔNS1ΔNS2ΔSH *in vivo* was determined in respiratory pathogen-free 4-week old cotton rats. Cotton rats in groups of 5 were inoculated intranasally under light methoxyflurane anesthesia with 10⁵ pfu per animal in a 0.1-ml inoculum of rA2 or rA2ΔNS1ΔNS2ΔSH. On day 4 postinoculation, animals were sacrificed by CO₂ asphyxiation and their nasal turbinates and lungs were obtained separately. Tissues were homogenized and virus titers were determined by plaque assay in Vero cells. As shown in Table 16, no virus replication was observed in the upper and lower respiratory tracts of cotton rats that were infected with rA2ΔNS1ΔNS2ΔSH. This indicated that deletion of the NS2 and SH genes attenuated RSV and this recombinant RSV with NS2 and M2-2 deletion might serve as a good vaccine candidate for human use.

Table 16. Replication of rA2ΔNS1ΔNS2ΔSH and rA2 in cotton rats

Virus	Virus titer (mean log ₁₀ pfu/g tissue ± SE)	
	Nasal turbinates	Lung
rA2	2.30 ± 0.26	4.23 ± 0.10
rA2ΔNS1ΔNS2ΔSH	<1.4	<1.4

^a Groups of five cotton rats were immunized intranasally with 10⁵ pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue were determined.

In conclusion, 11 different gene deletion mutants have been obtained as summarized in Table 17. Four RSV accessory genes have been deleted either individually or in combination. These different deletion mutants showed different plaque formation and growth properties. A good correlation was demonstrated between plaque size *in vitro* and attenuation *in vivo*. These different RSV deletion mutants provide several choices for use as potential RSV vaccine candidates.

Table 17. Summary of RSV gene deletion mutants

	Virus	Recovered
5	Δ M2-2	Yes
	Δ SH	Yes
	Δ NS1	Yes
	Δ NS2	Yes
	Δ M2-2 Δ SH	Yes
10	Δ M2-2 Δ NS1	ND ^a
	Δ M2-2 Δ NS2	Yes
	Δ NS1 Δ NS2	Yes
	Δ SH Δ NS1	Yes
	Δ SH Δ NS2	Yes
15	Δ SH Δ NS1 Δ NS2	Yes
	^a ND Not determined. Replication of rA2 Δ M2-2 Δ NS1 was not detected in tissue culture.	

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12. **EXAMPLE: GENERATION OF AN ATTENUATED
HUMAN RESPIRATORY SYNCYTIAL VIRUS VACCINE
(RSV) CANDIDATE BY MUTAGENESIS OF THE VIRAL
M2-1 GENE**

5 Rationale:

The ability to generate infectious RSV from cDNA allows defined changes to be introduced into the RSV genome. The phenotype of the rescued viruses can be directly attributed to the engineered changes in the genome. Changes in the virus genome can be easily verified by sequencing the region in which mutations are introduced. Different point mutations and lesions can be combined in a single virus to create suitably attenuated and genetically stable RSV vaccine candidates. The RSV genome encodes several auxiliary proteins: NS1, NS2, SH, M2-1 and M2-2 proteins that do not have counterparts in other paramyxoviruses. The function of these genes in the viral life cycle is the subject of ongoing investigations.

The product of the M2-1 gene is a 22 kDa protein which has been shown to promote processive sequential transcription and antitermination of transcription at each gene junction of the RSV genome in vitro (Collins, P. L. *et al. Proc. Natl. Acad. Sci. U S A* 93, 81-85 (1996); Hardy, R. W. *et al. J. Virol.* 72, 520-526 (1998)). M2-1 is also thought to be a structural component of the viral nucleocapsid and interaction of M2-1 with the N protein has been observed in RSV infected cells (Garcia *et al. Virology* 195:243-247 (1993)). The M2-1 protein contains a putative zinc binding motif (Cys3His motif) at its N-terminus (Worthington *et al.*, 1996, *Proc. Natl. Acad. Sci.* 93:13754-13759). This motif is highly conserved throughout the pneumovirus genus.

Two mutagenesis strategies are presented here to introduce mutations in the M2-1 protein. The first method involves changing each of the cysteine residues individually to glycine (cysteine scanning mutagenesis). The second method involves engineering premature stop codons at the carboxyl terminus of the protein to produce truncated M2-1 proteins of various length. These

strategies provide different approaches to making attenuated RSV for use as live vaccines.

12.1 CYSTEINE SCANNING MUTAGENESIS OF M2-1

Four cysteine residues are present in the M2-1 protein at amino acid positions 7, 15, 21 and 96. Cys7, Cys15 and Cys21 lie in the putative zinc binding motif, the Cys3His motif. DNA oligonucleotides were designed to change these cysteine codons to that for glycine by Quickchange site-directed mutagenesis (Stratagene). Mutagenesis was performed using a cDNA subclone (pET-S/B) that contained nucleotide 4482 to nucleotide 8505 of the RSV genome. The oligonucleotides corresponding to the positive sense of the RSV genome used for the mutagenesis reactions are listed in Table 18.

The engineered changes in the pET-S/B RSV subclone were verified by DNA sequence analysis. Each Sac I to Bam HI restriction fragment that contained the mutated cysteine codon in M2-1 was individually cloned into an infectious RSV antigenomic cDNA clone that was derived from RSV strain A2 (Jin, H. *et al. Virology* 251, 206-214 (1998)). The full-length RSV antigenomic cDNA clone with an engineered cysteine to glycine codon change was designated pA2MC1, 2, 3, or 4.

Table 18. Primers used for changing each cysteine codon in the M2-1 gene^a

Primer	Position in RSV antigenome	Sequence
MC1	nt 7609-7641	5'TCACGAAGGAATCCT <u>G</u> GCAAATTTGAAATT CGA
MC2	nt 7633-7665	5'GAAATTCGAGGTCAT <u>G</u> GTTTAAATGGTAA GAGG
MC3	nt 7648-7683	5'TGCTTAAATGGTAAGAGG <u>G</u> GACATTTTAGT CATAAT
MC4	nt 7876-7908	5'ACTAAACAATCAGCAG <u>G</u> GTGTTGCCATGAG CAAA

^a The numbers correspond to nucleotides in the RSV antigenome. Nucleotides that were mutated to change cysteine codons to glycine codons are in bold and underlined.

To produce infectious RSV that contained an individual cysteine mutation in M2-1, pA2MC1, 2, 3, or 4 was transfected into cells that expressed the T7 RNA polymerase together with plasmids that expressed the N, P and L protein. Briefly, monolayers of Hep-2 cells in 6 well dishes at a confluency of 70–80% were infected with modified vaccinia virus that expressed the T7 RNA polymerase (MVA) at a moi of 5. Absorption of MVA was performed at room temperature for 1 hour. The infected cells were washed with OPTI-MEM (Life Technologies) and transfected with pA2MC1, pA2MC2, pA2MC3 or pA2MC4 antigenomic plasmids together with a mixture of plasmids encoding the RSV N, P and L proteins each under the control of the T7 promoter. The amount of plasmids used for each transfection are: 0.5 µg antigenome plasmid, 0.4 µg N plasmid, 0.4 µg P plasmid and 0.2 µg L plasmid in a final volume of 0.1 ml OPTI-MEM. The final plasmid mixture was combined with 0.1 ml OPTI-MEM containing 10 µl lipofecTACE (Life Technologies). After 15 minutes incubation at room temperature, the transfection mixture was added to the MVA infected cells. The transfection reaction was incubation at 33°C for 5 hours. After 5 hours, the transfection medium was removed and replaced with MEM supplemented with 2% fetal bovine serum and incubated at 33°C for 3 days. Following the 3-day incubation, medium was harvested and passaged in Vero cells for 6 days. Positive immunostaining of the infected cell monolayers using goat anti-RSV antibody (Biogenesis) was then used to identify wells containing successfully rescued viruses. RT/PCR of genomic viral RNA was performed to verify that the engineered changes were present in the rescued viruses. A recombinant RSV bearing the introduced cysteine change at position of 96, rA2C4 was obtained. Replication *in vitro* and in an animal model of rA2C4 is currently being studied. Preliminary results indicated that rA2C4 showed reduced plaque size at 35°C and is therefore probably attenuated. Preliminary results indicated that rA2C4 has about a 10-fold reduction in replication of the lungs of cotton rats (See Table 19). Recovery of rA2C1, rA2C2 and rA2C3 are

currently being pursued. It is quite possible that changes in any of the three cysteine residues in the putative zinc binding motif may prove to be lethal to the M2-1 protein.

Table 19. Replication of M2-1 mutants in cotton rats

Virus	Virus titer (mean log ₁₀ pfu/g tissue ± SE)
	Lung
rA2	3.55 ± 0.07
RA2C4	2.29 ± 0.13
rA2MSCH3	1.97 ± 0.18

^a Groups of five cotton rats were immunized intranasally with 10⁵ pfu of the indicated virus on day 0. The level of infected virus replication at day 4 was determined by plaque assay on the indicated specimens, and the mean log₁₀ titer ± standard error (SE) per gram tissue was determined.

12.2 C-TERMINAL TRUNCATIONS OF THE M2-1 PROTEIN

Tandem termination codons were introduced at the C-terminus of the M2-1 protein by site-directed mutagenesis in order to create progressively longer truncations from the C-terminal end of the M2-1 protein. Mutagenesis was performed using a cDNA subclone (pET-S/B) that contained RSV sequences from nucleotide 4482 to nucleotide 8505. Oligonucleotides corresponding to the positive sense of the RSV genome that were used for creating premature tandem termination codons in M2-1 are listed in Table 20.

The engineered changes were verified by sequence analysis of the RSV subclone containing the introduced mutations. The Sac I to Bam HI restriction fragment containing the premature tandem termination codons in M2-1 was excised from RSV subclone pET-S/B and introduced into the full length infectious RSV antigenomic cDNA clone (Jin et al.,1998). Each reassembled full-length RSV antigenomic cDNA containing the engineered premature

tandem termination codons along with a unique Hind III site was designated pA2MCSCH1, pA2MSCH 2 or pA2MSCH3.

Table 20. Primers used to introduce tandem termination codons in the C-terminus of the M2-1 protein

Primer	Position in RSV antigenome	Sequence ^a
MSCH 1	nt 7960-8011	5'GAGCTAAATTCACCCAAGATA <i>AAGCTT</i> GTA <i>TAA</i> ACTGTCATATCATATATTG
MSCH2	nt 8035-8076	5' CAAACTATCCATCTGT <i>AATAA</i> <i>GCTT</i> GCCAGCA GACGTATTG
MSCH3	nt 8120-8169	5' CCATCAACAACCCAAA <i>ATAATAA</i> <i>GCTT</i> TAGTG ATACAAATGACCATGCC

^aThe numbers correspond to nucleotides in the RSV antigenome. Tandem stop codons are indicated in bold. Mutated nucleotides are underlined and unique Hind III sites introduced simultaneously with the tandem stop codon are shown in italics.

Recombinant RSV that contained deletion in the C-terminal of the M2-1 protein was generated by transfection of pA2MCSCH1, pA2MSCH2 or pA2MSCH3 together with plasmids expressing the N, P and L proteins as described above. Recovery of infectious RSV that contained the shortest deletion in the C-terminus of the M2-1 protein, derived from pA2MSCH3 has been obtained. This virus had a 17 amino acid truncation at the C-terminus of M2-1 because of the engineered two tandem stop codons at amino acid 178 and 179. Virus plaque purification, amplification and verification of the engineered tandem termination codons in rA2MSCH3 are currently being performed . The rescue of recombinant RSV containing longer deletions in the C-terminus of the M2-1 protein is also being pursued. Preliminary results indicate that rA2MSCH3 has about a 15-fold reduction in replication of the lungs of cotton rats (See Table 19). Viable M2-1 deletion mutants provide an alternative method to attenuating RSV by itself or in combination with other mutations in the RSV genome for vaccine use.

13. **EXAMPLE: CHIMERIC SUBGROUP A RESPIRATORY SYNCYTIAL VIRUS (RSV) WITH THE GLYCOPROTEINS OF SUBGROUP B AND RSV WITHOUT THE M2-2 GENE ARE ATTENUATED IN AFRICAN GREEN MONKEYS**

13.1 INTRODUCTION

In this study, rA2ΔM2-2 was evaluated for its attenuation, immunogenicity, and protective efficacy against subsequent wild type RSV challenge in African green monkeys. The replication of rA2ΔM2-2 was more than 1000-fold restricted in both the upper and lower respiratory tracts of the infected monkeys and it induced titers of serum anti-RSV neutralizing antibody that were slightly lower than those induced by wild type rA2. When rA2ΔM2-2-infected monkeys were challenged with wild type A2 virus, the replication of the challenge virus was reduced by approximately 100-fold in the upper respiratory tract and 45,000-fold in the lower respiratory tracts. To further attenuate rA-G_BF_B, the M2-2 open reading frame was removed from rA-G_BF_B. As described for rA2ΔM2-2, rA-G_BF_BΔM2-2 was restricted for growth in Hep-2 cells and was attenuated in cotton rats. rA2 and rA-G_BF_B bearing a deletion of the M2-2 gene could represent a bivalent RSV vaccine composition for protection against multiple strains from the two RSV subgroups.

African green monkeys (AGM) were evaluated as a non-human primate model for assessing the attenuation, immunogenicity and protective efficacy of RSV vaccine candidates. We showed that rA2 replicated to high titers in both the upper and lower respiratory tracts of AGM, whereas rA2ΔM2-2 and rA-G_BF_B replicated poorly in the respiratory tracts of monkeys. Both rA2ΔM2-2 and rA-G_BF_B induced neutralizing antibodies which protected the animals from experimental challenge.

13.2 MATERIALS AND METHODS

Cells and viruses

Monolayer cultures of HEp-2 and Vero cells (obtained from American Type Culture Collections, ATCC) were maintained in minimal essential medium

(MEM) containing 5% fetal bovine serum (FBS). Wild type RSV strains, A2 and B9320, were obtained from ATCC and grown in Vero cells. Modified vaccinia virus Ankara (MVA-T7) expressing bacteriophage T7 RNA polymerase was grown in CEK cells.

Construction of chimeric cDNA clone

The wild type RSV B9320 was grown in Vero cells and the viral RNA was extracted from infected cell culture supernatant. A cDNA fragment containing the G and F genes of RSV B9320 was obtained by RT/PCR using the following primers:

ATCAGGATCCACAATAACATTGGGGCAAATGCAACC and
CTGGCATTTCGGATCCGTTTTATGTAAGTATGAGTTG (the BamH I sites engineered for cloning is in italics and B9320 specific sequences are underlined). BamH I restriction enzyme sites were introduced upstream of the gene start sequence of G and downstream of the gene end sequence of F. The PCR product was first introduced into the T/A cloning vector (Invitrogen) and the sequences were confirmed by DNA sequencing. The BamH I restriction fragment containing the G and F gene cassette of B9320 was then transferred into a RSV cDNA subclone pRSV(R/H) that contained RSV sequences from nt 4326 to nt 9721 through the introduced Bgl II sites at nt 4655 (upstream of the gene start signal of G) and at nt 7552 (downstream of the gene end signal of F). Introduction of these two Bgl II sites were made by PCR mutagenesis using the QuickChange mutagenesis kit (Stratagene, La Jolla, CA). BamH I and Bgl II restriction enzyme sites have compatible ends but ligation obliterates both restriction sites. The Xho I (nt 4477) to BamH I (nt 8498) restriction fragment containing the G and F genes of B9320 was then shuttled into the infectious RSV antigenomic cDNA clone pRSVC4G (Jin et al., 1998). The chimeric antigenomic cDNA was designated pRSV-G_BF_B. To delete the M2-2 gene from pRSV-G_BF_B, the Msc I (nt 7692) to BamH I (nt 8498) fragment from rA2ΔM2-2 which contained the M2-2 deletion (Jin et al., 2000a) was introduced into pRSV-G_BF_B. The chimeric cDNA clone that lacks the M2-2 gene was designated pRSV-G_BF_BΔM2-2.

Recovery of recombinant RSV

Recovery of recombinant RSV from cDNA is described herein. Briefly, HEP-2 cells in 6-well plate at 80% confluence were infected with MVA at an m.o.i. of 5 pfu/cell for 1 h and then were transfected with full-length antigenomic plasmids (pRSV-G_BF_B or pRSV-G_BF_BΔM2-2), together with plasmids expressing the RSV N, P, and L proteins using LipofecTACE reagent (Life Technologies, Gaithersburg, MD). After incubating the transfected cells at 35°C for three days, the culture supernatants were passaged in Vero cells for six days to amplify rescued virus. The recovered recombinant viruses were biologically cloned by three successive plaque purifications and further amplified in Vero cells. Virus recovered from pRSV-G_BF_B transfected cells was designated rA-G_BF_B and that from pRSV-G_BF_BΔM2-2 transfected cells was designated rA-G_BF_BΔM2-2. Virus titer was determined by plaque assay and plaques were visualized by immunostaining using polyclonal anti-RSV A2 serum (Biogenesis, Sandown, NH).

Virus characterization

The expression of viral RNA for each recovered chimeric RSV was analyzed by Northern blotting. Total cellular RNA was extracted from virus infected cells at 48 hr post-infection. The RNA blot was hybridized with a γ -³²P-ATP labeled oligonucleotide probe specific for the F gene of B9320 (GAGGTGAGGTACAATGCATTAATAGCAAGATGGAGGAAGA) or a γ -³²P-ATP labeled probe specific for the F gene of A2 (CAGAAGCAAAACAAAATGTGACTGCAGTGAGGATTGTGGT). To detect the G mRNA of the chimeric viruses, RNA blots were hybridized with a 190-nt riboprobe specific to the G gene of B9320 or a 130nt riboprobe specific to the G gene of A2. Both riboprobes were labeled with α -³²P-CTP. Hybridization was performed at 65°C in Express Hyb solution (Clontech, Palo Alto, CA) overnight. Membranes were washed at 65°C under stringent condition and exposed to film.

Viral specific proteins from infected cells were analyzed by immunoprecipitation of the infected cell extracts or by Western blotting. To

immunoprecipitate viral proteins, Vero cells were infected with virus at an moi of 1.0 and labeled with ^{35}S -promix (100 $\mu\text{Ci/ml}$ ^{35}S -Cys and ^{35}S -Met, Amersham, Arlington Heights, IL) from 14 hr to 18 hr postinfection. The labeled cell monolayers were lysed with RIPA buffer and the polypeptides immunoprecipitated by polyclonal goat anti-RSV A2 serum (Biogenesis, Sandown, NH) or by a polyclonal antibody against the M2-2 protein. Immunoprecipitated polypeptides were electrophoresed on SDS-PAGE and detected by autoradiography. For Western blotting analysis, virus infected Vero cells were lysed in protein lysis buffer and the proteins were separated on 12% SDS-PAGE. The proteins were transferred to a nylon membrane and immunoblotting was performed as described herein, using a monoclonal antibody against the G protein of B9320 or a monoclonal antibody against the G protein of A2 (Storch and Park, 1987 J. Med. Virol. 22:345-356).

Growth of chimeric RSV *in vitro* was compared with wild type recombinant A2 (rA2) and rA2 Δ M2-2. Growth cycle analysis was performed in both HEP-2 and Vero cells. Cells grown in 6-cm dishes were infected with each virus at a moi of 0.01 or 0.1. After 1 hr adsorption at room temperature, the infected cell monolayers were washed three times with PBS, and incubated at 35°C with 4 ml of Opti-MEM in an incubator containing 5% CO_2 . At various times post-infection, 200 μl of the culture supernatant was collected, and stored at -70°C for virus titration. Each aliquot removed was replaced with an equal amount of fresh medium. Virus titer was determined by plaque assay in Vero cells on 12-well plates using an overlay of 1% methylcellulose and 1 X L15 medium containing 2% FBS.

Virus replication in cotton rats

Virus replication *in vivo* was determined in respiratory pathogen-free *S. Hispidus* cotton rats. Cotton rats in groups of 12 were inoculated intranasally under light methoxyflurane anesthesia with $10^{5.5}$ pfu of virus per animal in a 0.1-ml inoculum. On day 4 post-inoculation, six animals were sacrificed by CO_2 asphyxiation and their nasal turbinates and lungs were harvested separately. Tissues were homogenized and virus titers determined by plaque assay in Vero

cells. Three weeks later, the remaining 6 animals were anesthetized, their serum samples were collected, and a challenge inoculation of 10^6 pfu of biologically derived wild type RSV strain A2 or B9320 administered intranasally. Four days post-challenge, the animals were sacrificed and both nasal turbinates and lungs were harvested, homogenized and virus titer determined by plaque assay. Serum neutralizing antibodies against RSV A2 or B9320 strain were determined by a 50% plaque reduction assay (Coates et al., 1966, Am. J. Epidemiol. 83(2):299-313).

Virus replication in AGM

Recombinant RSV was evaluated for their replication, immunogenicity and protective efficacy in AGM (*Cercopithecus aethiops*). AGM, obtained from St. Kitts with an average age of 4.2 years and body weight ranging from 2.2 to 4.3 kg, were used in the first study (study A) to compare the replication of rA2 with wild type A2. The second study (study B) used AGM with ages ranging from 5.3 to 8.4 years and an average body weight of 4.15 kg. None of the monkeys had detectable serum neutralizing antibodies for RSV B9320 or A2 (titer $< 1:10$). Groups of 4 monkeys were inoculated with either wild type A2, rA2, rA2 Δ M2-2, wild type B9320 or rA-G_BF_B by both intranasal and intratracheal route with a dose of $10^{5.5}$ pfu in a 1.0 ml inoculum at each site. Following inoculation, daily nasopharyngeal (NP) swabs were collected from each monkey for 12 days under Telazol anesthesia and tracheal lavage (BAL) were collected on days 3, 5, 7 and 10 post-infection (Kakuk et al., 1993, J. Infect. Dis. 167(3):553-561). On day 28 post-infection, serum samples were collected from each infected monkey and the monkeys were challenged with either wild type A2 or B9320 at both the intranasal and intratracheal sites with a dose of $10^{5.5}$ pfu in a 1.0-ml inoculum. Replication of the challenge virus in the upper and lower respiratory tracts of monkeys was examined by quantitation of virus shed in NP and tracheal lavage specimens. The NP samples were collected daily for 10 days and BAL samples were collected on days 3, 5, 7 and 10 post-challenge. Fourteen days after wild type virus challenge, serum samples were collected for measurement of RSV neutralizing antibody by the 50% plaque

reduction assay using wild type A2 or B9320 viruses. The virus shed in the NP and BAL samples were quantitated by plaque assay using Vero cells.

13.3 RESULTS

Construction of cDNA and recovery of RSV A/B chimeric virus

Previously, we constructed an infectious antigenomic cDNA encoding wt RSV strain A2 and its derivative bearing a deletion of the M2-2 gene. Here, these cDNAs were modified by replacing the G and F genes of the A2 strain with those of B9320 to produce chimeric viruses expressing RSV subgroup B antigens. The gene start and gene end sequences are very conserved between the two RSV subgroups. Therefore, the complete G and F genes of B9320 including their own gene start and gene end signals were transferred to the A2 cDNA backbone (Fig. 26). The cDNA encoding the G and F genes of B9320 was obtained by RT/PCR and confirmed by sequence analysis. The constructed chimeric cDNA was designated pRSVA-G_BF_B. pRSVA-G_BF_BΔM2-2 was constructed by deleting the M2-2 gene from pRSVA-G_BF_B. The M2 gene containing the deletion of the M2-2 open reading frame from rA2ΔM2-2 was introduced into pRSVA-G_BF_B through the unique Msc I and BamH I restriction enzyme sites. Both chimeric viruses (rA-G_BF_B and rA-G_BF_BΔM2-2) were recovered from cDNA using the previously described rescue system. The recovered recombinant viruses were plaque-purified and amplified in Vero cells.

Characterization of the recombinant chimeric viruses in vitro

Expression of the subgroup specific proteins by the chimeric viruses was analyzed by Northern and Western blotting. Using strain specific probes, B9320-specific G and F mRNAs were detected in cells infected with rA-G_BF_B and rA-G_BF_BΔM2-2 (Fig. 27A). The M2-2 gene was not detected in cells infected with rA-G_BF_BΔM2-2, confirming that the M2-2 gene was deleted from this chimeric virus. The B9320 strain specific protein expression of the two chimeric viruses was also compared with that of rA2, rA2ΔM2-2 and wild type B9320 (Fig. 27B). The F1 protein of rA-G_BF_B and

rA-G_BF_BΔM2-2 showed the same rate of migration mobility as that of B9320, both migrated faster than that of A2. Western blotting analysis using strain specific monoclonal antibodies confirmed that the G protein of subgroup B was expressed by rA-G_BF_B and rA-G_BF_BΔM2-2 (Fig. 27B). Western blotting using a polyclonal antibody specific to the M2-2 protein further confirmed the ablation of the M2-2 gene in rA2ΔM2-2 and rA-G_BF_BΔM2-2.

Replication of chimeric viruses, rA-G_BF_B and rA-G_BF_BΔM2-2, was compared to rA2 and rA2ΔM2-2 in both the HEp-2 and Vero cells (Fig. 28). In Vero cells, infected at an moi of 0.1, both rA-G_BF_B and rA-G_BF_BΔM2-2 reached peak titers similar to that seen with wild type rA2 and rA2ΔM2-2 respectively. At a lower moi of 0.01, the peak titer of rA-G_BF_B was slightly reduced compared to rA2; the level of replication of rA-G_BF_BΔM2-2 was reduced by about 10-fold compared to rA-G_BF_B. In HEp-2 cells, at moi of 0.1, rA-G_BF_B showed a slightly lower peak titer compared to wt A2 whereas the replication of rA-G_BF_BΔM2-2 was reduced by about 100-fold. At moi of 0.01, the peak titer of rA-G_BF_B was reduced by about 10-fold compared to rA2 and the peak titer of rA-G_BF_BΔM2-2 was reduced by 100-fold. Therefore, similar to that observed for rA2ΔM2-2, rA-G_BF_BΔM2-2 also exhibited restricted replication in HEp-2 cells, whereas its replication in Vero cells was less impaired.

Replication of chimeric RSV in cotton rats

Cotton rats are susceptible to both subgroup A and B RSV infection. The levels of replication of rA-G_BF_B and rA-G_BF_BΔM2-2 in the nasal turbinates and lungs of cotton rats were compared with rA2, rA2ΔM2-2 and wild type B9320 (Table 21). The replication of rA-G_BF_B was below the limit of detection by plaque assay in the nasal turbinates, its replication in lung tissue was reduced by about 3.6 log₁₀ compared to wild type B9320 and by about 2.0 log₁₀ relative to rA2. The replication of rA2ΔM2-2 was not detected in the nasal turbinates and was 1.6 log lower in the lung compared to rA2. Removal of M2-2 from rA-G_BF_B further attenuated the chimeric

virus. No virus replication was detected in either the nasal turbinates or lungs of cotton rats infected with rA-G_BF_BΔM2-2.

Although rA-G_BF_B and rA-G_BF_BΔM2-2 were attenuated in cotton rats, both chimeric viruses induced sufficient immunity against RSV to protect the animals from challenge (Table 21). The level of serum anti-RSV neutralizing antibody induced by rA-G_BF_B was 2.85-fold lower relative to that induced by wild type B9320. Serum anti-RSV neutralizing antibody induced by rA-G_BF_BΔM2-2 was approximately 4-fold lower compared to that induced by B9320 and 1.5-fold lower than that of rA-G_BF_B. By comparison, the level of serum anti-RSV neutralizing antibody induced by rA2ΔM2-2 was similarly reduced by approximately 2-fold compared to that of rA2.

Replication of wt RSV and rA2ΔM2-2 in AGM

In order to investigate RSV attenuation and immunogenicity in primates, replication of recombinant RSV was further studied in AGM. Study A examined the replication of recombinant A2 and wild type A2 virus in the respiratory tracts of AGM. RSV sero-negative AGM were infected with 5.5 log₁₀pfu of rA2 or wt A2 intranasally and intratracheally and virus shedding was monitored over a period of 12 days in both the upper and lower respiratory tracts. As shown in Table 22, rA2 replicated well in both the upper and lower respiratory tracts of AGM. rA2 reached a peak titer of 4.18 and 4.28 log₁₀pfu/ml at each site respectively and shed virus over the same length of time as the wild type A2 virus (Table 22, study A), though the peak titer of rA2 in the respiratory tracts of AGM was slightly lower than that obtained for wild type A2 virus. Having confirmed a high level of replication of rA2 in AGM, rA2ΔM2-2 was evaluated for its attenuation, immunogenicity, and protective efficacy in AGM. In a separate study (study B, Table 22), rA2ΔM2-2 showed a greatly reduced level of replication in both the nasopharynx and trachea compared to rA2. Its peak titer in nasopharynx had a reduction of 3.1 log₁₀ while the peak titer in the trachea was reduced by 3.25 log₁₀. Despite the much lower level of replication in the

respiratory tracts, rA2ΔM2-2 induced a significant level of serum anti-RSV neutralizing antibody. The antibody titer induced by rA2ΔM2-2 was about 4-fold lower than that induced by rA2 at three weeks post-infection (Table 23). When challenged with wild type A2 virus, rA2ΔM2-2 provided partial protection against wild type RSV replication in the upper respiratory tract and virtually complete protection in the lower respiratory tract of immunized monkeys. Monkeys inoculated with rA2 were fully protected in both the upper and lower respiratory tracts (Table 23). Although rA2ΔM2-2 did not provide complete protection in the respiratory tracts of immunized monkeys, it reduced virus shedding by 5 days. Two weeks after challenge, the level of serum anti-RSV neutralizing antibody from rA2ΔM2-2 infected monkeys approached that induced by rA2.

Replication of chimeric rA-G_BF_B and wild type B9320 in AGM

The level of replication of chimeric rA-G_BF_B was compared with that of wild type B9320. RSV sero-negative AGM were inoculated with 5.5 log₁₀pfu of rA-G_BF_B or B9320 by intranasal and intratracheal instillation. The throat swab and tracheal lavage samples were collected over 12 days for virus quantitation. B9320 replicated to a level similar to that of wild type A2 virus (Table 22). The peak titer of rA-G_BF_B in the respiratory tracts of the infected monkeys was about 1000-fold reduced compared to that of B9320. Animals infected with rA-G_BF_B shed virus for a shorter period than those infected with B9320. Despite its significantly attenuated replication, rA-G_BF_B provided complete protection when challenged with wild type B9320. No challenge virus was detected in either the upper or lower respiratory tracts of the monkeys previously immunized with rA-G_BF_B(Table 23). Consistent with the level of protection seen in monkeys immunized with rA-G_BF_B, the level of serum anti-RSV neutralizing antibody from these monkeys was only marginally reduced (about 2-fold) compared to that observed for wild type B9320 infected animals. The level of serum anti-RSV neutralizing antibody induced by rA-G_BF_B was augmented by subsequent wild type RSV infections

13.4 DISCUSSION

To expedite vaccine development for subgroup B RSV, a recombinant A2 virus was used as a vector to express subgroup B RSV surface antigens. The chimeric virus should elicit a balanced immune response and provide protection against subgroup B RSV infection. As an approach to expressing RSV subgroup B antigens, we constructed a different chimeric virus in which the G and F genes of the A2 strain were completely replaced by the G and F genes of the B9320 strain. The chimeric RSV was then further attenuated using a strategy developed for attenuating the A2 virus.

The recovered chimeric RSV (rA-G_BF_B) replicated efficiently in Vero cells, but its growth in HEP-2 cells was reduced by 5- to 10-fold relative to rA2. rA-G_BF_B was attenuated in both the upper and lower respiratory tracts of cotton rats. To determine if the attenuation of rA-G_BF_B was host specific, this chimeric virus was further evaluated in AGM that are genetically more closely related to humans than rodents. RSV infection in AGM is less well characterized and there is a wide range in the reported peak titer (Crowe et al., 1996, J. Infect. Dis. 173:829-839); (Kakuk et al., 1993, J. Infect. Dis. 167:553-561). Therefore, RSV infection was first tested in AGM using wild type viruses. Both subgroup A and subgroup B RSV were shown to replicate equally well in AGM and virus titers recovered from the upper and lower respiratory tracts of AGM were comparable to those observed in infected Chimpanzees (Crow et al., 1994, Vaccine 12:783-790). When rA-G_BF_B was evaluated in AGM, it showed a mean peak titer reduction of 3.0 log₁₀ in the upper respiratory tract and a reduction of 2.59 log₁₀ in the lower respiratory tract.

The level of attenuation of rA-G_BF_B in AGM was consistent with those levels observed in cotton rats. However, this result was somewhat different from that reported for a recently described chimeric RSV in which the G and F genes of A2 were replaced with those of RSV B1 strain (rAB1, Whitehead et al., 1999, J. Virol. 73:9773-80). Though rAB1 and rA-G_BF_B

are similarly attenuated in cotton rats, rAB1 was not attenuated in Chimpanzees. In contrast to rA-G_BF_B, rAB1 replicated better than wt RSV B1 in both the upper and lower respiratory tracts of Chimpanzees (Whitehead et al., 1999, J. Virol. 73:9773-80). Part of this discrepancy may be explained by the semi-permissiveness of Chimpanzees to wild type subgroup B RSV infection. However, there exists the possibility that rA-G_BF_B is more attenuated than rAB1 because of differences in the subgroup B strain surface antigens or constellation effects when these antigens are introduced into an A2 background. Therefore, it appears that chimerization of closely related different heterologous proteins can result in different phenotypes. Chimerization of surface antigens resulting in an attenuated virus has been reported for several paramyxoviruses. A chimeric measles virus with the HN and F proteins replaced by the G protein of VSV was highly restricted in replication in vitro (Spielhofer et al., 1998, J. Virol. 72:2150-2159). A chimeric Rinderpest virus in which the F and H proteins were replaced by the heterologous surface proteins of a closely related peste-des-petits-ruminants virus was attenuated in vitro, as indicated by slow virus growth and low virus yield (Das et al., 2000, J. Virol. 74:9039-9047). Most recently, it was reported that the PIV3-PIV2 chimeric virus, in which the F and HN genes of PIV3 were replaced by those of PIV2 was not attenuated in vitro, but it was severely attenuated in hamsters, AGM and Chimpanzees (Tao et al., 2000, J. Virol 74:6448-6458). On the other hand, the chimeric PIV3-PIV1 was not attenuated in vivo (Tao et al., 1998, J. Virol. 72:2955-2961; Tao et al., 1999, Vaccine 17:1100-1108). Though attenuated in AGM, rA-G_BF_B induced significant levels of anti-RSV neutralizing antibody and provided complete protection against subsequent challenge with wild type subgroup B RSV.

In this study, rA2ΔM2-2 was evaluated for its attenuation, immunogenicity and protection against wild type RSV challenge in AGM. rA2ΔM2-2 was shown to be attenuated in the respiratory tracts of AGM and following challenge, much reduced replication of wild type RSV was

observed in animals previously infected with rA2ΔM2-2. The level of replication and protection observed for rA2ΔM2-2 in AGM is very similar to that reported in a Chimpanzee study for a similar recombinant RSV that had the M2-2 protein expression silenced (Bermingham and Collins, 1999, pNAS USA 96:11259-11264; Teng et al., 2000, J. Virol. 74: 9317-9321). rA2ΔM2-2 may prove to be more attenuated in humans than a previously tested vaccine candidate cpts248/404 (Teng et al., 2000, J. Virol. 74: 9317-9321). cpts248/404 was neither sufficiently attenuated nor genetically stable in naive infants (Crowe et al., 1994, Vaccine 12:783-790; Wright et al., 2000, J. Infect. Dis. 182:1331-1342). The serum anti-RSV neutralizing antibody titer induced by rA2ΔM2-2 was slightly lower than that induced by the wild type RSV infection. However, the augmentation of neutralizing antibody titer after the challenge suggests that the immunogenicity of rA2ΔM2-2 could be enhanced by repeat administrations.

Since rA2ΔM2-2 exhibits many of the desired features in a live attenuated vaccine, the deletion of the M2-2 gene was considered as an appropriate way to further attenuate the chimeric rA-G_BF_B. In vitro study indicated that rA-G_BF_BΔM2-2 had similar level of attenuation as rA2ΔM2-2, exhibiting increased syncytial formation, reduced growth in HEp-2 cells and unbalanced RNA transcription to replication. As the chimeric rA-G_BF_B virus is already attenuated in both the cotton rats and AGM, rA-G_BF_BΔM2-2 is expected to be more attenuated than rA2ΔM2-2. However, cotton rats studies indicated that rA-G_BF_BΔM2-2 was still capable of inducing a level of serum RSV neutralizing antibody approaching that induced by rA-G_BF_B and provided complete protection against subsequent experimental challenge. Therefore, rA-G_BF_BΔM2-2 may represent a suitable vaccine candidate for protecting against subgroup B RSV infection.

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Table 2 Replication, immunogenicity, and protection of recombinant RSV against wt RSV infection in the upper and lower respiratory tracts of cotton rats

Virus ^a	Virus titer ^b (mean log ₁₀ pfu/g ± SE) ^b		Neutralizing Ab titer (mean reciprocal log ₂) ^c	Challenge Virus ^d	Challenge virus titer (mean log ₁₀ pfu/g ± SE) ^b	
	Nasal Turbinates	Lung			Nasal Turbinates	Lung
rA2	3.9 ± 0.13	3.57 ± 0.07	9.75	A2	< 1.4	< 1.4
rA2ΔM2-2	< 1.4	2.02 ± 0.12	9.40	A2	< 1.4	< 1.4
Control	< 1.4	< 1.4	< 3.3	A2	4.2 ± 0.14	6.0 ± 0.06
B9320	2.8 ± 0.57	5.6 ± 0.05	10.64	B9320	< 1.4	< 1.4
rA-G _B F _B	< 1.4	1.94 ± 0.31	9.13	B9320	< 1.4	< 1.4
rA-G _B F _B ΔM2-2	< 1.4	< 1.4	8.57	B9320	< 1.4	< 1.4
Control	< 1.4	< 1.4	< 3.3	B9320	2.3 ± 0.53	5.2 ± 0.01

^a Cotton rats were administered with 5.5 log₁₀ PFU of virus intranasally under light anesthesia on day 0 and sacrificed on day 4.

^b Virus titers were determined in the nasal and lung tissues by plaque assay.

^c Serum RSV neutralizing antibody titers were determined by a complement-enhanced 50% plaque reduction assay with wt A2 or wt B9320.

^d On day 21 of virus infection; cotton rats in groups of six were challenged with wt A2 or wt B9320.

Table 3. Replication or recombinant RSV in the upper and lower respiratory tracts of African Green monkeys

Virus ^a	AGM Number	Virus shedding (Days)	Virus peak titer (Mean log ₁₀ pfu ± SE) ^b	
			Nasopharyngeal swab	Tracheal lavage
wt A2	4 (Study A)	8	4.67 ± 0.17	4.97 ± 0.04
rA2	4 (Study A)	8	4.18 ± 0.18	4.28 ± 0.27
rA2	4 (Study B)	9	3.44 ± 0.27	3.91 ± 0.18
rA2ΔM2-2	4 (Study B)	4	0.33 ± 0.26	0.66 ± 0.40
B9320	4 (Study B)	9	4.51 ± 0.18	4.36 ± 0.45
rA-G _B F _B	4 (Study B)	4	1.50 ± 0.42	1.77 ± 0.25

^a African green monkeys were administered with 5.5 log₁₀ PFU of virus intranasally and intratracheally. Nasopharyngeal swab samples were collected daily for 12 days, and tracheal-lavage samples were collected on days 3, 5, 7 and 10.

^b Virus titers were determined in the nasopharyngeal swab and tracheal-lavage by plaque assay and the peak titers were shown.

Table 3. Evaluation of level of immunogenicity and efficacy against wild type challenge virus in African Green monkeys

Virus	Challenge Virus ^a	Virus peak titer (Mean log ₁₀ pfu ± SE) ^b		Neutralizing Ab titer (mean reciprocal log ₂) ^c	
		Nasopharyngeal swab	Tracheal lavage	Pre-challenge	Post-challenge
rA2	A2	<0.7	<0.7	9.7	10.5
rA2ΔM2-2	A2	2.64 ± 0.07	0.46 ± 0.47	8.25	9.25
B9320	B9320	<0.7	<0.7	10.0	10.75
rA-G _B F _B	B9320	<0.7	<0.7	9.0	11.25

^a African green monkeys were administered with 5.5 log₁₀PFU of virus intranasally and intratracheally and on day 21, monkeys were challenged with wt A2 or wt B9320 at a dose of 5.5 log₁₀PFU intranasally and intratracheally.

^b Nasopharyngeal swab samples were collected daily for 10 days, and tracheal-lavage samples were collected on days 3, 5, 7 and 10. Challenge

virus titers were determined by plaque assay. Only the peak titers are shown.

^c Serum RSV neutralizing antibody titers from infected monkeys before challenge infection and 14 days post-challenge were determined by a complement-enhanced 50% plaque reduction assay with wt A2 or wt B9320.

The present invention is not to be limited in scope by the specific
embodiments described which are intended as single illustrations of
individual aspects of the invention, and any constructs, viruses or enzymes
which are functionally equivalent are within the scope of this invention.
Indeed, various modifications of the invention in addition to those shown
and described herein will become apparent to those skilled in the art from
the foregoing description and accompanying drawings. Such modifications
are intended to fall within the scope of the appended claims.

Various publications are cited herein, the disclosures of which are
incorporated herein by reference in their entireties.

WHAT IS CLAIMED IS:

1. An isolated infectious respiratory syncytial virus particle which comprises an respiratory syncytial virus antigenome or genome containing at least one functional deletion in a viral accessory gene.

2. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing an M2-2 gene mutation.

3. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing an SH gene mutation.

4. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing an NS1 gene mutation.

5. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing an NS2 gene mutation.

6. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing both an M2-2 gene mutation and an SH gene mutation.

7. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing both an M2-2 gene mutation and an NS1 gene mutation.

8. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing both an M2-2 gene mutation and an NS2 gene mutation.

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9. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing both an NS1 gene mutation and an NS2 gene mutation.

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10. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing both an NS1 gene mutation and an SH gene mutation.

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11. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing both an NS2 gene mutation and an SH gene mutation.

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12. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing an NS1 gene mutation, an NS2 gene mutation and an SH gene mutation.

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13. An isolated infectious respiratory syncytial virus particle having an attenuated phenotype comprising a respiratory syncytial virus antigenome or genome containing an M2-1 gene mutation.

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14. The isolated infectious respiratory syncytial virus particle of Claim 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13 which further comprises a

heterologous sequence.

15. The recombinant RNA molecule of Claim 14 in which the heterologous sequence is derived from the genome of another strain of respiratory syncytial virus.

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16. The recombinant RNA molecule of Claim 14 in which the heterologous sequence is derived from the genome of a virus other than respiratory syncytial virus.

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17. A vaccine comprising a respiratory syncytial virus, the genome of which contains the reverse complement of an mRNA coding sequence operatively linked to a polymerase binding site of a respiratory syncytial virus, and a pharmaceutically acceptable carrier.

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18. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of the complete M2-2 gene.

19. The vaccine of Claim 17 in which the mRNA coding sequence contains a mutagenized M2-1 gene.

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20. The vaccine of Claim 19 in which the M2-1 gene is mutagenized by cysteine scanning mutagenesis.

21. The vaccine of Claim 19 in which the M2-1 gene is mutagenized by C-terminal truncation of the M2-1 protein.

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22. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of the complete SH gene.

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23. The vaccine of Claim 17 in which the mRNA coding

sequence contains a deletion of the complete M2-2 gene.

24. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of the complete NS1 gene.

5 25. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of the complete NS2 gene.

10 26. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of both the complete M2-2 gene and the complete SH gene.

15 27. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of both the complete M2-2 gene and the complete NS1 gene.

20 28. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of both the complete M2-2 gene and the complete NS2 gene.

25 29. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of both the complete NS1 gene and the complete NS2 gene.

30 30. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of both the complete M2-2 gene and the complete SH gene.

35 31. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of both the complete NS1 gene and the complete SH gene.

32. The vaccine of Claim 17 in which the mRNA coding sequence contains a deletion of the complete NS1 gene, the complete NS2 gene and the complete SH gene.

5 33. The vaccine of Claim 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31 or 32 which further comprises a heterologous sequence

34. The vaccine of Claim 33 in which the heterologous gene is derived from the genome of influenza.

10 35. A pharmaceutical composition comprising the attenuated vaccine of Claim 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28 or 29.

ABSTRACT

The present invention relates to genetically engineered recombinant respiratory syncytial viruses and viral vectors which contain deletions of various viral accessory gene(s) either singly or in combination. In accordance with the present invention, the recombinant respiratory syncytial viral vectors and viruses are engineered to contain complete deletions of the M2-2, NS1, NS2, or SH viral accessory genes or various combinations thereof. In addition, the present invention relates to the attenuation of respiratory syncytial virus by mutagenesis of the M2-1 gene.

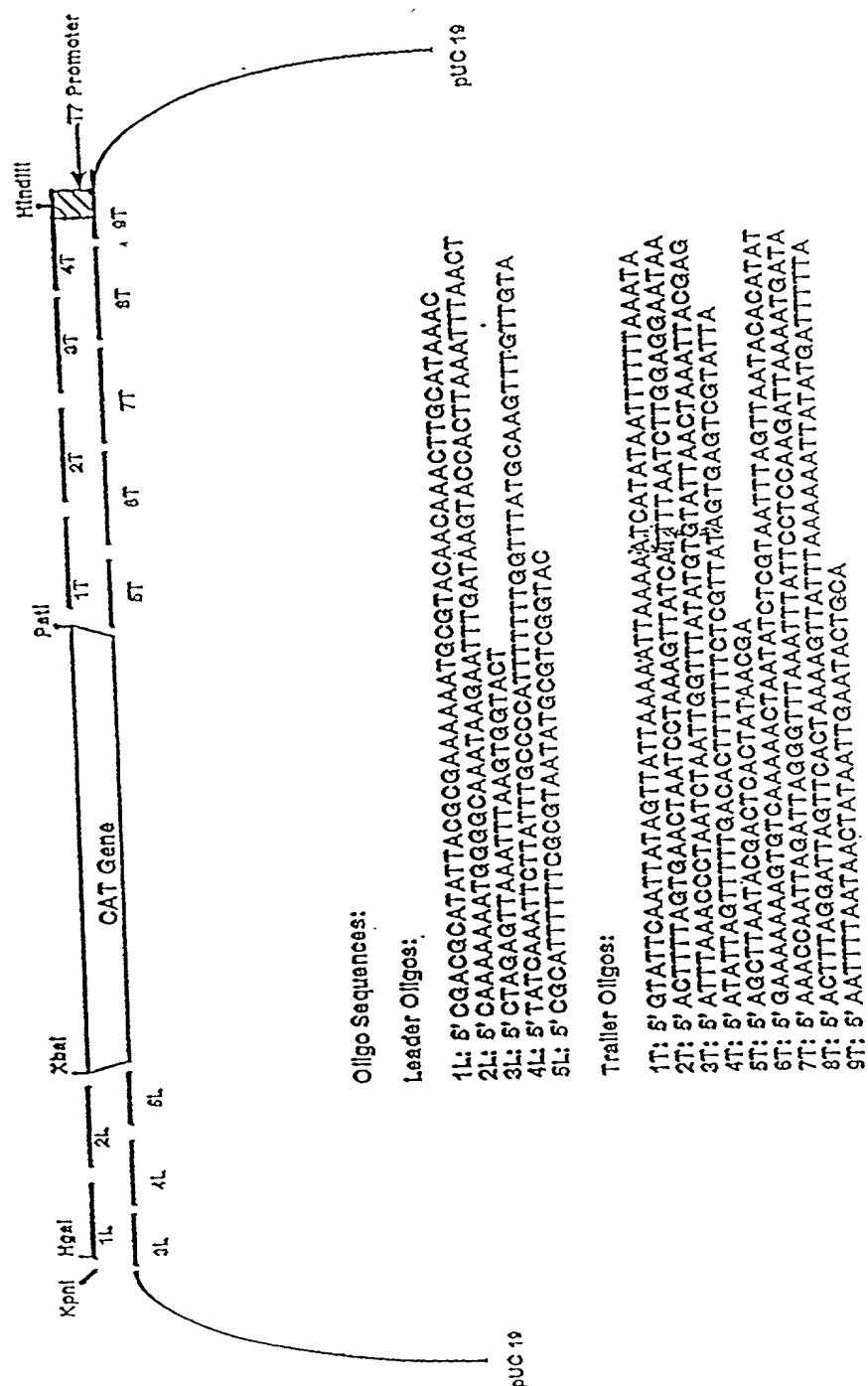


FIG. 1

7682-052 (Sheet 2 of 28)

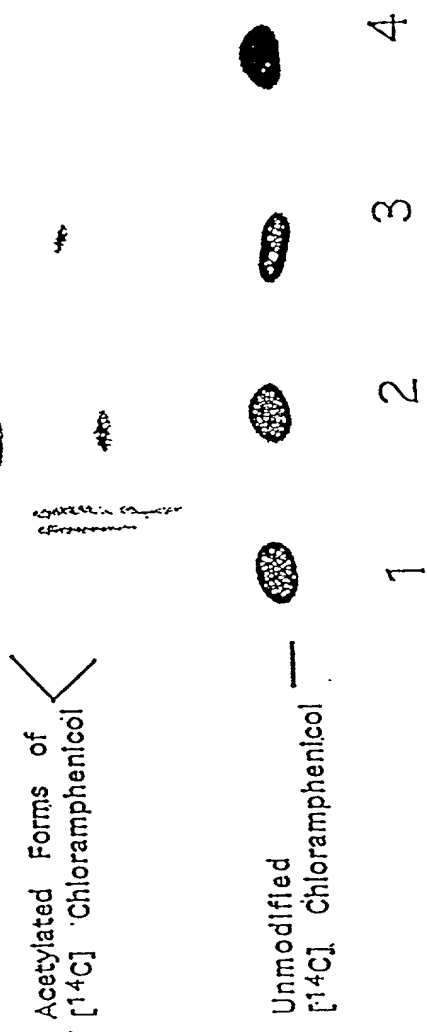


FIG. 2

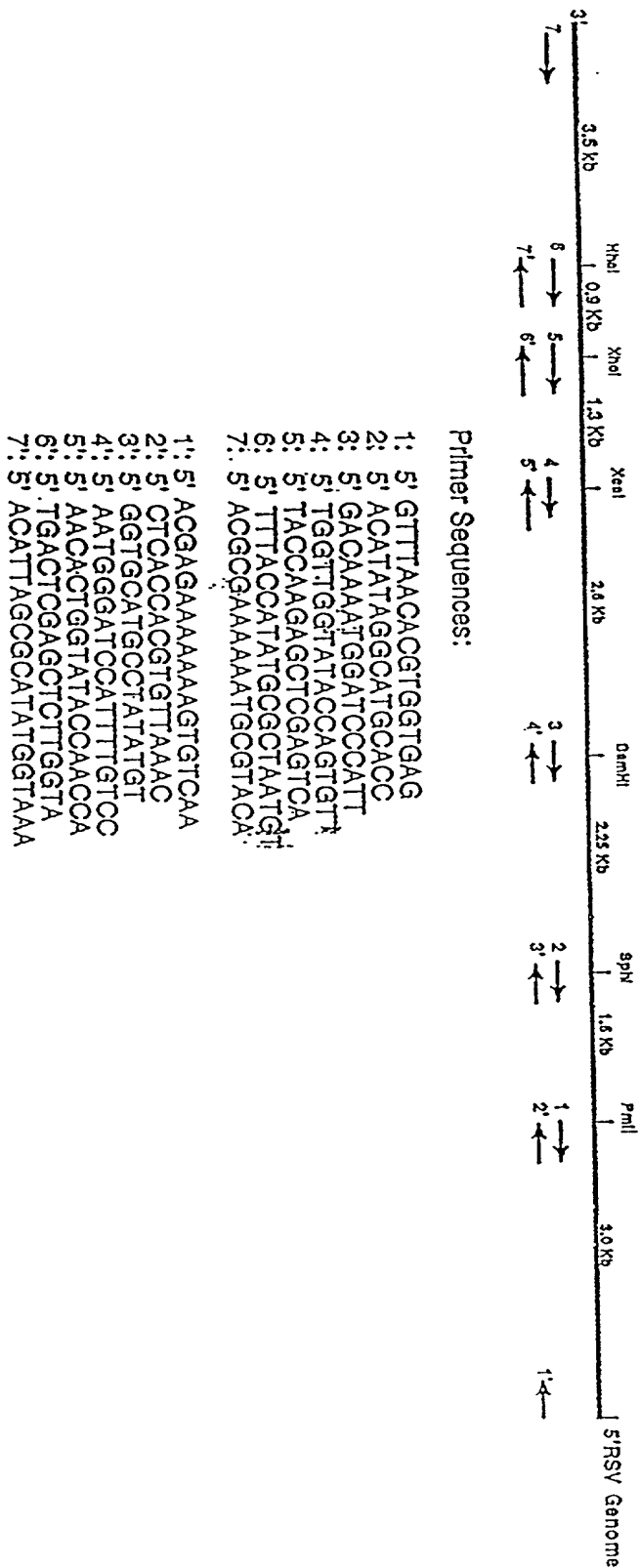
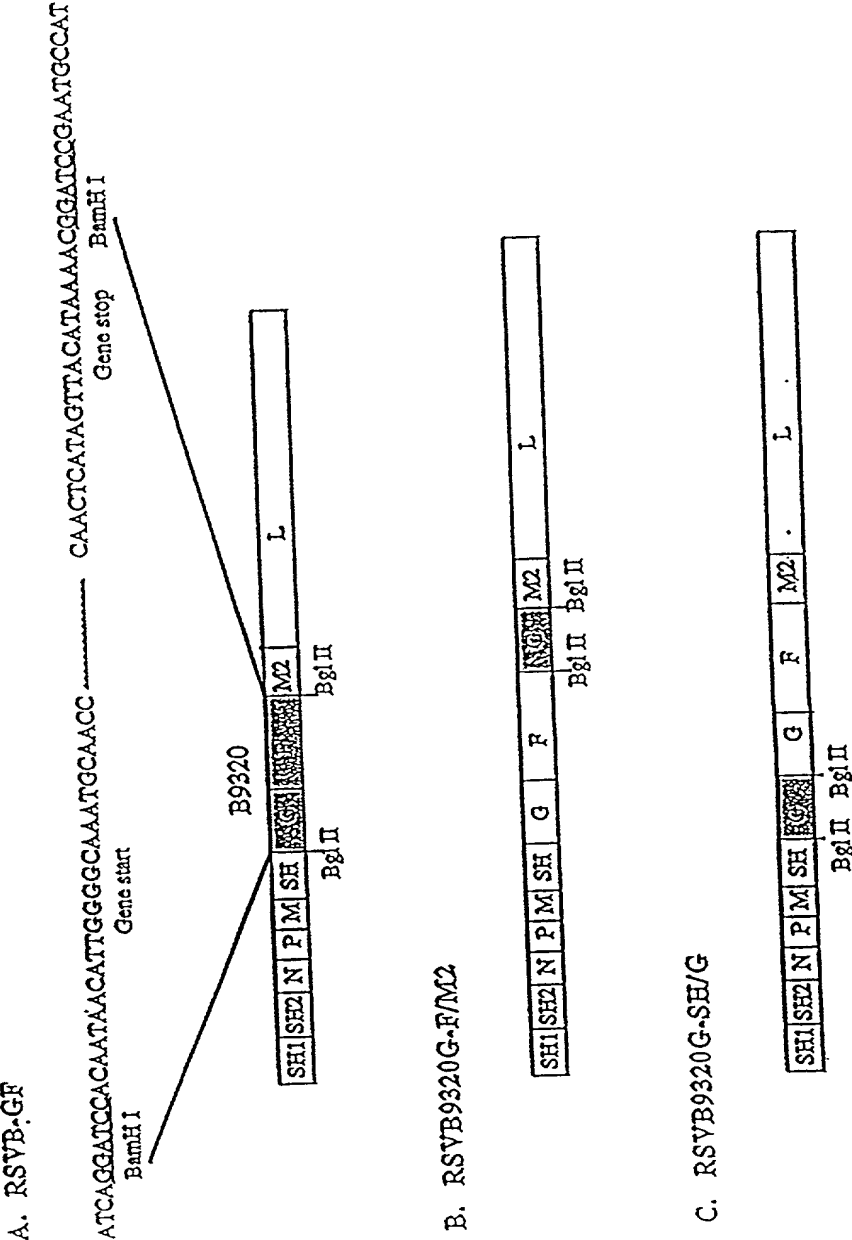


FIG. 3



FIGS. 4A-C

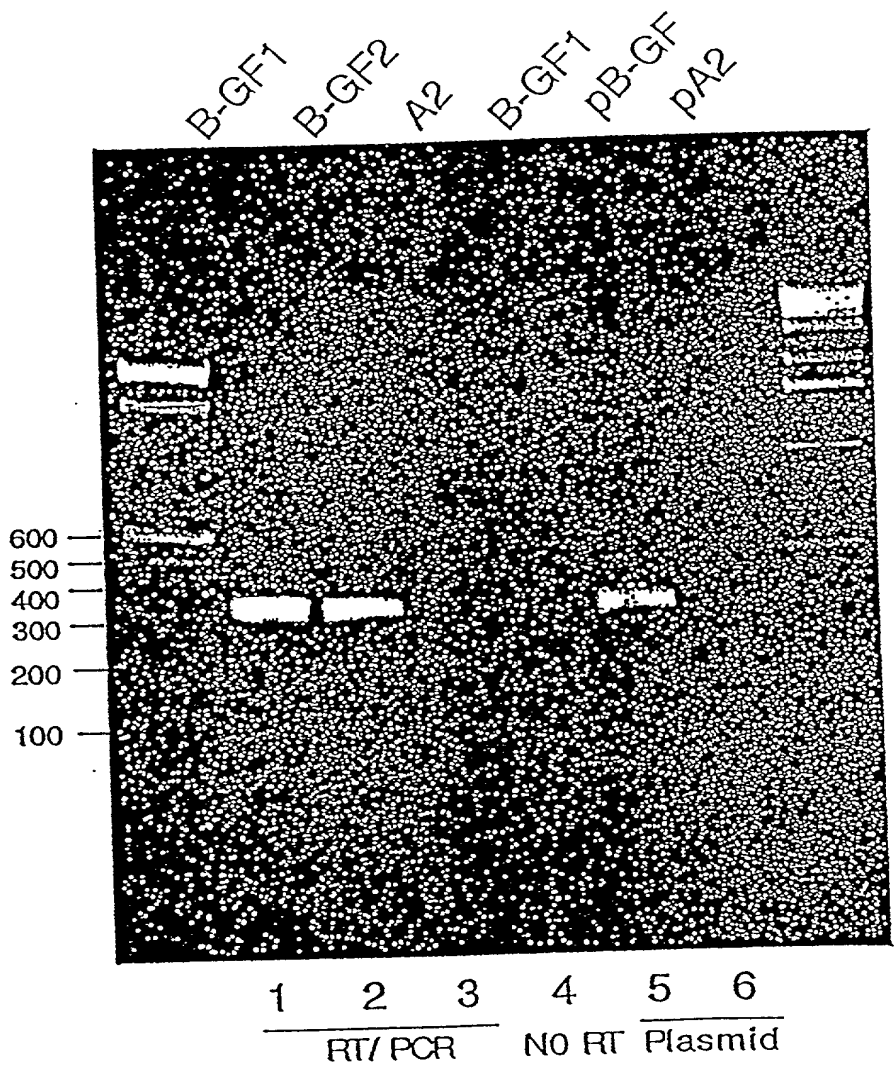
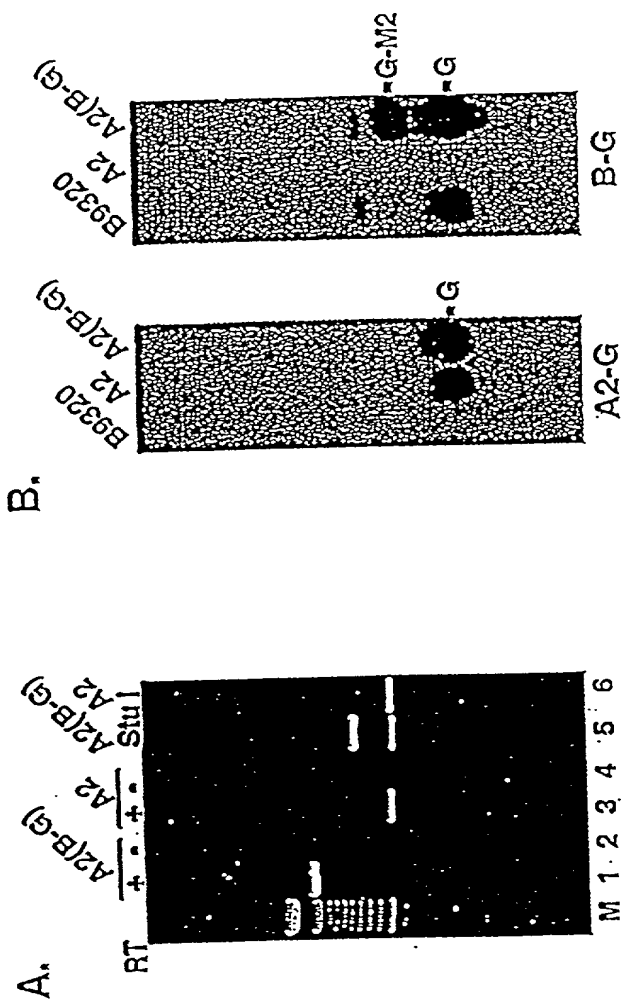


FIG. 5



FIGS. 6A-B

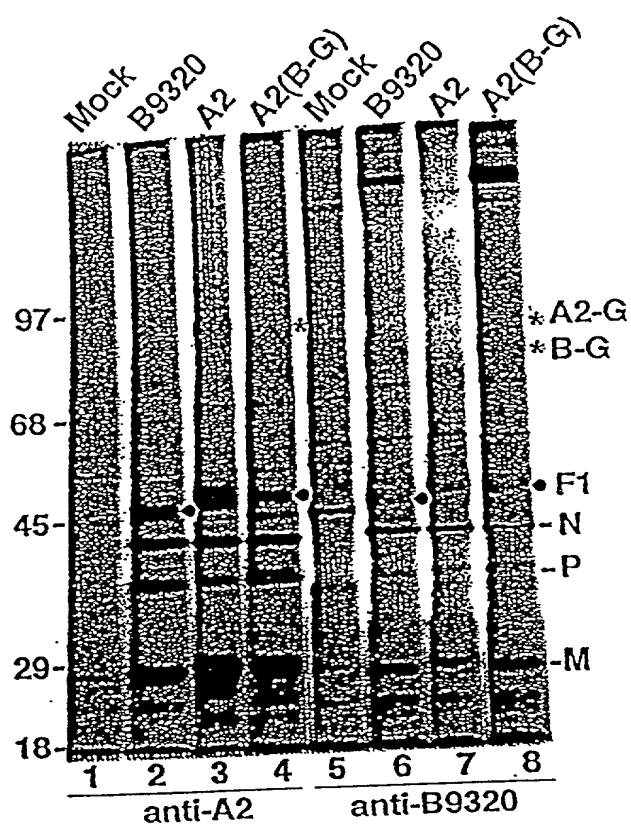


FIG. 7

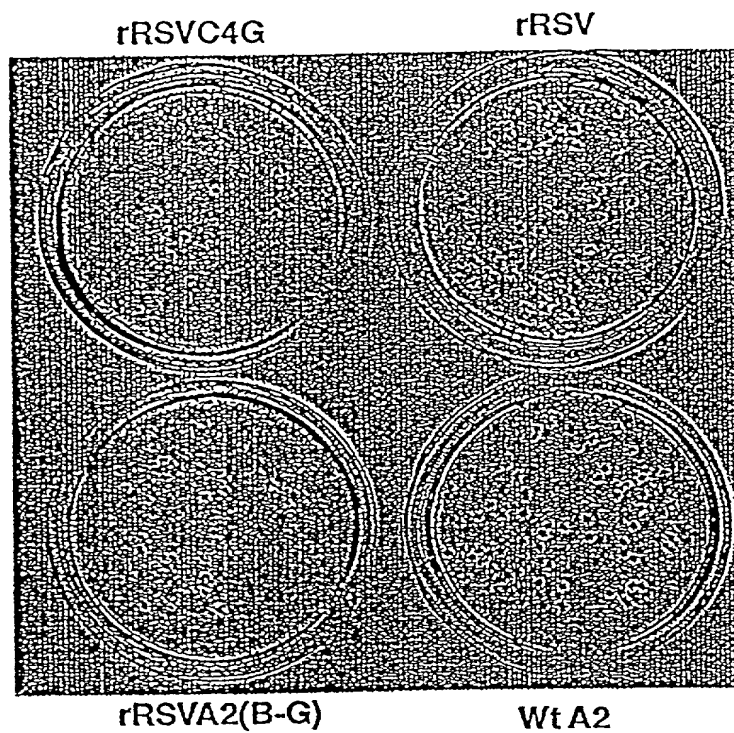


FIG. 8

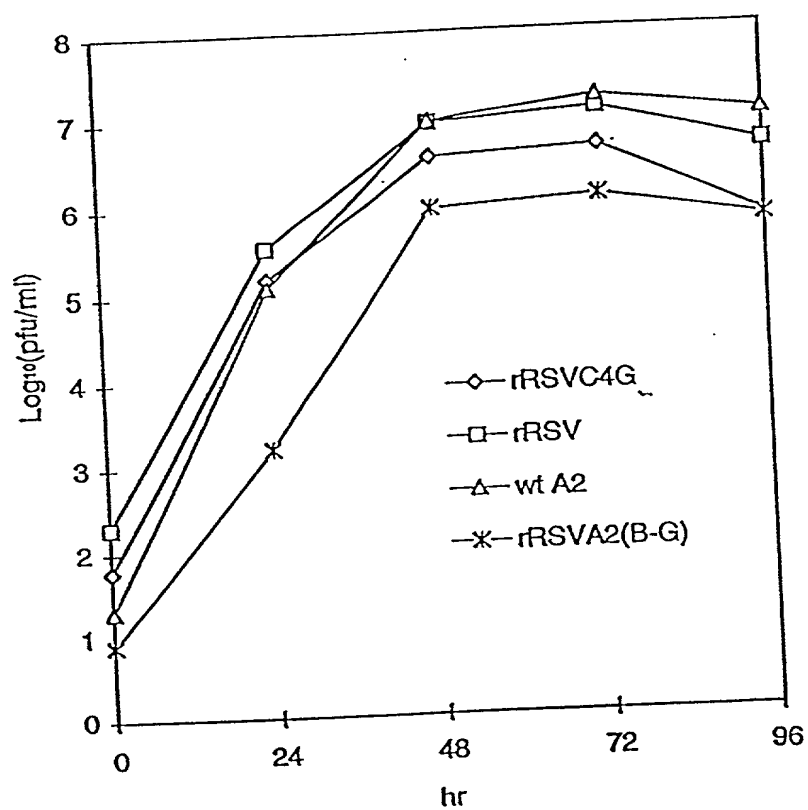


FIG. 9

MDPIINGNSANVILT DSVLKGVISFSECNA LGSYIFNGPYLKNDY TNLISRQNPFLIEHM LKKLNTQSLISKYH 75
 KGEIKLEPTFYQSL LMTYKSMTSSEQIAT TNLKKITIRRAIEIS DVKVYAILNKLGLKE KDKIKSNNGODEDNS 150
 VITTIKKDILSAVK DNQSHLKADKNHSTK QKDTIKTTLKKLMC SMQHPSPSWLIHWENL YTKLNNILTQYRSNE 225
 VQNHGFTLIDNQTLS GEQFTINQXGCIYH KKLKRIITVTFYNQFL TWKDI SLRINVCIL TWISNCLNTNKS LG 300
 LRCGFNNVILTQLFL YGDCILKLFHNEGFI IIEKEVEGFIMSLIN TTEEDQFKEKFFYNSM LNNITDAANKAQKQL 375
 LSRVCHTLLDKTVSD NIINGRWIILLSKFL KLKLAGDNMLNLS ELXFLFRIFGHPMD ERQAMD AVKINCNET 450
 KFYLLSSLMLRGAF IYRIIKGFVNINRW PTLRNAIVLPLRWLT YKLNTPPSLLELTH RDLIVLSGLRFYREF 525
 RLPKKVDLEMIINDK AISPKNLIMTSFPR NYMPSHIQNYIEHEK LKFSSEDSKRRVLEY VLRDNKFNECDLYNC 600
 VVNQSYLNNENHVS LTGKEBELSVGRMFA MQPGRQVQILAEK MIAENILQFFPESLT RYGDLELQKILEKA 675
 GISNKSNNRYNDNANN YISKCSIIITDLSKEN QAFRYETSCICSDVL DELHGVQSLSFWLH TTPHVTIICTYRHAP 750
 FYIGDHIVDLNNVDE QSLYRYHMGIEGW CQKLWTEATISLDDL ISLKGKFSITALING DNQSIDISKPIRLME 825
 GQTHAQADYLLAALS LKLLYKEYAGIGHKL KGTETIYISRDQMFS VMLYNQIALQLKSHA KTNKLYLDILKVLK 900
 FKVSLESIGSLTQEL EYRGESLLCSLIFFRN TEAIVHSVFILSYT NHDLDKDLQDLSDDR LNKFLTCLITFDKNP 1050
 TLYMNLPLFGGDP NLLYRSFYRRTPDFL VTEVLSTAPNKIFSK SAQHYTTTEIDLDNDI MONEPTYPHGLRVV 1125
 NAEFVTLMDPQALG SERQAKITSEINRLA AIDTIDIDRATENMR KNTILLIRILPLDCN ROKREILSMENLSIT 1200
 YESLPFYKAEKIVNL ISGTSIITNILEKTS KYTSTISSGIIIEK YVNSLITRGERGPTK PWVGSSTQEKTMPV 1275
 ELSKYVREBSWLSN IVGVTSPIIMYTMDI DEFMEELSIGTLGLT YEKAKKLPQYLSVN YLHRLTVSSRPCEFP 1350
 YNRQVLTKKQDQID LIAKLDWVYASIDNK IDIVFQNCISFGLSL MSVVEQFTNVCPNRI ILIPKLEIHLMKPP 1425
 ASIPAYRTTYHFDT SPINRILTEKYGDEP YVELELSNKTLSKGS HVNSNLILAHKISDY FHNTVILSTNLAGHW 1500
 IFTGDVDIHKLKQVI QKQHMFLPDKISLTQ LKVFENAYKTVLLCF HKGYGKAKLECDMT SDLLCVLELIDSSYW 1575
 ILTIQLMKDSKGIFE KDWGEGYITDHFMTN SFKLMFLKRLNVAEF TVCPWVNVNIDYHPTH MKAILTYIDLVRMGL 1650
 KSMKSVFLEQKVIKY ILSQDASLHRVKGCH SDNTHLLTKHIRIAN SELENNYNNKLYHPTP ETLENILANPIKSND 1725
 INIDRIHIKNHKEFN DEFYTSNLEFYINYNF AMIRTNYSKQDLVNL FPMVVIDRIIDHSGN TAKSNQLYTTTSHQI 1800
 KYKTINDYCIGKNVDS TMLPLLNNKLIKSS KISIEYILKDLKIKD PNCIAFIGEGAGNLL LRTVVELHPDIRYIY 1875
 SLVHNSTSLYCMLEPW HHINRFNFVFSSTCC LRTYNGHINIDYGEN YLHIKFAEPISLFVC DAELSVTNWSKIII 1950
 RSLKDCNDHSLPIEF LRLXNGHINIDYGEN LRTYNGHINIDYGEN YLHIKFAEPISLFVC DAELSVTNWSKIII 2025
 EWSKTVREKCKYQSSV NKCMILIVRYHRAQDDI DFKLDNITILKTYVC LGSKLKGESEVYLVT IGPANIFPVFNVVQN 2100
 AKLILSRKKNFIMPK KADKESIDANTIKSLI PFLCYPITKKGINTA LSKLKSVVSGDILSY SIAGRNEVFENKLIN 2165
 HKHNNILKWFNHLVNL FRSTELNYNHLYMVE STYPYLSLUNSLITT NELKLIKITGSLLY NFHNE

Charged Clusters (Amino Acids that are underlined were changed to alanines)

Mutations in cpts-248/404

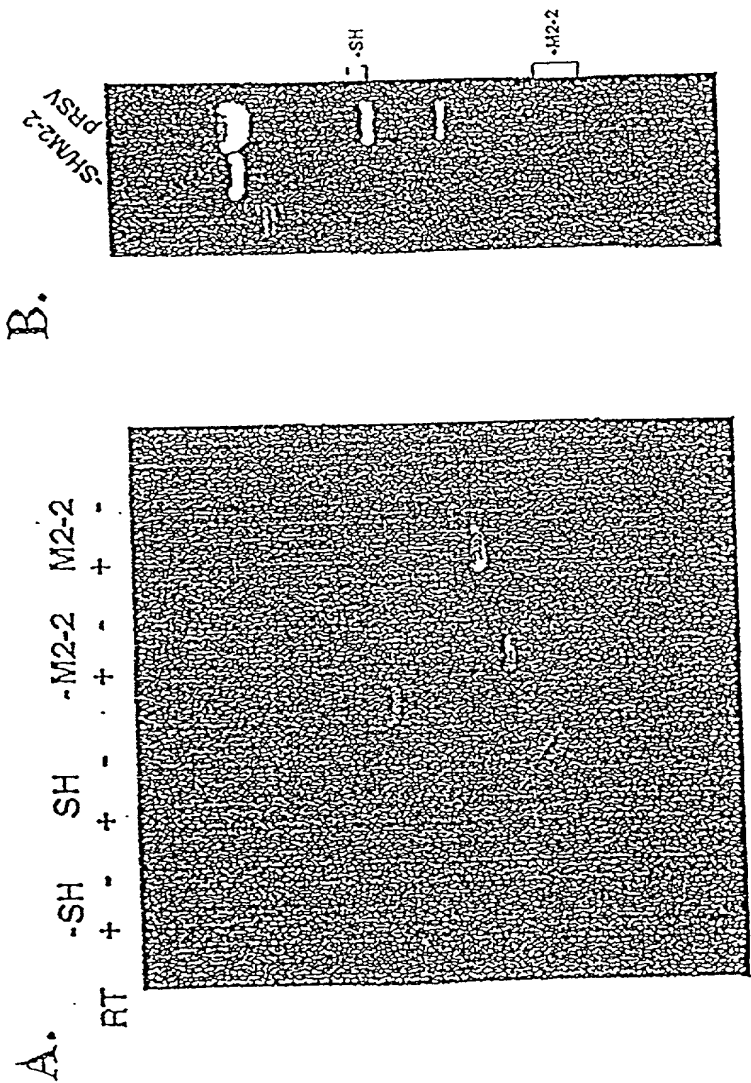
Mutation in cpts530

FIG. 10

MDPIINGNSANVYL	DSYLKGVLSFSECNA	LGSYIFNGPYLKNDY	TNLSRQNPFLIEHNM	LKKLNITQSLISKYH	75
KGEIKLEETTFQSL	IMTYKSMTSSEQIAT	TNLLKTIIRRAIETS	DVKVYAIINKLGLKE	KDKIKSNNGQDEDNS	150
VYTTIKDDILSAVK	DNQSHLKADKNHSTK	QKDTIKTKTLKKLMC	SMQHPSPSWLIHWENL	YTKLNNTLTQYRSNE	225
VKNHGFTLIDNQTL	GFQFILNQYGCIVYH	KELKRIITVTTYNQFL	TWKDISLSRLNVCLI	TWISNCINTLNKSLG	300
LRCGFNNVILLTQLFL	YGDCLLKLFEHNEGFI	ITKEVEGFIMSLILN	ITEEDQFRKRFYNM	LNNITDAANKAQKNL	375
LSRVCHILLDKTVSD	NIINGRWILLLSKFL	KLTKLAGDNNINLS	ELYFLFRIFGHPMVD	ERQAMDVAKINQNET	450
KFYLLSSLSMLRGAF	IYRIITKGFVANNYRW	PTLRNAIVLPLRWLT	YXKLNTPSLLLELTE	RDLIVLSGLRFYREF	525
RLPKKVDLEMTINDK	AISPPKQLIWTSEPR	NYPMSHIQNYIEHEK	LKFSSEDKSRRVLEY	YLRDNKFNEDDLYNC	600
VVNQSYLNNFNHVS	LTGKERELSVGRMFA	MQPGMERQVQIIAEK	MIAENILQFFPESLT	RYGDLELQKILELKA	675
GISNKSNRANDNYAN	YISKCSLITDLSKEN	QAFRYETSCICSDVL	DELHGVQSLFSWLHL	TIPHVTIICTYRHAP	750
PYIGDHTVDLNNVDE	QSGLYRYHMGGIEGW	QOKLWTIEAISLLDL	ISLKGKFSITALING	DNQSIDISKPIRLME	825
GQTHAQADYLLALNS	LKLLYKEYZAGIGHKL	KGTEYISRDMQFMS	KTIQHNQVYYPASIK	KVLRVGFWINTILDD	900
FKVSLSESIGSLTQEL	EYRGESLLQSLIFRN	VMLXNQIALQLKNHA	LCNNKLYLDILKVLK	HLKTFEFLDNIDTAL	975
TLXANLPMFLFGGGDP	NLLXRSFVRRTPDFL	TEATVHSVFIISYYT	NHDLKDKLQDLSDDR	LNKFLTCIITFDKNP	1050
NABFVILMRDPQALG	SERQAKITSEINRLA	VTEVLSTAPNKIFSK	SAQHYTTTELDIANDI	MQNIETPTYPHGLRW	1125
YESLPFYKAEKIVNL	ISGTKSITNILEKTS	ANDLTDIDRATENMR	KNITLLIRILPLDQN	RDKREILSMENLSIT	1200
ELSKYVRERSWSLSN	IVGVTSPSIMYTMDI	KYTTSTISSGIITIEK	YNVNSLTRGERGPTK	PWVGSSTQEKKTMPV	1275
YNRQVLTCKQRDQID	LLAKLDWVYASIDNK	DEFMEELSIGTLGLT	YEKAKKLFPQYLSVN	YLHRLTVSSRPQEF	1350
ASIPAYRTNTYHFDI	SPINRILTEKYGDED	IDIVFQNCISFGLSL	MSVVEQFTNVCPNRI	LLIPKLNETHLMKPP	1425
IFTGDVDIHLKQVT	QKQEMFLPKISLTQ	YVELEFLSNKTLKSGS	HVNSNLILAHKISDY	FHNTYILSTNLAGHW	1500
ILLIIQIMKDSKGIFE	KDWGEGYITDHFMIN	LKQFFENAYKTYLLCF	HKGYGKAKLECDMNT	SDLLCVLELIDSSYW	1575
KMSKVFLQKQVIKY	ILSQDASLHRVKGQH	SFKLWFLKRLNVAEF	TVCPWVNVNIDYHPTH	MKAILTYIDLVRMGL	1650
INIDRIHIKNKHKN	DEFTSNLFXINYNF	SDNTHLLTKHIRIAN	SELENNYNKLYHPTP	ETLENILANPIKSND	1725
KKTIANDYCIGKNVDS	IMLPLLSNKKLIKSS	AMIRTNYSKQDLXNL	FPMVVIDRIIDHSGN	TAKSNQLYTTTSHQI	1800
SLVHNSTSLYCMPLPW	HHINRFNFVFSSTGC	KISIEYILKDLKIKD	PNCIAFIGEGAGNLL	LRTVVELHPDIRYIY	1875
RSLLKDCNDHSLPIEF	LRLVNGHINIDYGEN	LTIPATDATNNIHS	YLHIKFAEPISLFCV	DAELSVTNVNSKIII	1950
EWSKHVRKCKYCSSV	NKCMILIVKYHAQDDI	DFKLDNITILKTYVC	LGSKLKGSEVYLVLT	IGPANIFPVFNVVQN	2025
AKLILSRKXNFIMPX	KADKESIDANIKSLI	PFLCYPTTKKGINTA	LSKLKSVVSGDILSY	SIAGRNEVFNSKLIN	2100
HKHMNILKWFNHVLN	FRSTELNVNHLVAVE	STYPYLSSELNLSLT	NELKKLIKITGSLLY	NFHNE	2165

C Cysteine residues
 C Cysteine residues that were changed to valine or aspartic acid
 C Cysteine residue deleted

FIG. 11



FIGS. 12A-B

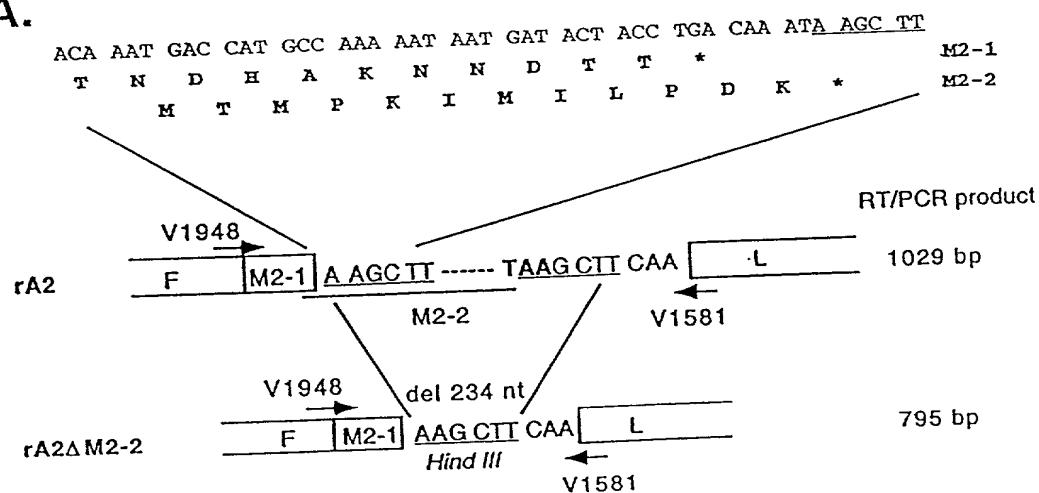
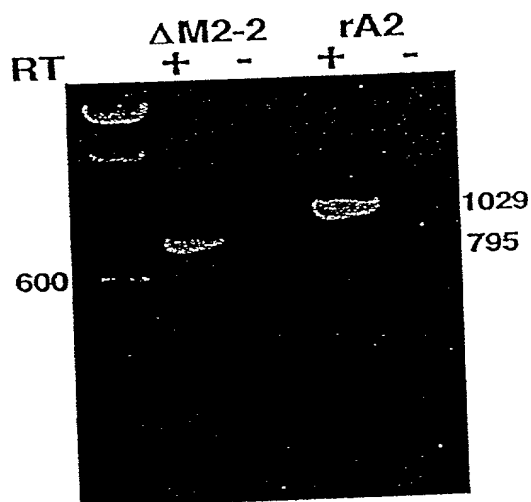
A.**B.**

FIG. 13

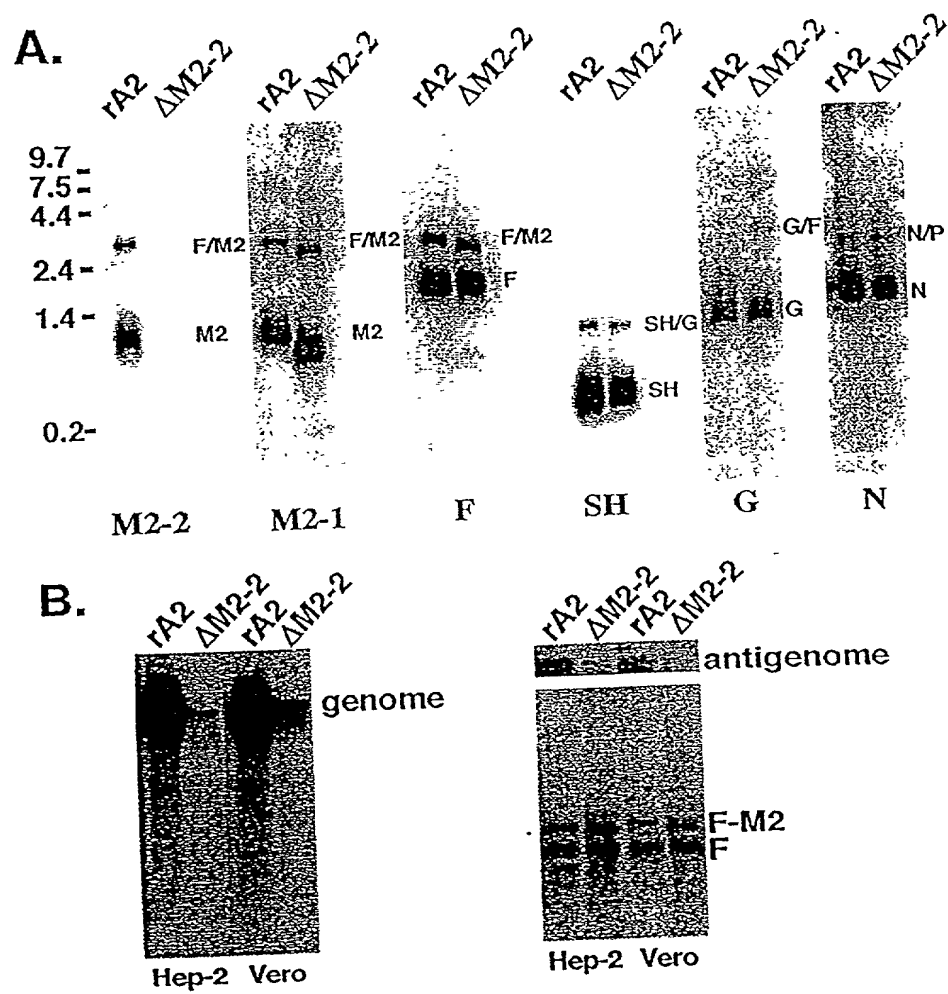


FIG. 14

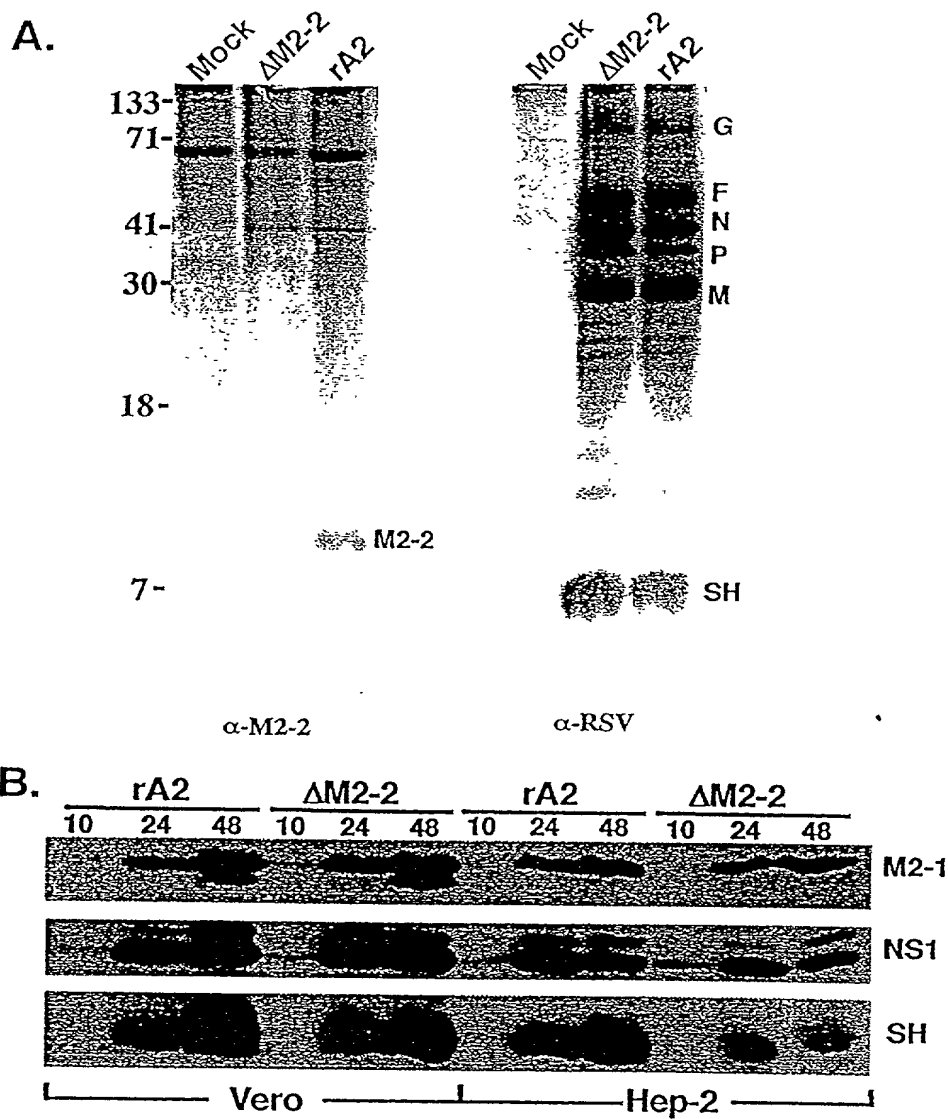


FIG. 15

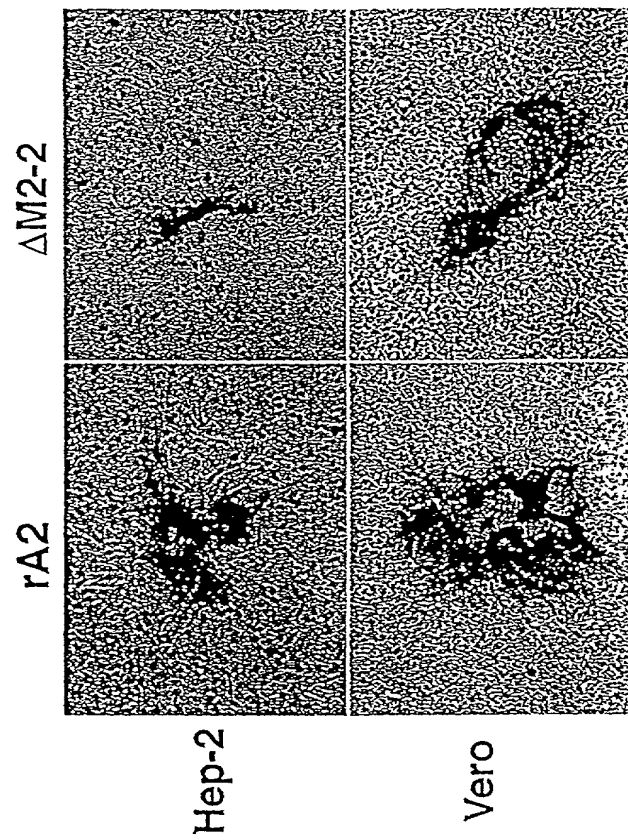


FIG. 16

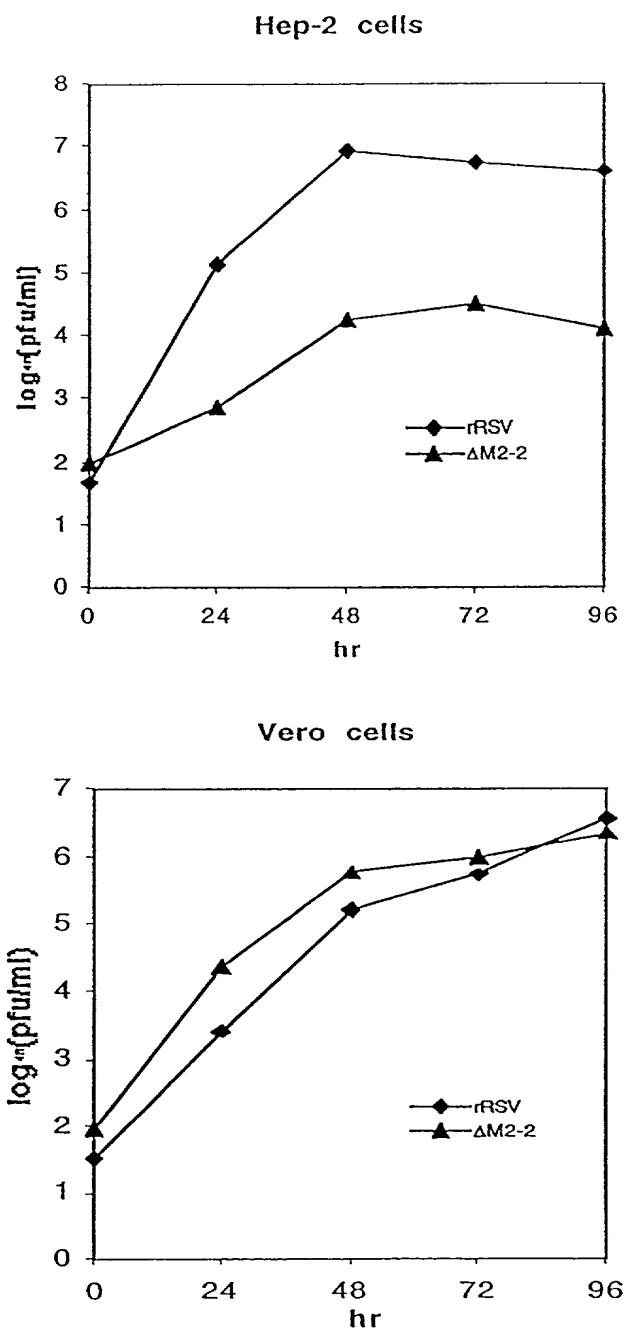


FIG. 17

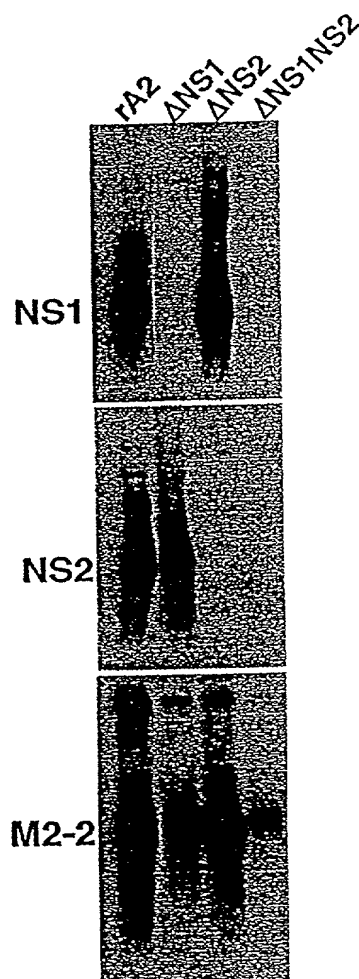


FIG. 18

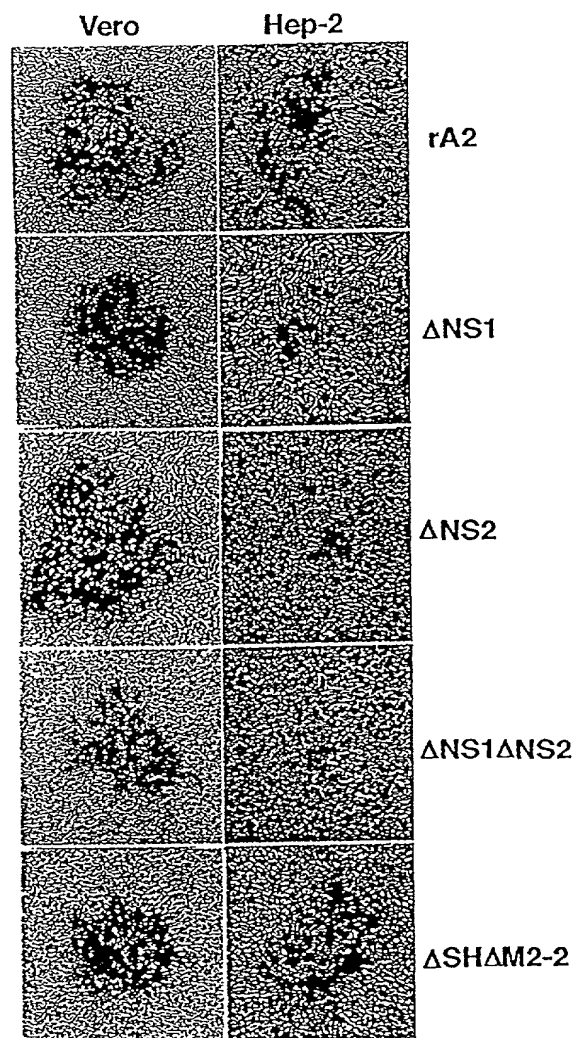


FIG. 19

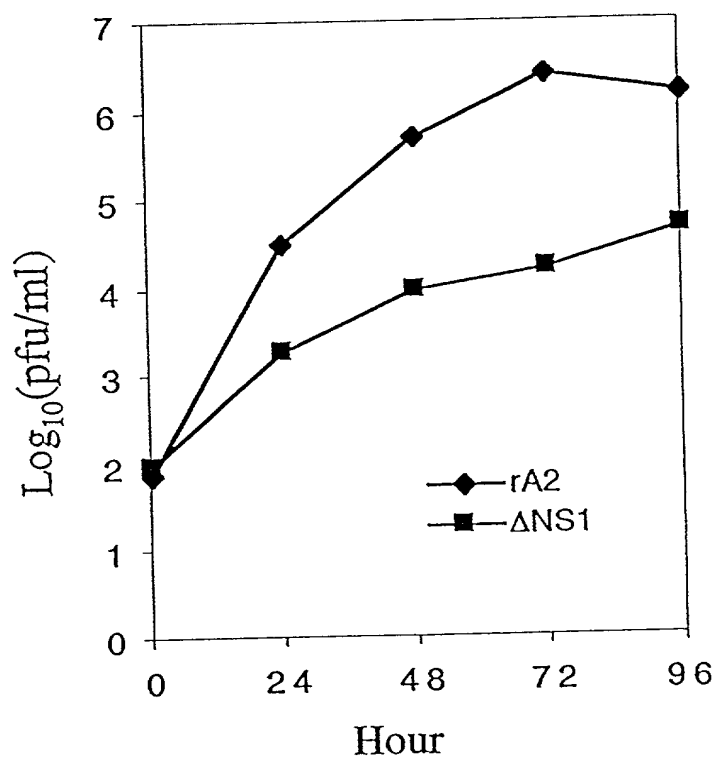


FIG. 20

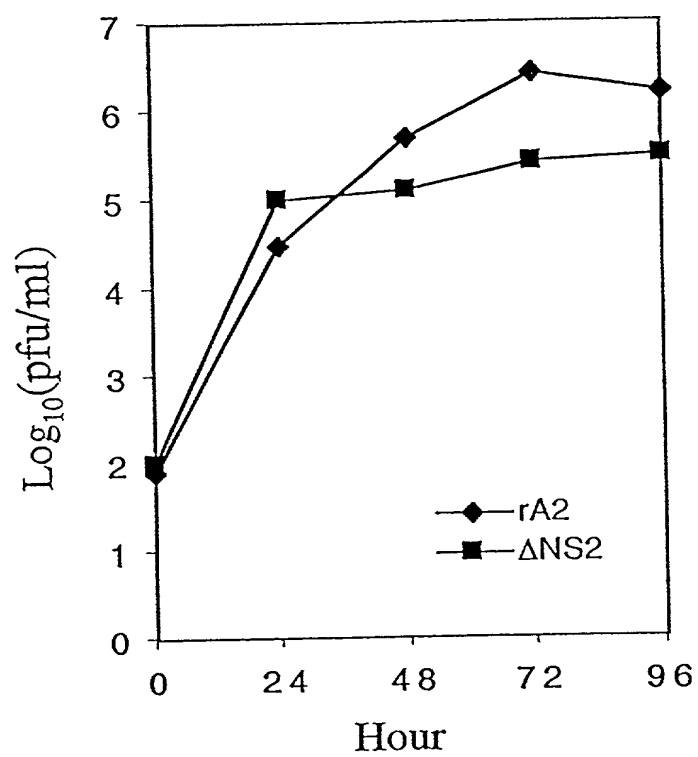


FIG. 21

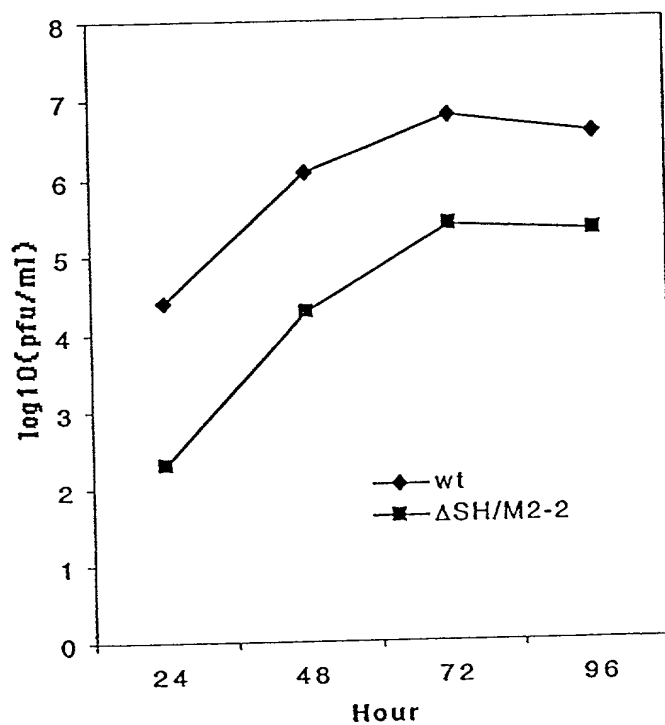


FIG. 22

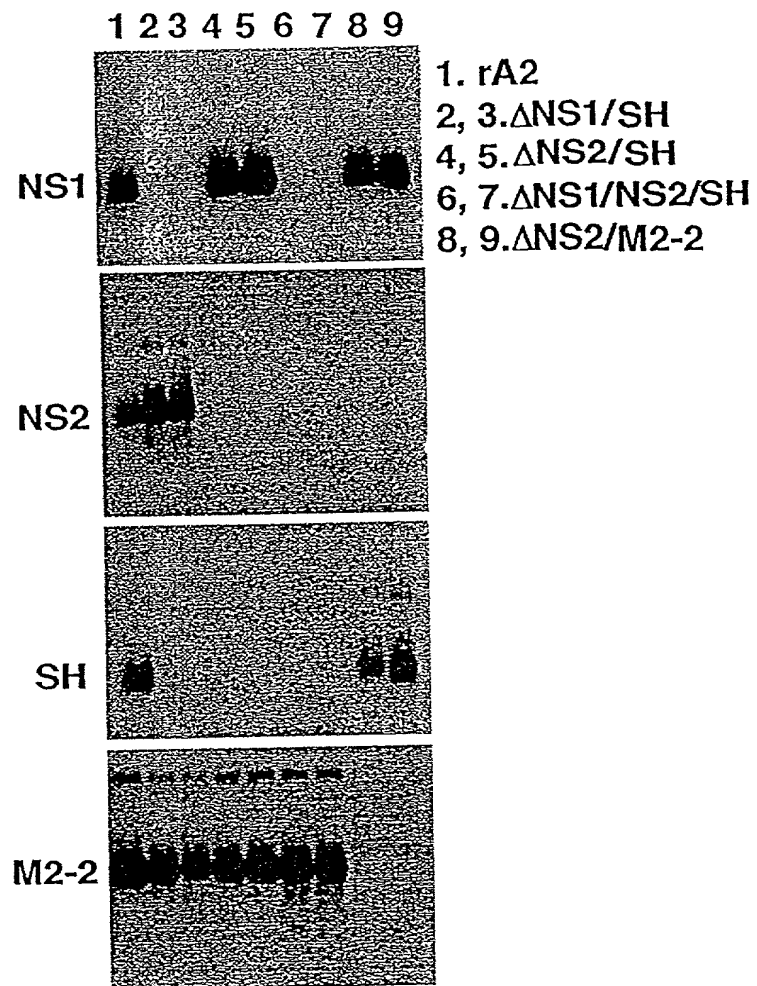


FIG. 23

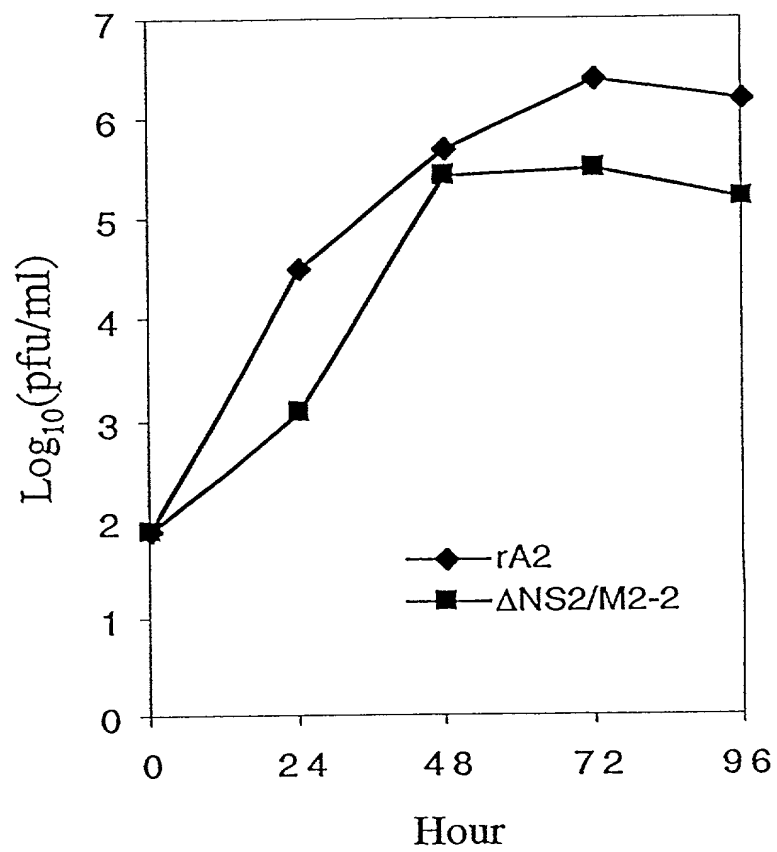


FIG. 24

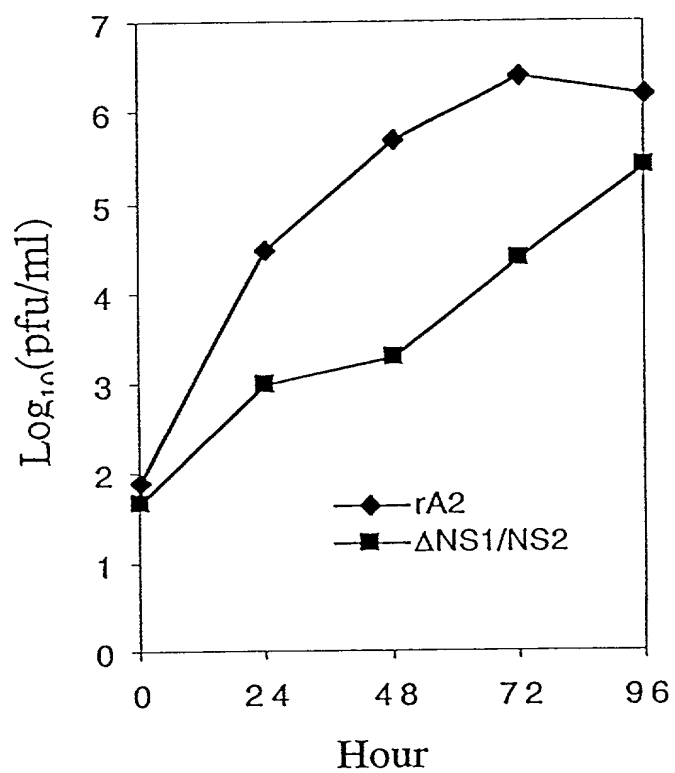


FIG. 25

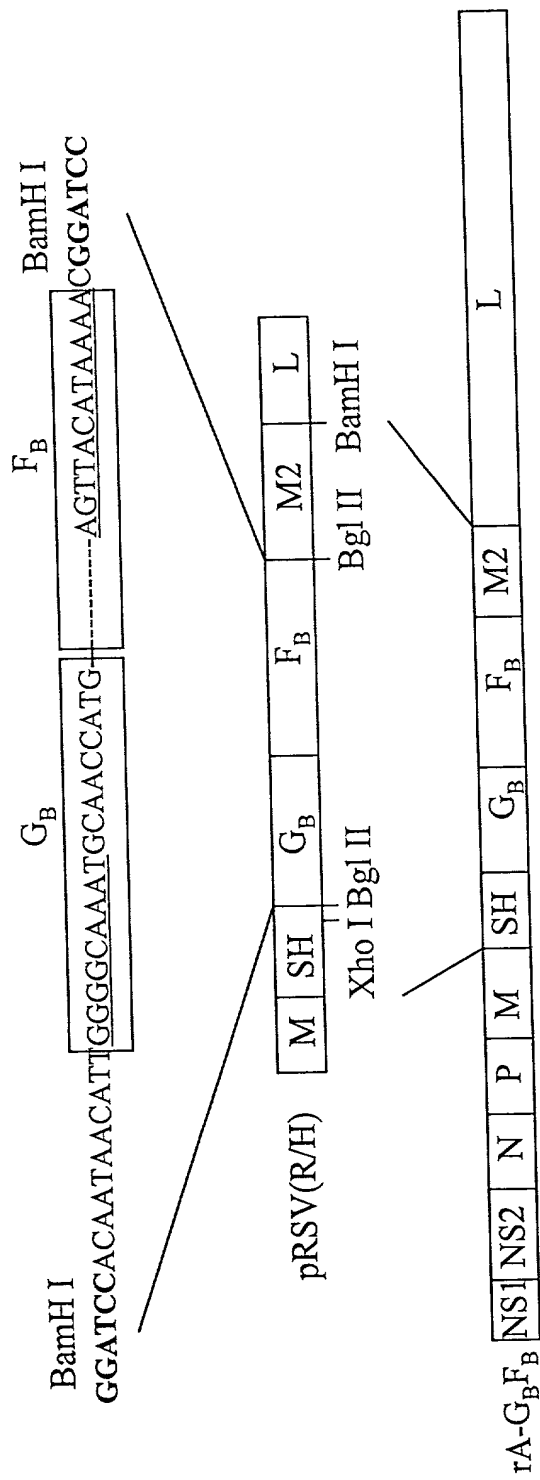
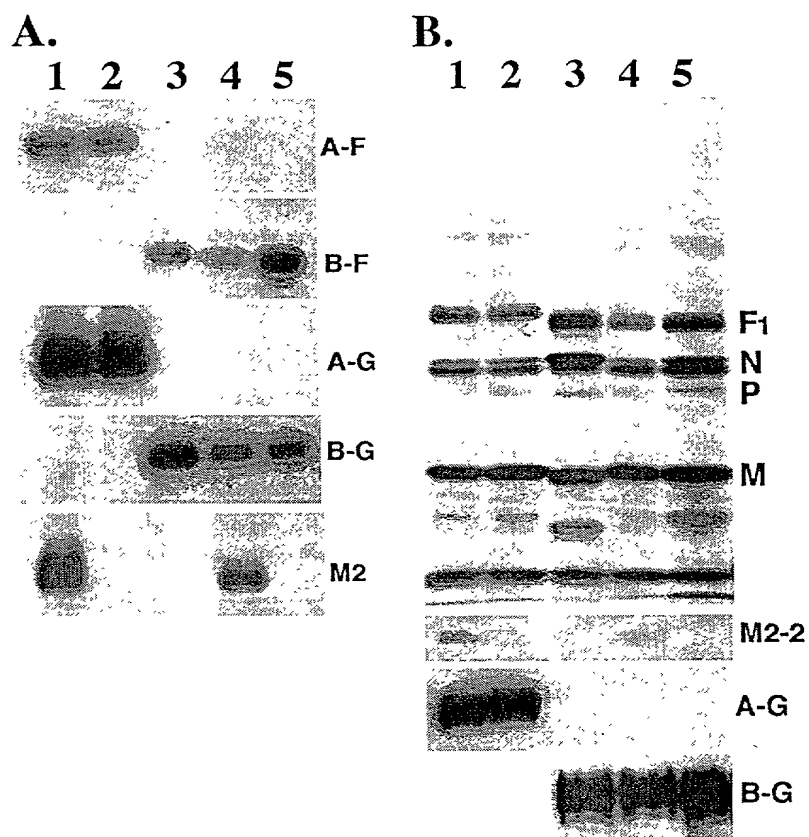


FIG. 26

**FIG. 27**

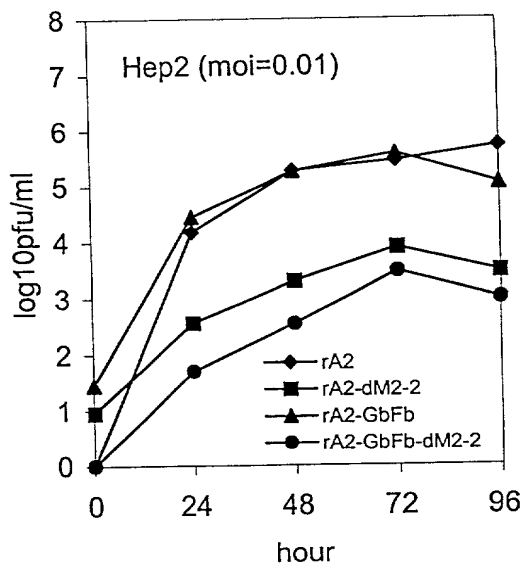
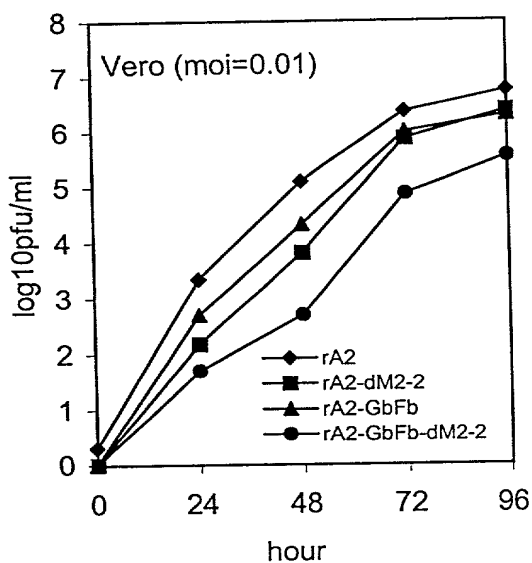
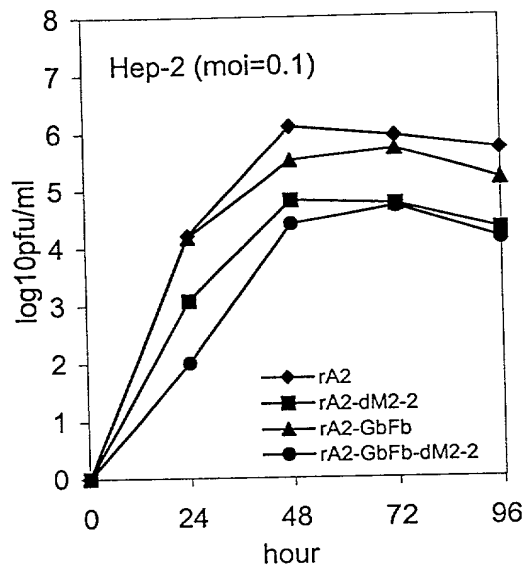
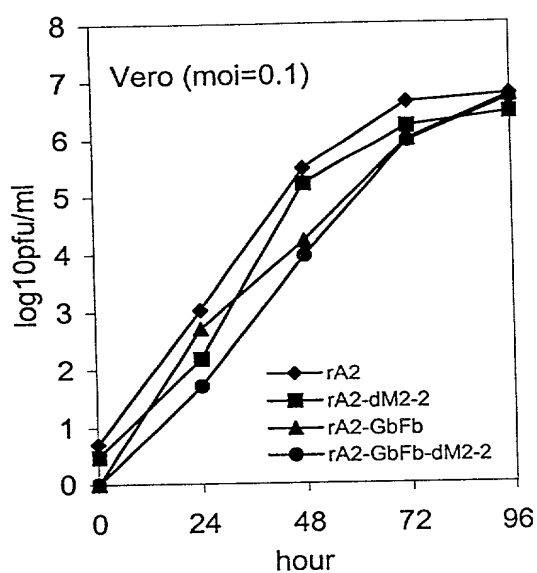


FIG. 28

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below at 201 et seq. underneath my name.

I believe I am the original, first and sole inventor if only one name is listed at 201 below, or an original, first and joint inventor if plural names are listed at 201 et seq. below, of the subject matter which is claimed and for which a patent is sought on the invention entitled

RECOMBINANT RSV VIRUS EXPRESSION SYSTEMS AND VACCINES

and for which a patent application:

- ☐ is attached hereto and includes amendment(s) filed on *(if applicable)*
☒ was filed in the United States on November 28, 2000 as Application No. *(for declaration not accompanying application)*
 with amendment(s) filed on *(if applicable)*
☐ was filed as PCT international Application No. on and was amended under PCT Article 19 on *(if applicable)*

I hereby state that I have reviewed and understand the contents of the above identified application, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119(a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

EARLIEST FOREIGN APPLICATION(S), IF ANY, FILED PRIOR TO THE FILING DATE OF THE APPLICATION			
APPLICATION NUMBER	COUNTRY	DATE OF FILING (day, month, year)	PRIORITY CLAIMED
			YES <input type="checkbox"/> NO <input type="checkbox"/>
			YES <input type="checkbox"/> NO <input type="checkbox"/>

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

APPLICATION NUMBER	FILING DATE
60/060,153	September 26, 1997
60/084,133	May 1, 1998
60/089,207	June 12, 1998

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

APPLICATION SERIAL NO.	FILING DATE	STATUS		
		PATENTED	PENDING	ABANDONED
09/161,122	September 25, 1998		X	
09/368,076	August 3, 1999		X	

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SEND CORRESPONDENCE TO:		PENNIE & EDMONDS LLP 1667 K Street, N.W. Washington, D.C. 20006		DIRECT TELEPHONE CALLS TO: PENNIE & EDMONDS LLP DOCKETING (202) 496-4721	
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	POST OFFICE ADDRESS	STREET 354 Connemara Way	CITY Sunnyvale	STATE OR COUNTRY California	ZIP CODE 94087
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	RESIDENCE & CITIZENSHIP	CITY Cupertino	STATE OR FOREIGN COUNTRY California	COUNTRY OF CITIZENSHIP People's Republic of China	
	POST OFFICE ADDRESS	STREET 21001 Seven Springs Parkway	CITY Cupertino	STATE OR COUNTRY California	ZIP CODE 94043
204	FULL NAME OF INVENTOR	LAST NAME Bryant	FIRST NAME Martin	MIDDLE NAME	
	RESIDENCE & CITIZENSHIP	CITY Los Altos	STATE OR FOREIGN COUNTRY California	COUNTRY OF CITIZENSHIP U.S.A.	
	POST OFFICE ADDRESS	STREET 1664 Clay Drive	CITY Los Altos	STATE OR COUNTRY California	ZIP CODE 94024

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

SIGNATURE OF HONG JIN	SIGNATURE OF RODERICK TANG	SIGNATURE OF SHENGQIANG LI
DATE	DATE	DATE
SIGNATURE OF MARTIN BRYANT		
DATE		